

1 **Supplementary document**

2

3 **Laboratory Measurement**

4

5 *Collection and processing of biospecimens*

6 For Cohort one, the 1 cm x 0.5 cm fresh cancerous liver tissue was be collected at the time of

7 core-needle biopsy or surgery. For Cohort two, before patients received systemic chemotherapy,

8 participants had been directed to collect their feces using a fecal collection kit for gut

9 microbiome study and targeted metabolomics of fecal short-chain fatty acids (SCFAs).

10 Laboratory process and testing was described in supplementary document. Ten milliliters of

11 whole blood were collected to investigate basic biochemical profile, oxidative stress,

12 inflammatory markers, bile acids, and serum ferritin. Quantification of serum ferritin and the

13 basic biochemical profile were conducted using the Cobas e801 immunoassay analyzer (Roche

14 Diagnosis GmbH, Mannheim, Germany). Plasma was prepared by centrifuging whole blood at

15 3000 rpm for ten minutes. All biospecimens were then preserved at -80°C.

16

17 ***Gut microbiota***

18 Bacterial genomic DNA was extracted from 0.25 g of fecal samples using the QIAamp

19 PowerFecal Pro DNA Kit (Qiagen, Hilden, Germany) according to the manufacturer's

20 instructions. The V3–V4 hypervariable regions of the 16S rRNA gene were amplified using the

21 341F and 805R primers and sequenced using paired-end reads on the Illumina NovaSeq 6000

22 platform (Novogene, Singapore) (1). Raw sequencing reads were processed using Quantitative

23 Insights into Microbial Ecology 2 (QIIME 2, v.2023.5). Sequence denoising, quality filtering,

24 chimera removal, and amplicon sequence variant (ASV) generation were performed using the

25 DADA2 plugin (2-4). Taxonomic classification of ASVs was conducted using a pretrained

26 classifier against the SILVA database (version 138) (5). Samples were rarefied to a minimum  
27 sequencing depth of 77,443 reads per sample to retain all samples for downstream diversity  
28 analyses. Alpha- and beta-diversity analyses were subsequently performed using QIIME2. To  
29 reduce potential noise from extremely rare taxa, taxa present in less than 10% of samples were  
30 excluded prior to differential abundance analysis. For differential abundance analysis, QIIME2  
31 outputs were imported into phyloseq environment in R. Taxonomic features were aggregated at  
32 multiple taxonomic levels (phylum, class, order, family, and genus). Differentially abundant  
33 taxa between groups were identified using Analysis of Compositions of Microbiomes with Bias  
34 Correction (ANCOM-BC) (6). The statistical model included the group variable as the main  
35 effect. Multiple-testing correction was applied using the Bonferreni method, and taxa with  
36 adjusted p-values  $< 0.05$  were considered statistically significant. Visualization of differential  
37 abundance results was performed using custom R scripts based on log fold changes estimated by  
38 ANCOM-BC.

39 Functional pathway prediction was performed using PICRUSt2 with the MetaCyc database as  
40 the reference pathway database. Predicted MetaCyc pathway abundance tables were analyzed in  
41 R using ggpicrust2. To reduce noise from sparse features, pathways with nonzero abundance in  
42 fewer than three samples were removed prior to differential abundance analysis. Differentially  
43 abundant pathways between groups were identified using Linear Models for Differential  
44 Abundance (LinDA), with the group variable specified as the main effect and a reference group.  
45 Pathway annotations were assigned using the MetaCyc database, and significant pathways were  
46 exported for Venn diagram analysis (7, 8). Associations between gut microbial taxa and  
47 continuous clinical or metabolite variables were evaluated using ANCOM-BC in R. QIIME2-  
48 derived feature tables were imported into the phyloseq environment and aggregated at the

49 phylum, class, order, family, and genus levels. For each clinical or metabolite variable, an  
50 independent ANCOM-BC model was fitted with the variable treated as a continuous predictor.  
51 ANCOM-BC estimates taxon-specific log-fold changes using a bias-corrected generalized linear  
52 model framework that accounts for the compositional nature of microbiome data. Multiple  
53 hypothesis testing was controlled using Bonferroni correction, and taxa with adjusted p-values <  
54 0.05 were considered statistically significant. Estimated log-fold changes and corresponding  
55 standard errors were extracted for visualization and interpretation.

56

### 57 ***Tissue bacterial quantification***

58 Genomic DNA was extracted from cancerous liver tissue using the DNeasy Blood and Tissue  
59 Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. Extracted DNA  
60 was normalized to a concentration of 50 ng/uL prior to downstream analyses. Representative  
61 bacterial taxa were quantified using quantitative real-time PCR (RT-qPCR), including Eubacteria  
62 (total bacterial load), Firmicutes (estimated from the combined abundance of Clostridiales and  
63 *Lactobacillus acidophilus*), Bacteroidota, and Enterobacteriaceae, following previously  
64 established protocols. RT-qPCR assays were performed in accordance with the Minimum  
65 Information for Publication of Quantitative Real-Time PCR Experiments (MIQE) guidelines (9).  
66 Standard curves were generated using serial dilutions of plasmids containing the target gene  
67 sequences from reference bacterial strains, and the standards were included in the same reaction  
68 plate as the study samples. No-template controls (NTC) were included in each run to monitor  
69 potential contamination during PCR amplification. Differences in bacterial composition between  
70 intrahepatic cholangiocarcinoma (ICCA) and hepatocellular carcinoma (HCC) tissues were

71 evaluated based on the abundance of these taxa together with the Firmicutes/Bacteroidota (F/B)  
72 ratio (10).

73

#### 74 ***Determination of Opisthorchis viverini infection***

75 Serum sample in 50  $\mu$ L was used to diagnose OV infection by immunochromatographic point-  
76 of-care testing kit (11).

77

#### 78 ***Oxidative stress measurement in peripheral blood mononuclear cells***

79 Three milliliters of blood were used for peripheral blood mononuclear cells (PBMCs) isolation.

80 Reactive oxygen species is a marker of oxidative stress, to determine ROS production in

81 PBMCs, PBMCs were incubated with 2  $\mu$ M Dichlorohydro-Fluorescein Diacetate dye (DCFH-

82 DA), at 25°C for 20 minutes. The DCFH-DA diffused through the mitochondrial cell membrane,

83 and it was deacetylated by intracellular esterases to a non-fluorescent compound, which was later

84 oxidized by ROS into a dichlorofluorescein (DCF) form. DCF was a highly fluorescent

85 compound, which can be detected by fluorescence spectroscopy with maximum excitation. An

86 increased level of DCF indicates an increased ROS production. Fluorescent intensity of the DCF

87 was measured with an excitation wavelength at 485 nm, and an emission wavelength at 530 nm

88 using a flow cytometer (FACS Celesta, BD biosciences, San Jose, CA, USA) (12, 13).

