

International Journal of Medical Sciences

2024; 21(8): 1438-1446. doi: 10.7150/ijms.96627

**Research Paper** 

# Analysis of prognostic biomarker models of TXNIP/NLRP3/IL1B inflammasome pathway in patients with acute myeloid leukemia

Junjie Chen<sup>1#</sup>, Qi Hou<sup>1#</sup>, Tao Chang<sup>2#</sup>, Jiamian Zheng<sup>1,3</sup>, Chao Yao<sup>1</sup>, Junyi He<sup>1</sup>, Shengting Chen<sup>4⊠</sup>, Xiuli Wu<sup>1,3⊠</sup>, Zhenyi Jin<sup>3,5⊠</sup>

1. The First Affiliated Hospital and Institute of Hematology, School of Medicine, Jinan University, Guangzhou, China.

- 2. Department of Biochemistry, School of Medicine, Jinan University, Guangzhou, China.
- 3. School of Medicine, Key Laboratory for Regenerative Medicine of Ministry of Education, Jinan University, Guangzhou, China.
- 4. Department of Blood Transfusion, The First Affiliated Hospital, Jinan University, Guangzhou, China.
- 5. Department of Pathology, School of Medicine, Jinan University, Guangzhou, China.

#Junjie Chen, Qi Hou, and Tao Chang contributed equally to this work.

🖂 Corresponding authors: Shengting Chen (shengtingchen@sina.cn), Xiuli Wu (siulier@163.com), and Zhenyi Jin (jinzhenyijnu@163.com).

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2024.03.25; Accepted: 2024.05.10; Published: 2024.05.27

#### Abstract

**Background:** Exploring potential biomarkers for predicting clinical outcomes and developing targeted therapies for acute myeloid leukemia (AML) is of utmost importance. This study aimed to investigate the expression pattern of the thioredoxin-interacting protein (TXNIP)/nucleotide-binding oligomerization domain (NOD)-like receptor protein 3 (NLRP3) pathway and its role in the prognosis of AML patients.

**Methods:** In this study, we examined the prognostic value of TXNIP/NLRP3 pathway in AML patients using microarray data from Gene Expression Omnibus (GEO) and transcriptome data from the Cancer Genome Atlas (TCGA) to develop a prognostic model and validated the results by quantitative real-time PCR (qRT-PCR) in a validation cohort of 26 AML patients and 18 healthy individuals from Jinan University (JNU) database.

**Results:** Analysis of the GSE13159 database revealed that TXNIP, interleukin 1 beta (IL1B) within the TXNIP/NLRP3 pathway were significantly upregulated and caspase1 (CASP1) was downregulated in AML patients (TXNIP, P = 0.031; IL1B, P = 0.042; CASP1, P = 0.038). Compared to high NLRP3 expression, AML patients with low NLRP3 expression had a longer overall survival (OS) in the GSE12417 dataset (P = 0.004). Moreover, both the training and validation results indicated that lower TXNIP, NLRP3, and IL1B expression were associated with favorable prognosis (GSE12417, P = 0.009; TCGA, P = 0.050; JNU, P = 0.026). According to the receiver operating characteristic curve analysis, this model demonstrated a sensitivity of 84% for predicting three-year survival. These data might provide novel predictors for AML outcome and direction for further investigation of the possibility of using TXNIP/NLRP3/IL1B genes in novel targeted therapies for AML.

Keywords: Acute myeloid leukemia, TXNIP, NLRP3, biomarker, prognosis

#### Introduction

Acute myeloid leukemia (AML) is a genetically heterogeneous disease, characterized by abnormal hematopoiesis, differentiation blockade and inadequate production of blood cells (1,2). It is the most common myeloid leukemia in adults (3,4). Previous research in our laboratory have shown that the prognosis of AML is closely related to immune checkpoints (5-7). In the era of tumor immunotherapy, the impact of inflammatory pathways on the effectiveness of anti-tumor immunity and tumor progression is increasing evident (8-10). Therefore, it is essential to study the inflammatory response in the tumor microenvironment.