89

#### 90 ***Inflammatory cytokine expression***

91 The expression of tumor necrosis factor (TNF)- $\alpha$ , transforming growth factor (TGF)- $\beta$ ,

92 interleukin (IL)-1 $\beta$ , and IL-6 genes were measured by SYBR Green-based Quantitative Reverse

93 Transcription Polymerase Chain Reaction (RT-qPCR) of mRNA extracted from the buffy coated

94 blood. The thermal cycling conditions were used denaturation at 95°C for 20 s, alignment at  
95 54°C for 20 s, and elongation at 72°C for 20 s, for 40 cycles. A second pair of beta-actin primers  
96 were used as an internal control: forward, 5'-CCAGATCATGTTTGAGACC-3' and reverse, 5'-  
97 ATGTCACGCACGATTTCCC-3'. All reactions were performed in duplicate. Reaction  
98 mixtures, without RNA, were used as negative controls in each run (13).

99

### 100 ***Bile acid analysis***

101 Bile acid concentrations were quantitated using a 1260 infinity II liquid chromatography/6546  
102 quadrupole time-of-flight mass spectrometry (Agilent technologies, Santa Clara, CA, USA)  
103 under reversed phase liquid chromatography negative ion mode. Mobile phase A was 0.1%  
104 formic acid in water and mobile phase B was 0.1% formic acid in acetonitrile. The column  
105 setting and gradient of mobile phases were described in a previous study (14). The mass  
106 spectrometry setting was as follows: dual Agilent jet stream electrospray ionization, full-scan  
107 mass spectrometry detection (m/z 50 to 1,200), acquisition rate 1 spectrum/sec, capillary voltage  
108 3,500 V, nozzle voltage 1,500 V, gas temperature 350 °C, drying gas 10 L/min, nebulizer  
109 pressure 20 psig, and reference mass correction was enabled (15).

110

### 111 ***Targeted metabolomics of SCFAs***

112 Gas chromatography-mass spectrometry has been primarily designated for use in targeted  
113 metabolomics, specifically for SCFAs (16). To quantify the targeted metabolomics of SCFAs in  
114 both fecal content and plasma, gas chromatography-mass spectrometry (Agilent Technologies,  
115 California, USA) was employed, as described in our previous study (4). The actual concentration  
116 of SCFAs was calculated by comparison with a standard mixture (Restek, Pennsylvania, USA)

117 using MassHunter Quantitative Analysis Software v.10.1 (Agilent Technologies, California,  
118 USA) (14).

119  
120 **References**

- 121  
122 1. Sriwichain S, Kittichotirat W, Chunchai T, Chattipakorn N, Chattipakorn SC. Profiles of  
123 gut microbiota in obese-insulin-resistant rats treated with biotics. *Eur J Nutr.* 2022.
- 124 2. Weiss S, Xu ZZ, Peddada S, Amir A, Bittinger K, Gonzalez A, et al. Normalization and  
125 microbial differential abundance strategies depend upon data characteristics. *Microbiome.*  
126 2017;5(1):1-18.
- 127 3. Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, et al.  
128 Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. *Nat*  
129 *Biotechnol.* 2019;37(8):852-7.
- 130 4. Sriwichain S, Thiennimitr P, Thonusin C, Sarichai P, Buddhasiri S, Kumfu S, et al.  
131 Deferiprone has less benefits on gut microbiota and metabolites in high iron-diet induced iron  
132 overload thalassemic mice than in iron overload wild-type mice: A preclinical study. *Life Sci.*  
133 2022;307:120871.
- 134 5. Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, et al. The SILVA  
135 ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic*  
136 *Acids Research.* 2012;41(D1):D590-D6.
- 137 6. Lin H, Peddada SD. Analysis of compositions of microbiomes with bias correction.  
138 *Nature communications.* 2020;11(1):1-11.
- 139 7. Yang C, Mai J, Cao X, Burberry A, Cominelli F, Zhang L. ggpicrust2: an R package for  
140 PICRUST2 predicted functional profile analysis and visualization. *Bioinformatics.* 2023;39(8).

- 141 8. Douglas GM, Maffei VJ, Zaneveld JR, Yurgel SN, Brown JR, Taylor CM, et al.  
142 PICRUSt2 for prediction of metagenome functions. *Nat Biotechnol.* 2020;38(6):685-8.
- 143 9. Winter SE, Thiennimitr P, Winter MG, Butler BP, Huseby DL, Crawford RW, et al. Gut  
144 inflammation provides a respiratory electron acceptor for Salmonella. *Nature.*  
145 2010;467(7314):426-9.
- 146 10. Saiyasit N, Chunchai T, Prus D, Suparan K, Pittayapong P, Apaijai N, et al. Gut dysbiosis  
147 develops before metabolic disturbance and cognitive decline in high-fat diet-induced obese  
148 condition. *Nutrition.* 2020;69:110576.
- 149 11. Sadaow L, Sanpool O, Rodpai R, Yamasaki H, Ittiprasert W, Mann VH, et al.  
150 Development of an Immunochromatographic Point-of-Care Test for Serodiagnosis of  
151 Opisthorchiasis and Clonorchiasis. *Am J Trop Med Hyg.* 2019;101(5):1156-60.
- 152 12. Sawaddiruk P, Apaijai N, Paiboonworachat S, Kaewchur T, Kasitanon N, Jaiwongkam T,  
153 et al. Coenzyme Q10 supplementation alleviates pain in pregabalin-treated fibromyalgia patients  
154 via reducing brain activity and mitochondrial dysfunction. *Free Radic Res.* 2019;53(8):901-9.
- 155 13. Khuankaew C, Apaijai N, Sawaddiruk P, Jaiwongkam T, Kerdphoo S, Pongsiriwet S, et  
156 al. Effect of coenzyme Q10 on mitochondrial respiratory proteins in trigeminal neuralgia. *Free*  
157 *Radic Res.* 2018;52(4):415-25.
- 158 14. Thonusin C, IglayReger HB, Soni T, Rothberg AE, Burant CF, Evans CR. Evaluation of  
159 intensity drift correction strategies using MetaboDrift, a normalization tool for multi-batch  
160 metabolomics data. *J Chromatogr A.* 2017;1523:265-74.
- 161 15. Thonusin C, Nawara W, Khuanjing T, Prathumsup N, Arinno A, Ongnok B, et al. Blood  
162 metabolomes as non-invasive biomarkers and targets of metabolic interventions for doxorubicin  
163 and trastuzumab-induced cardiotoxicity. *Arch Toxicol.* 2023;97(2):603-18.

164 16. Hoving LR, Heijink M, van Harmelen V, van Dijk KW, Giera M. GC-MS Analysis of  
165 Short-Chain Fatty Acids in Feces, Cecum Content, and Blood Samples. *Methods Mol Biol.*  
166 2018;1730:247-56.  
167

168 **Supplementary Table 1.** Baseline demographic and clinical characteristics.

Characteristics	CCA (n=56)	HCC (n=14)	p-value
<b>Demographics</b>			
Age, year	64.0 (60.0,68.3)	60.5 (52.0,68.0)	0.28
Sex, n (%)			<b>0.04</b>
Male	33 (58.9)	12 (85.7)	
Female	23 (41.1)	2 (14.3)	
BW, kg (mean, SD)	55.4 (44.6 - 66.2)	63.9 (51.9 – 75.9)	<b>0.01</b>
Risk factor			
Smoking (%)	9 (16.1)	0	0.19
Alcohol consumption (%)	16 (28.6)	3 (27.4)	1.00
Raw food consumption (%)	11 (19.6)	0.0	0.11
NAFLD (%)	0	1 (7.1)	0.19
Cirrhosis (%)	8 (14.3)	10 (71.4)	<b>&lt;0.01</b>
HBV infection (%)	2 (3.6)	7 (50.0)	<b>&lt;0.01</b>
HCV infection (%)	1 (1.8)	6 (42.9)	<b>&lt;0.01</b>
Antibiotics use (%)	2 (3.6)	0	1
Treatment (%)			<b>&lt;0.01</b>
Surgery	5 (8.9)	8 (57.1)	
Chemotherapy	48 (85.7)	2 (14.3)	
Multi-kinase inhibitor	0	4 (28.6)	
Supportive care	6 (10.7)	0	
<b>Tumor staging</b>			
T (%)			0.31
1-2	39 (69.6)	8 (57.1)	
3-4	17 (30.4)	6 (42.9)	
N (%)			<b>&lt;0.01</b>
0	9 (16.1)	13 (92.9)	
1	47 (83.9)	1 (7.1)	
M (%)			<b>&lt;0.01</b>
0	10 (17.9)	12 (85.7)	
1	46 (82.1)	2 (14.3)	
Metastatic site (%)			<b>&lt;0.01</b>
0-1	36 (64.3)	14 (100.0)	
≥ 2	20 (33.9)	0	

169 **Abbreviations:** BW, body weight; CCA, cholangiocarcinoma; HBV, hepatitis B virus; HCC,  
170 hepatocellular carcinoma; HCV, hepatitis C virus; kg, kilogram; NAFLD, non-alcoholic fatty  
171 liver diseases

172 **Supplementary Table 2.** Tissue bacterial profiles using RT-qPCR comparing between ICCA and HCC patients.

Bacterial profiles	ICCA	HCC	Unadjusted mean difference (95 % CI)	p-value	Adjusted mean difference* (95 % CI)	p-value
	Mean (range) n=56	Mean (range) n=14				
Eubacteria (copies/uL, total read count)	102190.5 (0.01–1211055.9)	243694.1 (0.7–1434258.1)	141503.6 (-18383.6 - 301390.8)	0.08	199930.2 (-79723.5 - 479583.8)	0.16
Clostridiales, (%)	50.6 (0.0–1422.5)	53.6 (1.1–356.3)	2.9 (-104.5 - 110.3)	0.96	20.2 (-168.4 - 208.7)	0.83
<i>Lactobacillus</i> , (%)	6.6 (0.0–111.6)	13.7 (0.0–172.8)	7.1 (-9.9 - 24.0)	0.41	1.9 (-27.2 - 31.0)	0.90
Bacteroidota, (%)	192.2 (0.0–10519.3)	25.1 (0.0–346.8)	-167.1 (-921.1 - 586.9)	0.66	47.1 (-1285.3 - 1379.4)	0.94
<i>Enterobacteriaceae</i> , (%)	9.2 (0.0–159.1)	34.34 (0.0–236.11)	25.16 (-1.68 to 51.99)	0.07	-25.1 (-68.7 - 18.6)	0.26
Clostridiales and <i>Lactobacillus</i> , (%)	58.16 (0.00–1422.45)	67.22 (1.09–529.11)	9.06 (-103.43 to 121.55)	0.87	22.3 (-174.8 - 219.3)	0.82
F/B ratio	327.58 (0.00–4232.03)	155.18 (0.00–1241.98)	-172.40 (-605.02 to 260.23)	0.43		0.07

173

174 **Abbreviations:** F/B, Firmicutes per Bacteroidota ratio; RT-qPCR, reverse transcriptase-quantitative polymerase chain reaction; 16S

175 rRNA, 16S ribosomal ribonucleic acid.

176 **\*Adjusted for cirrhosis status, HBV infection, HCV infection, and metastatic status**

**Supplementary Table 3.** Tissue bacterial profiles by survival and OV infection status in ICCA patients.

Tissue bacterial profiles	> 6m-PFS (n= 16)	≤ 6m-PFS (n=14)	p- value	> 1yr-OS (n=12)	≤ 1yr-OS (n=18)	p- value	Non-OV (n=21)	OV (n=9)	p- value
Eubacteria (copies/uL)	11410.88 (2883.89, 174963.69)	1887.55 (262.84, 19188.13)	0.23	18049.90 (3274.28, 163937.53)	1896.12 (739.16, 24323.47)	0.37	3585.94 (361.17, 136468.32)	3782.11 (1879.00, 24323.48)	0.92
Clostridiales (%)	3.18 (1.15,5.87)	1.52 (0.89,2.95)	0.21	3.91 (2.03,5.11)	1.47 (0.87,3.02)	1.00	2.45 (1.10,6.01)	1.43 (0.68,2.74)	0.15
Lactobacillus (%)	0.00 (0.00,0.07)	0.01 (0.00,0.29)	0.76	0.02 (0.00,0.18)	0.01 (0.00,0.10)	0.71	0.02 (0.00,0.21)	0.01 (0.00,0.01)	0.48
Bacteroidota (%)	0.07 (0.04,0.26)	0.03 (0.00,0.76)	0.63	0.09 (0.06,0.51)	0.04 (0.00,0.21)	0.24	0.09 (0.04,0.51)	0.01 (0.00,0.07)	0.17
<i>Enterobacteriaceae</i> (%)	0.06 (0.04,0.28)	0.19 (0.03,3.02)	0.43	0.06 (0.05,0.35)	0.10 (0.03,3.00)	0.64	0.101 (0.06,0.57)	0.04 (0.03,0.27)	0.20
Clostridiales and <i>Lactobacillus</i> (%)	3.59 (1.15,5.96)	1.80 (0.98,3.11)	0.27	4.56 (2.50,5.22)	1.58 (0.97,3.14)	0.13	2.61 (1.10,7.86)	1.44 (1.01,3.02)	0.24
F/B ratio	49.59 (14.16, 461.14)	94.42 (21.04, 492.84)	0.78	28.83 (14.21, 80.50)	96.31 (18.82, 754.88)	0.40	28.83 (14.21,96.31)	407.47 (70.34, 522.75)	0.31

**Abbreviations:** F/B, Firmicutes per Bacteroidota ratio; OS, overall survival; OV, *Opisthorchis viverini*; PFS, progression-free survival.