Dysregulation of redox-controlled gene expression may be a common event in the pathogenesis of AML (11). It has been reported that increased levels of thioredoxin-interacting protein (TXNIP) contribute to the growth of leukemic cells (12). TXNIP, as a negative regulator of the key antioxidant system thioredoxin (TRX), is sensitive to reactive oxygen species (ROS) (13). It binds to the active cysteine residue of TRX and inhibits its antioxidative function. TXNIP also participates in other signaling pathways, playing roles in immune and inflammatory responses, glucose metabolism, and lipid metabolism. It is also involved in cell proliferation, differentiation, and apoptosis. TXNIP acts as a ligand for nucleotide-binding oligomerization domain (NOD)-like receptor protein 3 (NLRP3) (14). In the presence of high levels of ROS, elevated TXNIP protein activates NLRP3 inflammasome and activates a range of downstream genes, leading to chronic inflammatory and promoting tumor occurrence and progression (15,16). The NLRP3 inflammasome is one of the most well-known inflammasomes, consisting of NLRP3, apoptosisassociated speck-like protein containing CARD (ASC) and a CARD-domain effector protein pro-caspase-1 (17). In canonical or classical activation pathways, pathogen-associated molecular pattern molecules (PAMPs) or danger-associated molecular pattern molecules (DAMPs) induce NLRP3 oligomerization, recruiting the adapter ASCs and pro-caspase-1 to assemble the NLRP3 inflammasomes (18). This ultimately leads to the self-cleavage of the pro-caspase-1, forming activated caspase-1 (CASP1), which then cleaves pro- interleukin-1 $\beta$  (pro-IL-1 $\beta$ ) and pro- interleukin-18 (pro-IL-18) into their form active mature forms, IL-1 $\beta$  and IL-18, inducing inflammation and even cell death (19). It has been reported that the NLRP3 inflammasome is overexpressed and highly activated in AML bone marrow leukemia cells, and it is correlated with poor prognosis (20). However, the role of TXNIP/NLRP3 inflammation in AML patients remains unclear. This study aims to evaluate the potential value of inflammasome molecules in AML by examining their expression and prognostic significance. Based on the analysis from Gene Expression Omnibus (GEO), the Cancer Genome Atlas (TCGA) and our Jinan University (JNU) databases, we observed that lower expression pattern inflammasome-related of molecules was associated with better clinical outcomes. These results highlight the potential of inflammasome pathway as a promising biomarker for prognosis of AML patients.

## Materials and methods

#### Microarray data from GEO

We obtained microarray data from GEO database. We considered studies were eligible

according to the following standards: (1) studies with peripheral blood (PB) or bone marrow (BM) samples from healthy individuals (HIs) and AML (2) studies with information about the technology and platform utilized for studies. Based on these, we downloaded two datasets (GSE13159 and GSE12417) from the repository. GSE13159, including 542 AML patients and 74 HIs, was used to analyze relative gene expression and GSE12417, including 79 AML patients, was used to perform survival analysis.

#### **RNA** sequence data from TCGA database

Gene expression quantification data and clinical information of AML patients were collected from TCGA database. We utilized the R package "TCGAbiolinks" to download data that include a total of 167 AML patients from TCGA. Overall survival (OS) was defined as the time from diagnosis to death or last follow-up.

#### **PB** samples information from JNU dataset

A total of 26 PB samples were collected from the newly diagnosed AML patients at the First Affiliated Hospital of JNU database from March 1, 2014, to October 1, 2023. Additionally, 18 PB samples from age matching HIs, including 10 males and 8 females, with ages ranging from 24 to 66 years, with a median age of 36 years, were included as controls. The clinical information of the patients in the validation cohort was listed in Table 1. This study was performed in accordance with the principles of the Declaration of Helsinki and was approved by the Ethics Committee of the First Affiliated Hospital of Jinan University. All participants provided written informed consent.