**Supplementary tables 4.** Differential abundance of bacterial taxa between two groups

**4.1 Comparison between progression of disease on response evaluation by CT scan and no progression**

Taxon	FC	SD	p_value	beta_direction	p_significance	color_to_fill_p	tax_level
Monoglobales	-2.18	0.72	0.00	Decreased	Sig	down_red	Order
Clostridiales	-1.58	0.79	0.05	Decreased	Sig	down_red	Order
Burkholderiales	0.75	0.37	0.05	Increased	Sig	up_blue	Order
Monoglobaceae	-2.19	0.69	0.00	Decreased	Sig	down_red	Family
Lactobacillaceae	-2.06	1.04	0.05	Decreased	Sig	down_red	Family
[Eubacterium]_coprostanoligenes_group	-1.76	0.76	0.02	Decreased	Sig	down_red	Family
Peptostreptococcaceae	-1.74	0.84	0.04	Decreased	Sig	down_red	Family
Clostridiaceae	-1.59	0.79	0.04	Decreased	Sig	down_red	Family
UCG-010	-1.50	0.60	0.01	Decreased	Sig	down_red	Family
Clostridia_vadinBB60_group	-1.26	0.63	0.04	Decreased	Sig	down_red	Family
Sutterellaceae	0.94	0.40	0.02	Increased	Sig	up_blue	Family
Coriobacteriaceae	1.72	0.76	0.02	Increased	Sig	up_blue	Family
Selenomonadaceae	2.04	1.03	0.05	Increased	Sig	up_blue	Family
Ruminococcus	-2.78	0.80	0.00	Decreased	Sig	down_red	Genus
Monoglobus	-2.15	0.65	0.00	Decreased	Sig	down_red	Genus
Romboutsia	-2.07	0.88	0.02	Decreased	Sig	down_red	Genus
NK4A214_group	-2.07	0.74	0.01	Decreased	Sig	down_red	Genus
Lactobacillus	-2.02	1.02	0.05	Decreased	Sig	down_red	Genus
[Ruminococcus]_gauvreauii_group	-1.82	0.71	0.01	Decreased	Sig	down_red	Genus
UCG-002	-1.78	0.81	0.03	Decreased	Sig	down_red	Genus
Lachnospiraceae_FCS020_group	-1.77	0.67	0.01	Decreased	Sig	down_red	Genus
Lachnospiraceae_ND3007_group	-1.73	0.81	0.03	Decreased	Sig	down_red	Genus
[Eubacterium]_coprostanoligenes_group	-1.72	0.68	0.01	Decreased	Sig	down_red	Genus
Intestinibacter	-1.70	0.78	0.03	Decreased	Sig	down_red	Genus

UCG-005	-1.50	0.74	0.04	Decreased	Sig	down_red	Genus
Christensenellaceae_R-7_group	-1.49	0.72	0.04	Decreased	Sig	down_red	Genus
Lachnospiraceae_UCG-001	-1.49	0.71	0.04	Decreased	Sig	down_red	Genus
Intestinimonas	-1.46	0.45	0.00	Decreased	Sig	down_red	Genus
UCG-010	-1.46	0.55	0.01	Decreased	Sig	down_red	Genus
UCG-003	-1.44	0.63	0.02	Decreased	Sig	down_red	Genus
Negativibacillus	-1.34	0.65	0.04	Decreased	Sig	down_red	Genus
Paeniclostridium	-1.32	0.58	0.02	Decreased	Sig	down_red	Genus
[Eubacterium]_brachy_group	-1.29	0.63	0.04	Decreased	Sig	down_red	Genus
d_Bacteria_Actinobacteriota_Coriobacteriia_Coriobacteriales_Eggerthellaceae_uncultured	-1.26	0.56	0.03	Decreased	Sig	down_red	Genus
Clostridia_vadinBB60_group	-1.22	0.59	0.04	Decreased	Sig	down_red	Genus
UCG-009	-0.98	0.45	0.03	Decreased	Sig	down_red	Genus
Faecalitalea	-0.95	0.45	0.04	Decreased	Sig	down_red	Genus
d_Bacteria_Firmicutes_Bacilli_Erysipelotrichales_Erysipelotrichaceae_uncultured	-0.84	0.42	0.05	Decreased	Sig	down_red	Genus
Merdibacter	-0.78	0.39	0.04	Decreased	Sig	down_red	Genus
Anaerofilum	-0.75	0.37	0.04	Decreased	Sig	down_red	Genus
Sutterella	1.02	0.40	0.01	Increased	Sig	up_blue	Genus
Collinsella	1.76	0.73	0.02	Increased	Sig	up_blue	Genus
Acidaminococcus	1.93	0.95	0.04	Increased	Sig	up_blue	Genus

#### 4.2 Comparison between ≤ 6m-PFS compared to > 6m-PFS group

Taxon	FC	SD	p_value	beta direction	p significance	color to fill p	tax level
Desulfuromonadia	-1.60	0.78	0.04	Decreased	Sig	down_red	Class
Clostridiales	-2.09	0.71	0.00	Decreased	Sig	down_red	Order
Monoglobales	-2.08	0.69	0.00	Decreased	Sig	down_red	Order
Christensenellales	-1.56	0.75	0.04	Decreased	Sig	down_red	Order
Clostridia_vadinBB60_group	-1.40	0.65	0.03	Decreased	Sig	down_red	Order
Burkholderiales	0.85	0.35	0.02	Increased	Sig	up_blue	Order
Clostridiaceae	-2.09	0.71	0.00	Decreased	Sig	down_red	Family
Monoglobaceae	-2.08	0.66	0.00	Decreased	Sig	down_red	Family
[Eubacterium]_coprostanoligenes_group	-1.86	0.69	0.01	Decreased	Sig	down_red	Family
UCG-010	-1.70	0.58	0.00	Decreased	Sig	down_red	Family
Christensenellaceae	-1.56	0.71	0.03	Decreased	Sig	down_red	Family
Bradymonadales	-1.45	0.74	0.05	Decreased	Sig	down_red	Family
Clostridia_vadinBB60_group	-1.40	0.62	0.02	Decreased	Sig	down_red	Family
Sutterellaceae	1.08	0.38	0.00	Increased	Sig	up_blue	Family
Ruminococcus	-2.50	0.74	0.00	Decreased	Sig	down_red	Genus
NK4A214_group	-2.33	0.67	0.00	Decreased	Sig	down_red	Genus
Monoglobus	-2.06	0.62	0.00	Decreased	Sig	down_red	Genus
UCG-002	-2.04	0.75	0.01	Decreased	Sig	down_red	Genus
Lachnospiraceae_FCS020_group	-2.02	0.63	0.00	Decreased	Sig	down_red	Genus
Clostridium_sensu_stricto_1	-1.97	0.67	0.00	Decreased	Sig	down_red	Genus
[Eubacterium]_coprostanoligenes_group	-1.84	0.62	0.00	Decreased	Sig	down_red	Genus
Romboutsia	-1.82	0.86	0.04	Decreased	Sig	down_red	Genus
UCG-005	-1.72	0.72	0.02	Decreased	Sig	down_red	Genus
UCG-010	-1.69	0.53	0.00	Decreased	Sig	down_red	Genus
Lachnospiraceae_ND3007_group	-1.67	0.76	0.03	Decreased	Sig	down_red	Genus
Christensenellaceae_R-7_group	-1.62	0.68	0.02	Decreased	Sig	down_red	Genus
Paeniclostridium	-1.61	0.60	0.01	Decreased	Sig	down_red	Genus

Enterobacter	-1.53	0.74	0.04	Decreased	Sig	down_red	Genus
[Ruminococcus]_gavreauii_group	-1.52	0.69	0.03	Decreased	Sig	down_red	Genus
Intestinimonas	-1.51	0.44	0.00	Decreased	Sig	down_red	Genus
Prevotellaceae_UCG-004	-1.51	0.71	0.03	Decreased	Sig	down_red	Genus
Negativibacillus	-1.49	0.63	0.02	Decreased	Sig	down_red	Genus
d_Bacteria_Actinobacteriota_Coriobacteriia_Coriobacteriales_Eggerthellaceae_uncultured	-1.48	0.54	0.01	Decreased	Sig	down_red	Genus
Bradymonadales	-1.43	0.71	0.04	Decreased	Sig	down_red	Genus
Family_XIII_AD3011_group	-1.43	0.67	0.03	Decreased	Sig	down_red	Genus
Clostridia_vadinBB60_group	-1.38	0.58	0.02	Decreased	Sig	down_red	Genus
UCG-003	-1.34	0.60	0.02	Decreased	Sig	down_red	Genus
[Eubacterium]_xylanophilum_group	-1.30	0.60	0.03	Decreased	Sig	down_red	Genus
[Eubacterium]_brachy_group	-1.23	0.60	0.04	Decreased	Sig	down_red	Genus
UCG-009	-1.12	0.43	0.01	Decreased	Sig	down_red	Genus
Lactococcus	-1.08	0.54	0.04	Decreased	Sig	down_red	Genus
d_Bacteria_Firmicutes_Bacilli_Erysipelotrichales_Erysipelotrichaceae_uncultured	-0.95	0.44	0.03	Decreased	Sig	down_red	Genus
Sutterella	1.05	0.40	0.01	Increased	Sig	up_blue	Genus
Acidaminococcus	1.80	0.89	0.04	Increased	Sig	up_blue	Genus

### 4.3 Comparison between OV-infected patients and non-OV infected patients

Taxon	FC	SD	p value	beta direction	p significance	color to fill p	tax level
Alphaproteobacteria	-1.80	0.56	0.00	Decreased	Sig	down_red	Class
Monoglobales	-2.43	0.77	0.00	Decreased	Sig	down_red	Order
Clostridiales	-2.18	0.84	0.01	Decreased	Sig	down_red	Order
Rhodospirillales	-1.95	0.54	0.00	Decreased	Sig	down_red	Order
Pasteurellales	-1.55	0.56	0.01	Decreased	Sig	down_red	Order
Micrococcales	-1.40	0.64	0.03	Decreased	Sig	down_red	Order
Clostridia_UCG-014	-1.31	0.65	0.04	Decreased	Sig	down_red	Order
Monoglobaceae	-2.41	0.74	0.00	Decreased	Sig	down_red	Family
Clostridiaceae	-2.16	0.84	0.01	Decreased	Sig	down_red	Family
Leuconostocaceae	-1.97	0.65	0.00	Decreased	Sig	down_red	Family
d_Bacteria_Proteobacteria_Alphaproteobacteria_Rhodospirillales_uncultured	-1.94	0.54	0.00	Decreased	Sig	down_red	Family
Staphylococcaceae	-1.66	0.76	0.03	Decreased	Sig	down_red	Family
Pasteurellaceae	-1.53	0.55	0.01	Decreased	Sig	down_red	Family
Coriobacteriales_Incertae_Sedis	-1.45	0.50	0.00	Decreased	Sig	down_red	Family
Micrococcaceae	-1.37	0.64	0.03	Decreased	Sig	down_red	Family
Clostridia_UCG-014	-1.29	0.63	0.04	Decreased	Sig	down_red	Family
Moraxellaceae	-0.92	0.36	0.01	Decreased	Sig	down_red	Family
Ruminococcus	-2.72	0.88	0.00	Decreased	Sig	down_red	Genus
Enterobacter	-2.57	0.65	0.00	Decreased	Sig	down_red	Genus
Roseburia	-2.54	1.05	0.02	Decreased	Sig	down_red	Genus
Monoglobus	-2.30	0.68	0.00	Decreased	Sig	down_red	Genus
[Eubacterium]_ruminantium_group	-2.29	0.66	0.00	Decreased	Sig	down_red	Genus
Terrisporobacter	-2.20	0.91	0.02	Decreased	Sig	down_red	Genus
Romboutsia	-2.11	0.93	0.02	Decreased	Sig	down_red	Genus
Clostridium_sensu_stricto_1	-1.98	0.79	0.01	Decreased	Sig	down_red	Genus
Weissella	-1.86	0.61	0.00	Decreased	Sig	down_red	Genus
[Eubacterium]_brachy_group	-1.84	0.50	0.00	Decreased	Sig	down_red	Genus