## Quantitative real-time PCR (qRT-PCR)

Peripheral blood mononuclear cells (PBMCs) were separated from PB samples of the AML patients by Ficoll density centrifugation (Sigma Aldrich). Then total RNA was extracted from the PBMCs using TRIzol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions and was reversed transcribed into complementary DNA (cDNA) using PrimeScript<sup>™</sup> RT reagent Kit (Takara) according to the experimental instructions. The relative expression levels of TXNIP, NLRP3, CASP1, and IL1B were measured by qRT-PCR with SYBR Master Mix (TIANGEN, Beijing, China), and B2M was selected as an internal control. The primer sequences for qRT-PCR are shown in Table S1. The expression levels of TXNIP, NLRP3, CASP1, and IL1B are presented as  $2^{-\Delta\Delta CT*100}$ .

#### Statistical analysis

Statistical analyses were performed using Statistical Product and Service Solution (SPSS) (version 22.0, IBM, Armonk, NY, USA), GraphPad Prism (version 8.0, CA, USA) software and R (version 4.3.2). Comparison between the differences in mRNA expression levels between HIs and AML patients were analyzed by Mann Whitney U test for non-parametric values. The log-rank test conducted by R package "survminer" was used to compare differences in Kaplan-Meier curves. The restricted mean survival time (RMST) was obtained by the "survRM2" R package. The receiver operating characteristic (ROC) curve was performed by the "timeROC" R package. A two-tailed *P* value < 0.05 was statistically significant.

 Table 1. Clinical information of patients with acute myeloid leukemia

Variables	TCGA	JNU	GSE12417	GSE13159
Total, n	167	26	79	542
Gender, n (%)				
female	78 (46.8%)	10 (38.5%)	0 (0%)	0 (0%)
male	89 (53.2%)	16 (61.5%)	0 (0%)	0 (0%)
Missing	0 (0%)	0 (0%)	79 (100%)	542 (100%)
Age, years, median(range)				
Mean (SD)	55.0 (16.08)	52.8 (18.61)	59.8 (14.0)	-
Median (Min, Max)	58.0 (18.0, 88.0)	57.5 (20-86)	62.0 (18.0, 85.0)	-
Missing	0 (0%)	0 (0%)	0 (0%)	542 (100%)
FAB				
non-M3	154 (92.2%)	19 (73.1%)	79 (100%)	0 (0%)
M3	16 (7.8%)	2 (7.7%)	0 (0%)	0 (0%)
Missing	0 (0%)	5 (19.2%)	0 (0%)	542 (100%)
Risk, n (%)				
Favorable	33 (19.7%)	2 (7.7%)	0 (0%)	0 (0%)
Intermediate/Normal	97 (58.1%)	10 (38.5%)	0 (0%)	0 (0%)
Poor	35 (21.0%)	14 (53.8%)	0 (0%)	0 (0%)
Missing	2 (1.2%)	0 (0%)	79 (100%)	542 (100%)

Abbreviations: TCGA, the cancer genome atlas; GSE12417, gene expression omnibus series 12417 dataset; GSE13159, gene expression omnibus series 13159 dataset; JNU, Jinan University; FAB, French-American-British classification systems.

\*Due to rounding, not all percentages total 100%.

## Results

## Expression and clinical features of TXNIP/NPRP3 pathway related genes in AML

We initially assessed the expression levels of altered TXNIP/NLRP3 pathway in AML patients by analyzing the microarray data from the GEO database. The differentiation was analyzed between the 542 AML patients and 74 HIs from GSE13159 datasets. The expression levels of *TXNIP* and *IL1B* were significantly higher in AML patients than in HIs (P = 0.031 and P = 0.042, Figure 1A). However, the expression level of *CASP1* was lower in AML patients than in the HIs (P = 0.038). To further investigate the role of TXNIP/NLRP3 pathway in the clinical prognosis of AML patients, we analyzed the association between the expression levels of

TXNIP/NLRP3 pathway genes and OS of microarray data from 79 AML patients in the GSE12417 dataset by Kaplan-Meier curves. Based on median of gene expression level, we divided the patients into high and low expression groups. The results demonstrated that AML patients with lower *NLRP3* expression had a longer survival time and better OS (1-year OS: *NLRP3*<sup>low</sup> vs. *NLRP3*<sup>high</sup> 79% vs 44%, hazard ratio (HR) = 0.44, 95% confidence interval (CI): 0.24 to 0.79, *P* = 0.004, Figure 1B). However, *TXNIP*, *CASP1*, and *IL1B* expression in the GSE12417 dataset were not statistically significant (*P* = 0.139, *P* = 0.356, and *P* = 0.574, Figure 1B).