d_Bacteria_Proteobacteria_Alphaproteobacteria_Rhodospirillales_uncultured_uncultured	-1.83	0.55	0.00	Decreased	Sig	down_red	Genus
[Eubacterium]_coprostanoligenes_group	-1.66	0.83	0.05	Decreased	Sig	down_red	Genus
Haemophilus	-1.63	0.49	0.00	Decreased	Sig	down_red	Genus
d_Bacteria_Actinobacteriota_Coriobacteriia_Coriobacteriales_Coriobacteriales_Incertae_Sedis_uncultured	-1.56	0.44	0.00	Decreased	Sig	down_red	Genus
Staphylococcus	-1.56	0.78	0.04	Decreased	Sig	down_red	Genus
Lactococcus	-1.53	0.52	0.00	Decreased	Sig	down_red	Genus
Sellimonas	-1.51	0.74	0.04	Decreased	Sig	down_red	Genus
Oscilibacter	-1.51	0.54	0.01	Decreased	Sig	down_red	Genus
Negativibacillus	-1.44	0.69	0.04	Decreased	Sig	down_red	Genus
d_Bacteria_Firmicutes_Clostridia_Oscillospirales_Oscillospiraceae_uncultured	-1.32	0.63	0.04	Decreased	Sig	down_red	Genus
Frisingicoccus	-1.30	0.54	0.02	Decreased	Sig	down_red	Genus
Fournierella	-1.24	0.46	0.01	Decreased	Sig	down_red	Genus
Anaerostipes	-1.22	0.56	0.03	Decreased	Sig	down_red	Genus
Intestinimonas	-1.19	0.50	0.02	Decreased	Sig	down_red	Genus
Clostridia_UCG-014	-1.18	0.60	0.05	Decreased	Sig	down_red	Genus
DTU089	-0.82	0.40	0.04	Decreased	Sig	down_red	Genus
Acinetobacter	-0.80	0.38	0.04	Decreased	Sig	down_red	Genus
d_Bacteria_Bacteroidota_Bacteroidia_Bacteroidales_Prevotellaceae_uncultured	2.27	1.07	0.03	Increased	Sig	up_blue	Genus

#### 4.4 Comparison between $\leq 1$ yr-OS and $> 1$ yr-OS

Taxon	FC	SD	p value	beta direction	p significance	color to fill p	tax level
Desulfuromonadia	-2.10	0.91	0.02	Decreased	Sig	down_red	Class
Clostridiales	-1.42	0.67	0.03	Decreased	Sig	down_red	Order
Burkholderiales	0.85	0.36	0.02	Increased	Sig	up_blue	Order
Bradymonadales	-1.75	0.88	0.05	Decreased	Sig	down_red	Family
Monoglobaceae	-1.56	0.70	0.03	Decreased	Sig	down_red	Family
Clostridiaceae	-1.54	0.66	0.02	Decreased	Sig	down_red	Family
[Eubacterium]_coprostanoligenes_group	-1.35	0.68	0.05	Decreased	Sig	down_red	Family
Defluviitaleaceae	-1.34	0.55	0.02	Decreased	Sig	down_red	Family
UCG-010	-1.33	0.66	0.04	Decreased	Sig	down_red	Family
Streptococcaceae	-1.02	0.48	0.04	Decreased	Sig	down_red	Family
Sutterellaceae	1.00	0.42	0.02	Increased	Sig	up_blue	Family
Ruminococcus	-2.10	0.67	0.00	Decreased	Sig	down_red	Genus
Bradymonadales	-1.70	0.83	0.04	Decreased	Sig	down_red	Genus
Prevotellaceae_UCG-004	-1.65	0.83	0.05	Decreased	Sig	down_red	Genus
[Eubacterium]_xylanophilum_group	-1.62	0.66	0.01	Decreased	Sig	down_red	Genus
NK4A214_group	-1.58	0.69	0.02	Decreased	Sig	down_red	Genus
Monoglobus	-1.51	0.66	0.02	Decreased	Sig	down_red	Genus
Lachnospiraceae_FCS020_group	-1.45	0.67	0.03	Decreased	Sig	down_red	Genus
Clostridium_sensu_stricto_1	-1.36	0.61	0.02	Decreased	Sig	down_red	Genus
[Eubacterium]_coprostanoligenes_group	-1.30	0.63	0.04	Decreased	Sig	down_red	Genus
Defluviitaleaceae_UCG-011	-1.29	0.55	0.02	Decreased	Sig	down_red	Genus
UCG-010	-1.29	0.59	0.03	Decreased	Sig	down_red	Genus
Intestinimonas	-1.05	0.46	0.02	Decreased	Sig	down_red	Genus
Sutterella	1.01	0.43	0.02	Increased	Sig	up_blue	Genus
Collinsella	1.80	0.83	0.03	Increased	Sig	up_blue	Genus
Acidaminococcus	2.41	0.78	0.00	Increased	Sig	up_blue	Genus

**Supplementary tables 5.** Gut microbiota correlation with clinical parameters

Feature	taxon	FC	SD	p value	q value	beta direction	p significance	q significance	color to fill p	color to fill q	tax level
hb_1_g_dl	Fusobacteriota	-1.08	0.31	0.00	0.01	Decreased	Sig	Sig	down_red	down_red	Phylum
hb_1_g_dl1	Fusobacteriia	-1.09	0.30	0.00	0.01	Decreased	Sig	Sig	down_red	down_red	Class
hb_1_g_dl2	Fusobacteriales	-1.08	0.30	0.00	0.02	Decreased	Sig	Sig	down_red	down_red	Order
alb_1_g_dl.1	Monoglobales	2.54	0.70	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Order
alb_1_g_dl.2	Christensenellales	2.68	0.70	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Order
db_1_mg_dl	Bacillales	-0.63	0.16	0.00	0.00	Decreased	Sig	Sig	down_red	down_red	Order
hb_1_g_dl3	Fusobacteriaceae	-1.10	0.31	0.00	0.03	Decreased	Sig	Sig	down_red	down_red	Family
cr_1_mg_dl	Tannerellaceae	-2.33	0.66	0.00	0.03	Decreased	Sig	Sig	down_red	down_red	Family
alb_1_g_dl.14	Monoglobaceae	2.51	0.69	0.00	0.02	Increased	Sig	Sig	up_blue	up_blue	Family
alb_1_g_dl.21	Christensenellaceae	2.66	0.68	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Family
tb_1_mg_dl	Coriobacteriales_Incertae_Sedis	-0.53	0.12	0.00	0.00	Decreased	Sig	Sig	down_red	down_red	Family
db_1_mg_dl.1	Bacillaceae	-0.65	0.16	0.00	0.01	Decreased	Sig	Sig	down_red	down_red	Family
db_1_mg_dl.2	Coriobacteriales_Incertae_Sedis	-0.56	0.12	0.00	0.00	Decreased	Sig	Sig	down_red	down_red	Family
hb_1_g_dl4	Hungatella	-0.94	0.23	0.00	0.01	Decreased	Sig	Sig	down_red	down_red	Genus
alb_1_g_dl.15	UCG-009	1.70	0.40	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.22	Turicibacter	2.03	0.50	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.3	Family_XIII_UCG-001	2.14	0.38	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.4	Monoglobus	2.49	0.67	0.00	0.04	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.5	UCG-003	2.84	0.59	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.6	Coprococcus	3.13	0.82	0.00	0.03	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.7	UCG-005	3.17	0.61	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.8	Christensenellaceae_R-7_group	3.23	0.62	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.9	Lachnospiraceae_FCS020_group	3.32	0.50	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.10	UCG-002	3.38	0.82	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.11	Family_XIII_AD3011_group	3.55	0.65	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
alb_1_g_dl.12	Lachnospiraceae_NK4A136_group	3.66	0.77	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus

alb_1_g_dl.13	Dorea	3.74	0.97	0.00	0.03	Increased	Sig	Sig	up_blue	up_blue	Genus
tb_1_mg_dl.1	Turicibacter	-0.51	0.13	0.00	0.03	Decreased	Sig	Sig	down_red	down_red	Genus
	d_Bacteria_Actinobacteri										
	ota_Coriobacteriia										
	Coriobacteriales_Coriobac	-0.50	0.12	0.00	0.01						
	teriales_Incertae_Sedis_un										
tb_1_mg_dl.2	cultured					Decreased	Sig	Sig	down_red	down_red	Genus
db_1_mg_dl.11	Turicibacter	-0.54	0.14	0.00	0.02	Decreased	Sig	Sig	down_red	down_red	Genus
	d_Bacteria_Actinobacteri										
	ota_Coriobacteriia										
	Coriobacteriales_Coriobac	-0.49	0.12	0.00	0.01						
	teriales_Incertae_Sedis_un										
db_1_mg_dl.21	cultured					Decreased	Sig	Sig	down_red	down_red	Genus
Acetic	Actinomycetales	-0.13	0.04	0.00	0.01	Decreased	Sig	Sig	down_red	down_red	Order
Isobutyric.1	Peptostreptococcales-	1.32	0.38	0.00	0.02	Increased	Sig	Sig	up_blue	up_blue	Order
	Tissierellales										
Isobutyric.2	Clostridiales	2.51	0.66	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Order
Isovaleric.1	Lactobacillales	-1.18	0.35	0.00	0.03	Decreased	Sig	Sig	down_red	down_red	Order
Isovaleric.2	Peptostreptococcales-	1.38	0.42	0.00	0.05	Increased	Sig	Sig	up_blue	up_blue	Order
	Tissierellales										
Isovaleric.3	Clostridiales	2.54	0.70	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Order
Acetic1	Actinomycetaceae	-0.13	0.04	0.00	0.02	Decreased	Sig	Sig	down_red	down_red	Family
Isobutyric.110	Anaerovoracaceae	1.48	0.36	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Family
Isobutyric.21	Oscillospiraceae	1.68	0.38	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Family
Isobutyric.3	Clostridiaceae	2.47	0.66	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Family
Isovaleric.16	Anaerovoracaceae	1.43	0.38	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Family
Isovaleric.21	Oscillospiraceae	1.83	0.46	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Family
Isovaleric.31	Clostridiaceae	2.51	0.69	0.00	0.02	Increased	Sig	Sig	up_blue	up_blue	Family
Isobutyric.111	Unknown	0.95	0.24	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.22	Lachnospiraceae_NC2004	1.07	0.26	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
	_group										
Isobutyric.31	UCG-009	1.39	0.35	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.4	Christensenella	1.42	0.34	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.5	Intestinimonas	1.60	0.39	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Firmicutes_C										
Isobutyric.6	lostridia_Oscillospirales_	1.83	0.38	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus

	Ruminococcaceae_uncultured										
Isobutyric.7	Anaerofilum	1.87	0.25	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.8	Lachnospiraceae_FCS020_group	1.93	0.50	0.00	0.03	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.9	Family_XIII_UCG-001	1.93	0.45	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.10	Phoceae	2.01	0.33	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.11	DTU089	2.02	0.52	0.00	0.02	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.12	Oscillibacter	2.06	0.51	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Firmicutes_Clostridia_Oscillospirales_Oscillospiraceae_uncultured	2.10	0.39	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.13	Negativibacillus	2.27	0.49	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.14	Romboutsia	2.34	0.63	0.00	0.05	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Actinobacteriota_Coriobacteriia_Coriobacteriales_Atopobiacae_uncultured	2.42	0.52	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.15	UCG-002	2.46	0.56	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.16	Clostridium_sensu_stricto_1	2.57	0.61	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.17	UCG-005	2.78	0.55	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.18	Family_XIII_AD3011_group	2.94	0.46	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isobutyric.19	Lachnospiraceae_NC2004_group	1.02	0.27	0.00	0.04	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.17	UCG-009	1.42	0.38	0.00	0.04	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.22	Christensenella	1.46	0.34	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.32	Anaerofilum	1.79	0.30	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Firmicutes_Clostridia_Oscillospirales_Ruminococcaceae_uncultured	1.83	0.44	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.4	Family_XIII_UCG-001	1.90	0.49	0.00	0.03	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.5	Phoceae	2.02	0.37	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.6	Oscillibacter	2.06	0.54	0.00	0.03	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Firmicutes_Clostridia_Oscillospirales_	2.13	0.45	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus

	Oscillospiraceae_uncultured										
Isovaleric.10	Negativibacillus	2.31	0.52	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.11	UCG-002	2.43	0.62	0.00	0.02	Increased	Sig	Sig	up_blue	up_blue	Genus
	d_Bacteria_Actinobacteriota_Coriobacteriia_Coriobacteriales_Atopobiacae_uncultured	2.51	0.52	0.00	0.00						
Isovaleric.12	Clostridium_sensu_stricto_1					Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.13		2.60	0.65	0.00	0.01	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.14	UCG-005	2.80	0.56	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Isovaleric.15	Family_XIII_AD3011_group	2.92	0.49	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus
Valeric	Anaerofilum	1.05	0.21	0.00	0.00	Increased	Sig	Sig	up_blue	up_blue	Genus

**Supplementary tables 6.** Shared predictive metabolite pathway and clinical outcomes

<b>PD (n=12)</b>	PD and Death (n=0)	PD, 6mpd and Death (n=0)	PD and 6mpd (n=7)	PD and OV (n=4)	PD, OV and Death (n=0)	PD, 6mpd, OV, and Death (n=0)	PD, 6mpd and OV (n=0)	OV (n=8)	OV and Death (n=0)	6mpd, OV and Death (n=0)	6mpd and OV (n=0)	Death (n=3)	6mpd and Death (n=2)	6mpd (n=4)
superpathway of L-arginine, putrescine, and 4-aminobutanoate degradation	not found	not found	erythromycin D biosynthesis	phospholipases	not found	not found	not found	Mandelate degradation I	not found	not found	not found	mixed acid fermentation	toluene degradation VI (anaerobic)	glycine betaine degradation I
<b>superpathway of L-arginine and L-ornithine degradation</b>			superpathway of 2,3-butanediol biosynthesis	mandelate degradation to acetyl-CoA				mycolyl-arabinogalactan-peptidoglycan complex biosynthesis				ppGpp bio-synthesis	androstene dione degradation	myo-inositol degradation I
<b>D-galactarate degradation I</b>			6-hydroxymethyl-dihydropterin diphosphate biosynthesis III (Chlamydia)	superpathway of aerobic toluene degradation				4-hydroxyphenylacetate degradation				tRNA processing		paromamine biosynthesis II
<b>superpathway of D-glucarate and D-galactarate degradation NAD salvage pathway II</b>			glycerol degradation to butanol	ethylmalonyl-CoA pathway				toluene degradation II I (aerobic) (via p-cresol)						coenzyme M biosynthesis I
<b>aerobactin biosynthesis</b>			6-hydroxymethyl-dihydropterin diphosphate biosynthesis I	D-glucarate degradation I				mannan degradation						
<b>superpathway of ornithine degradation</b>				androstenedione degradation				L-methionine salvage cycle I (bacteria and plants)						
<b>enterobacterial common antigen biosynthesis</b>								toluene degradation I (aerobic) (via o-cresol)						
<b>creatinine degradation II</b>								toluene degradation II (aerobic) (via 4-methylcatechol)						
<b>peptidoglycan biosynthesis II (staphylococci) sulfoglycolysis</b>														
<b>enterobactin biosynthesis</b>														

**Supplementary Table 7.** Univariate analysis of prognostic factors associated with poor outcomes.

Characteristics	Response evaluation	6m-PFS	1y-OS	OV
	p-value	p-value	p-value	p-value
Age	0.65	0.35	0.85	0.17
Sex	0.24	0.07	0.08	0.34
BW	0.74	1.00	0.51	0.64
Height	0.37	0.24	0.33	0.29
Smoking	0.49	0.28	0.30	0.93
Alcohol consumption	0.87	0.83	0.87	0.59
Raw food consumption	0.15	0.28	0.71	0.44
Cirrhosis	0.34	0.48	0.80	0.99
HBV infection	0.77	0.92	0.77	0.53
HCV infection	0.99	1.00	0.99	1.00
Antibiotics use	0.99	0.99	1.00	0.99
OV	0.06	<b>0.04</b>	0.21	1.00
Staging T	0.29	0.90	0.95	0.57
Staging N	0.80	0.48	0.80	0.89
Staging M	0.52	0.37	0.66	0.82
Hb	0.76	0.71	0.71	0.18
WBC	0.60	0.86	0.86	0.10
ANC	0.56	0.73	0.77	0.08
PMN	0.60	0.55	0.19	0.30
ALC	0.18	0.11	<b>0.02</b>	0.55
Lymphocyte	0.06	0.09	<b>0.03</b>	0.36
AEC	0.06	0.10	0.14	0.78
Eosinophils	<b>0.04</b>	0.06	0.11	0.52
Platelets	0.46	0.34	0.33	0.11
N/L ratio	0.23	0.11	0.07	0.22
P/L ratio	0.33	0.12	<b>0.05</b>	0.39
Creatinine	<b>0.03</b>	0.07	0.38	0.57
Alb	0.09	0.09	0.41	0.40
Glb	0.73	0.97	0.30	0.72
ALP	0.37	0.54	0.51	0.55
AST	0.30	0.44	0.39	1.00
ALT	0.49	0.50	0.39	0.60
TB	0.89	0.94	0.69	0.65
DB	0.85	0.92	0.56	0.64
CEA	0.42	0.58	0.33	0.41
CA 19-9	0.54	0.49	0.38	0.48
Log <sub>10</sub> CA 19-9	0.51	0.84	0.59	0.29
AFP	1.00	1.00	0.75	0.70
Oxidative stress	0.61	0.65	0.83	0.37
IL-10	0.61	0.51	0.55	0.63
IL-1B	0.43	0.37	0.56	0.61

Characteristics	Response evaluation	6m-PFS	1y-OS	OV
	p-value	p-value	p-value	p-value
IL-6	0.56	0.51	0.55	0.63
MCP-1	0.75	0.51	0.64	0.42
TNF-alpha	0.67	0.47	0.56	0.63
Chenodeoxycholic acid	0.49	0.51	0.78	0.15
Cholic acid	0.71	0.53	0.66	0.19
Glycocholic acid	0.37	0.44	0.79	0.66
Glycodeoxycholic acid and glyoursodeoxycholic acid	0.28	0.30	0.38	0.99
Taurochenodeoxycholic acid and tauroursodeoxycholic acid	0.06	0.12	0.15	0.62
Taurocholic acid	0.50	0.50	0.54	0.64
Acetic acid	0.76	0.37	0.51	0.11
Propionic acid	0.70	0.55	0.87	0.11
Isobutyric acid	0.25	0.41	0.85	0.28
Butyric acid	0.54	0.76	0.71	0.14
Isovaleric acid	0.31	0.51	0.98	0.35
Valeric acid	0.30	0.86	0.86	0.28
<i>Acidaminococcus</i>	0.10	0.18	0.13	0.31
<i>Sutterella</i>	0.07	0.13	0.13	0.29
<i>NK4A214_group</i>	0.93	0.68	0.55	0.82
<i>Lachnospiraceae_FCS020_group</i>	0.09	<b>0.04</b>	0.07	0.36
UCG-010	0.23	0.14	0.25	0.26
<i>Ruminococcus</i>	0.09	0.05	<b>0.05</b>	0.09
<i>Monoglobus</i>	0.06	<b>0.03</b>	<b>0.03</b>	0.18
<i>(25)_coprostanoligenes_group</i>	0.07	<b>0.02</b>	0.17	0.41
<i>Intestinimonas</i>	<b>0.05</b>	<b>0.03</b>	0.35	0.28

**Abbreviations:** AFP, alpha-fetoprotein; Alb, albumin; ALP, alkaline phosphatase; AST, aspartate aminotransferase; ALT, alanine transaminase; AEC, absolute eosinophil count; ALC, absolute lymphocyte count; ANC, absolute neutrophil count; BW, body weight; CA 19-9, cancer antigen 19-9; CEA, carcinoembryonic antigen; DB, direct bilirubin; Glb, globulin; Hb, hemoglobin; HBV, hepatitis B virus; HCV, hepatitis C virus; ICCA, intrahepatic cholangiocarcinoma; IL, interleukin; MCP-1, monocyte chemoattractant protein-1; N/L, neutrophil per lymphocyte; OS, overall survival; OV, *Opisthorchis viverrini*; PFS, progression-free survival; P/L, platelet per lymphocyte; TB, total bilirubin; TNF, tumor necrotic factor; WBC, white blood count.