## Lower co-expression patterns of TXNIP/NPRP3 pathway related genes are associated with favorable OS in AML patients

Considering an additive effect on the outcome with multiple genes involved in the TXNIP/NLRP3 pathway, we further characterized the predictive value of co-expression patterns in AML. We found that AML patients with low expression of both TXNIP and NLRP3 had better OS in comparison with those with high expression of both genes or high expression of either gene alone in GSE12417 database (1-year OS:  $TXNIP^{high}NLRP3^{high}$ TXNIP<sup>low</sup>NLRP3<sup>low</sup> vs. vs. TXNIPhigh or NLRP3high, 91% vs. 48% vs. 50%, P =0.005, Figure 2A). To further investigate the relationship between TXNIP and its related genes, we combined three genes to screen out the combination mode that best significatively predict the OS in AML patients. According to the median values of single genes, AML patients were divided into triple low, triple high, and other groups. Intriguingly, we found that patients with low expression of TXNIP, NLRP3 and CASP1 or low expression of TXNIP, NLRP3 and IL1B had а better prognosis (1-year OS: TXNIPlowNLRP3lowCASP1low vs. TXNIPhighNLRP3high CASP1high vs. TXNIPhigh or NLRP3high or CASP1high, 89% 56% 51%, Р = 0.014;vs VS TXNIP<sup>high</sup>NLRP3<sup>high</sup> TXNIP<sup>low</sup>NLRP3<sup>low</sup>IL1B<sup>low</sup> vs. IL1Bhigh vs. TXNIPhigh or NLRP3high or IL1Bhigh, 100% vs 57% vs 50%, P = 0.009, Figure 2B, C). Meanwhile, RMST was used to evaluate the performance of the Kaplan-Meier curve, and we found had longer 3-year RMST in AML patients who are co-low expression of TXNIP, NLRP3 and CASP1 or TXNIP, NLRP3 and IL1B (3-year RMST: TXNIPlowNLRP3lowCASP1low vs. TXNIPhighNLRP3highCASP1high vs. *TXNIP*<sup>high</sup> or NLRP3high or CASP1high, 902 vs. 556 vs. 479 days; RMST: TXNIPlowNLRP3lowIL1Blow 3-year vs. TXNIPhighNLRP3highIL1Bhigh VS. **TXNIP**high or NLRP3high or IL1Bhigh, 946 vs. 600 vs. 490 days, Figure 2B and 2C).



Figure 1. Relative expression and OS analysis of TXNIP, NLRP3, CASP1, and IL1B in GSE13159 and GSE12417. (A) Relative expression of TXNIP, NLRP3, CASP1 and IL1B in AML patients and HIs. (B) OS analysis of high and low expression of TXNIP, NLRP3, CASP1, IL1B in GSE12417 dataset, Kaplan-Meier curves were plotted.

To elucidate the prognostic importance and verify the effects of co-expression patterns of the relationship between TXNIP and its related genes in AML patients, we further analyzed these genes in 167 AML patients from TCGA database. Using the co-expression patterns of TXNIP and related genes to evaluated the OS, higher OS was observed in AML patients with low expression of both TXNIP and NLRP3, and low expression of TXNIP and CASP1 compared to those with high expression of either gene or both genes (1-year OS: TXNIPlowNLRP3low vs. TXNIPhighNLRP3high vs. TXNIPhigh or NLRP3high, 77% vs. 53% vs. 45%, P = 0.001;  $TXNIP^{low}CASP1^{low}$  vs. TXNIPhighCASP1high vs. TXNIPhigh or CASP1high, 82% vs. 55% vs. 37%, *P* = 0.001; Figure 3A). Furthermore, AML patients with low expression of TXNIP, NLRP3 and CASP1 and low expression of TXNIP, NLRP3 and IL1B were significantly associated with favorable OS TXNIPlowNLRP3lowCASP1low (1-year OS: VS. TXNIPhighNLRP3highCASP1high **TXNIP**high vs. or  $NLRP3^{high}$  or  $CASP1^{high}$ , 86% vs 56% vs 44%, P = 0.001; TXNIPlowNLRP3lowIL1Blow vs. TXNIPhighNLRP3high IL1Bhigh vs. TXNIPhigh or NLRP3high or IL1Bhigh, 81% vs 45% vs 54%, P = 0.050, Figure 3B, C). Similarly, in TCGA database, AML patients with co-low

expression of *TXNIP*, *NLRP3* and *CASP1* or *TXNIP*, *NLRP3* and *IL1B* had longer RMST (3-year RMST: *TXNIP*<sup>low</sup>*NLRP3*<sup>low</sup>*CASP1*<sup>low</sup> vs. *TXNIP*<sup>high</sup>*NLRP3*<sup>high</sup> *CASP1*<sup>high</sup> vs. *TXNIP*<sup>high</sup> or *NLRP3*<sup>high</sup> or *CASP1*<sup>high</sup>, 787 vs. 558 vs. 476 days; 3-year RMST: *TXNIP*<sup>low</sup>*NLRP3*<sup>low</sup>*IL1B*<sup>low</sup> vs. *TXNIP*<sup>high</sup>*NLRP3*<sup>high</sup> *IL1B*<sup>high</sup> vs. *TXNIP*<sup>high</sup> or *NLRP3*<sup>high</sup> or *IL1B*<sup>high</sup>, 736 vs. 447 vs. 550 days, Figure 3B and 3C). These results were also confirmed in the validation cohort. Compared with patients with low co-expression of *TXNIP*, *NLRP3* and *IL1B*, those with high expression are at higher risk of death.

#### Validation of the prognosis value of lower co-expression of TXNIP/NLRP3/IL1B genes in JNU dataset

To further validate the prognostic value of TXNIP/NLRP3 pathway genes AML patients, we collected 26 AML patients and 18 HIs in our center. Compared to HIs, *CASP1* expression was significantly higher in AML patients (P = 0.014, Figure, 4A). In addition, the low expression of *TXNIP* had better OS in AML patients (1-year OS: *TXNIP*<sup>low</sup> vs. *TXNIP*<sup>high</sup>, 77% vs 45%, HR = 0.19, 95%CI: 0.05 to 0.81, P = 0.025, Figure 4B). The co-expression patterns in JNU dataset

Logrank P = 0.009

Times (days)

of TXNIP and related genes to evaluate the OS, higher OS was observed in AML patients with low expression of both TXNIP and NLRP3, compared to those with high expression of either gene or both genes, had a better prognosis (1-year OS: TXNIP<sup>low</sup>NLRP3<sup>low</sup> TXNIPhighNLRP3high vs. vs. TXNIPhigh or NLRP3high, 100% vs 25% vs 60%, P =0.003, Figure 4C). Furthermore, AML patients with low expression of TXNIP, NLRP3 and IL1B had a longer survival time compared to those with high

expression of either gene or all three genes (1-year OS: TXNIP<sup>low</sup>NLRP3<sup>low</sup>IL1B<sup>low</sup> vs. TXNIPhighNLRP3high IL1B<sup>high</sup> vs. TXNIP<sup>high</sup> or NLRP3<sup>high</sup> or IL1B<sup>high</sup>, 100% vs 33% vs 57%, P = 0.026, Figure 4D and 4E). In conclusion. think AML patients we with TXNIPlowNLRP3lowIL1Blow had а better OS. Meanwhile, the area under the receiver operating characteristic curve (AUC) showed that the sensitivity of this model for predicting three-year survival was 84% (95% CI: 52.91 to 114.16, Figure 4E).



Figure 2. Co-expression patterns of TXNIP, NLRP3, CASPI, and ILIB in AML patients in GSE12417 dataset. (A) OS analysis of TXNIPowNLRP3/ow, TXNIPowCASP1/ow, TXNIPowIL1B/ow, and (B)TXNIPowNLRP3/owCASP1/ow, (C) TXNIPowNLRP3/owIL1B/ow (left panel). The analysis of RMST, 3-year RMST was plotted (right panel).

1000

1500 0

3-year RMST:

500

490 (374 to 606)

1000

Survival Times (days)

1500

0 200

3-year RMST:

500

600 (343 to 857)

0.2

0.0

0

3-year RMST:

946 (826 to 1066)

600

1000 1400



Figure 3. Co-expression patterns of TXNIP, NLRP3, CASPI, and ILIB in AML patients in TCGA database. (A) OS analysis of TXNIPiowNLRP3low, TXNIPiowCASP11ow, TXNIPiowNLRP3lowCASP11ow, TXNIPiowNLRP3lowCASP11ow, (C) TXNIP

## Discussion

Inflammatory responses play a crucial role in the development and maintenance of inflammatory tumor environment, which supports tumor growth and promotes neoplastic transformation, invasion, and metastasis (21,22). The *TXNIP/NLRP3* pathway has been extensively studied in the context of inflammation and its inhibition by drugs, shedding light on the relationship between inflammation and disease (23,24).

We mainly focused on the prognostic biomarkers of TXNIP/NLRP3 pathway in the AML in this study.

We analyzed a total of 274 AML patients from 3 different databases (GEO, TCGA, and JNU) to assess OS and validate our findings. The results showed that low expression of NLRP3 predicted a better prognosis in AML patients in the GEO database and low expression of TXNIP predicted a better prognosis in AML patients in JNU database. Moreover, the co-expression of low TXNIP and NLRP3 were consistently associated with improved OS across all three databases. NLRP3 is considered an important intermediator between stressful stimuli and inflammatory responses, and its deregulation has been implicated in tumor progression. Previous studies have reported increased expression levels of NLRP3 inflammasome molecules in AML, suggesting their involvement in the disease (25, 26). Basiorka *et al.* reported activation of the NLRP3 inflammasome in hematopoietic stem and progenitor cells as a critical convergence signal in myelodysplastic syndromes (MDS), which direct activation of NLRP3 complexes and CASP1 and generation of IL-1 $\beta$  and pyroptotic cell death (27). These findings drive pyroptotic cell death and  $\beta$ -catenin activation and delineation of the

role of the pyroptosis in the clinical phenotype of MDS patients which suggesting new avenues for therapeutic intervention (28). There are also various mechanisms contribute to the activation of NLRP3 inflammasome, including TXNIP, calcium flux, and ROS (29). Specifically, TXNIP has been shown to bind to NLRP3 after dissociation of TXNIP from TRX in response to oxidative stress, thereby activating NLRP3 inflammasome (30). However, the functional roles of TXNIP in carcinogenesis remain controversial.



Figure 4. Validation results of JNU dataset (A) Relative expression of TXNIP, NLRP3, CASP1 and IL1B in AML patients and HIs. (B) Single expression patterns of TXNIP, NLRP3, CASP1, and IL1B were plotted to be forest plot. (C) Low co-expression TXNIP/NLRP3 AML patients had a better OS. (D) and (E) triple expression patterns of TXNIP, NLRP3, CASP1, IL1B and AUC of TXNIP/NLRP3/LL1B (right panel).

Although the expression of TXNIP alone, NLRP3 alone, or co-expression of TXNIP and NLRP3 cannot accurately predict the prognosis for all patients, combining TXNIP, NLRP3 and other genes may yield a more precise prognostic model. Our finding demonstrated that lower co-expression of TXNIP, NLRP3, and IL1B were associated with better OS of AML in all three databases. These findings could serve as valuable predictor of OS for AML. Several studies have reported a correlation between dysregulated IL-1β secretion and leukemia progression and poor prognosis (31). In addition, chronic stress has been shown to enhance infiltration and proliferation of AML cells, thereby worsening OS in AML mice models (32). These highlighted the oncogenic role of NLRP3/CASP1/IL-1β signaling in AML development, with IL-1 $\beta$  acting as a key mediator in disease progression (33). Therefore, targeting the combination of TXNIP, NLRP3, and IL-1 $\beta$  with more specific pharmacological inhibitors might be more beneficial for the treatment of AML.

## Conclusions

Taken together, our study reveals that lower co-expression of *TXNIP*, *NLRP3*, and *IL1B* is associated with a favorable prognosis in AML patients. These findings provide novel insight into evaluation and the design of combination targeted therapies for AML.

## Abbreviations

AML: ASC: Acute myeloid leukemia; Apoptosis-associated speck-like protein containing CARD; AUC: Area under the receiver operating characteristic curve; CASP1: Caspase1; CI: Confidence interval; DAMPs: Danger-associated molecular pattern molecules; GEO: Gene Expression Omnibus; HR: Hazard ratio; IL1B: Interleukin 1 beta; JNU: Jinan University; NLRP3: Nucleotide-binding oligomerization domain (NOD)-like receptor protein 3; OS: Overall survival; PAMPs: Pathogen-associated molecular pattern molecules; PBMCs: Peripheral blood mononuclear cells; pro-IL-18: Prointerleukin-18; pro-IL-1β: Pro-interleukin-1 $\beta$ ; **qRT-PCR**: Quantitative real-time PCR; **RMST**: Restricted mean survival time; ROC: Receiver operating characteristic; ROS: Reactive oxygen species; TCGA: The Cancer Genome Atlas; TXNIP: Thioredoxin-interacting protein; MDS: Myelodysplastic syndromes.

## Supplementary Material

Supplementary table. https://www.medsci.org/v21p1438s1.pdf

## Acknowledgments

We are immensely grateful to the healthy volunteers who generously donated their blood.

## Funding

This study was supported by grants from the Guangdong Basic and Applied Basic Research Foundation (Nos. 2020A1515010817; 2022A15150 10313; 2023A1515030271), the National Natural Science Foundation of China (82170220), the Science and Technology Program of Guangzhou City (No. 202201010164), the National Innovation and Entrepreneurship Training Program for Undergraduate (No. 202310559054) and the Guangdong College Students' Scientific and Technological Innovation (Nos. CX22446 and CX23304).

## Author contributions

ZYJ, XLW, and STC contributed to the concept development and study design. QH, TC, JMZ and CY performed the laboratory studies. JJC, JYH and CY collected the clinical data. ZYJ, JMZ and JJC participated in the manuscript and figure preparation. ZYJ, XLW and STC coordinated the study and helped draft the manuscript.

## Availability of data and materials

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

## **Ethical statement**

This study was performed in accordance with the principles of the Declaration of Helsinki and was approved by the Ethics Committee of the First Affiliated Hospital of Jinan University (JUNKY-2022-077). All participants provided written informed consent.

## **Consent for publication**

All authors read and approved the final manuscript.

## ORCID

Xiuli Wu: https://orcid.org/0000-0002-5202-6515; Zhenyi Jin: https://orcid.org/0000-0002-3812-3626.

## **Competing Interests**

The authors have declared that no competing interest exists.

## References

Döhner H, Weisdorf DJ, Bloomfield CD. Acute Myeloid Leukemia. New England Journal of Medicine 2015; 373:1136-1152.

- [2]. Zhong M, Gao R, Zhao R, Huang Y, Chen C, Li K, et al. BET bromodomain inhibition rescues PD-1-mediated T-cell exhaustion in acute myeloid leukemia. Cell Death Dis 2022; 13:671.
- [3]. Shallis RM, Wang R, Davidoff A, Ma X, Zeidan AM. Epidemiology of acute myeloid leukemia: Recent progress and enduring challenges. Blood Rev 2019; 36:70-87.
- [4]. Bewersdorf JP, Huntington SF, Zeidan AM. Cost-Effectiveness Analyses in AML: What Have We Learned, How Should This Impact Patient Care, and What Needs to Be Done in the Future? J Natl Compr Canc Netw 2023; 21:522-528.
- [5]. Chen C, Liang C, Wang S, Chio CL, Zhang Y, Zeng C, et al. Expression patterns of immune checkpoints in acute myeloid leukemia. J Hematol Oncol 2020; 13:28.
- [6]. Chen C, Wang P, Mo W, Zhang Y, Zhou W, Deng T, et al. IncRNA-CCDC26, as a novel biomarker, predicts prognosis in acute myeloid leukemia. Oncol Lett 2019; 18:2203-2211.
- [7]. Zhong M, Gao R, Zhao R, Huang Y, Chen C, Li K, et al. Correction to: BET bromodomain inhibition rescues PD-1-mediated T-cell exhaustion in acute myeloid leukemia. Cell Death Dis 2022; 13:743.
- [8]. Chen DW, Fan JM, Schrey JM, Mitchell DV, Jung SK, Hurwitz SN, et al. Inflammatory recruitment of healthy hematopoietic stem and progenitor cells in the acute myeloid leukemia niche. Leukemia 2024; 38:741-750.
- [9]. Mu M, Huang CX, Qu C, Li PL, Wu XN, Yao W, et al. Targeting Ferroptosis-Elicited Inflammation Suppresses Hepatocellular Carcinoma Metastasis and Enhances Sorafenib Efficacy. Cancer Res 2024; 84:841-854.
- [10]. He Y, Qu Y, Jin S, Zhang Y, Qin L. ALDH3A1 upregulation inhibits neutrophils n2 polarization and halts oral cancer growth. Oral Dis 2024;
- [11]. Huang D, Zhang C, Xiao M, Li X, Chen W, Jiang Y, et al. Redox metabolism maintains the leukemogenic capacity and drug resistance of AML cells. Proc Natl Acad Sci U S A 2023; 120:e2210796120.
- [12]. Erkeland SJ, Palande KK, Valkhof M, Gits J, Danen-van Oorschot A, Touw IP. The gene encoding thioredoxin-interacting protein (TXNIP) is a frequent virus integration site in virus-induced mouse leukemia and is overexpressed in a subset of AML patients. Leuk Res 2009; 33:1367-71.
- [13]. Hwang J, Suh HW, Jeon YH, Hwang E, Nguyen LT, Yeom J, et al. The structural basis for the negative regulation of thioredoxin by thioredoxin-interacting protein. Nat Commun 2014; 5:2958.
- [14]. Deng J, Pan T, Liu Z, McCarthy C, Vicencio JM, Cao L, et al. The role of TXNIP in cancer: a fine balance between redox, metabolic, and immunological tumor control. Br J Cancer 2023; 129:1877-1892.
- [15]. Liang M, Chen X, Wang L, Qin L, Wang H, Sun Z, et al. Cancer-derived exosomal TRIM59 regulates macrophage NLRP3 inflammasome activation to promote lung cancer progression. J Exp Clin Cancer Res 2020; 39:176.
- [16]. Jin P, Zhou Q, Xi S. Low-dose arsenite causes overexpression of EGF, TGFα, and HSP90 through Trx1-TXNIP-NLRP3 axis mediated signaling pathways in the human bladder epithelial cells. Ecotoxicol Environ Saf 2022; 247:114263.
- [17]. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. Cell 2006; 124:783-801.
- [18]. Ting JP-Y, Lovering RC, Alnemri ES, Bertin J, Boss JM, Davis BK, et al. The NLR gene family: A standard nomenclature. Immunity 2008; 28:285-7.
- [19]. Liu J, Qi X, Gu P, Wang L, Song S, Shu P. Baicalin Induces Gastric Cancer Cell Pyroptosis through the NF-κB-NLRP3 Signaling Axis. J Cancer 2024; 15:494-507.
- [20]. Zhong C, Wang R, Hua M, Zhang C, Han F, Xu M, et al. NLRP3 Inflammasome Promotes the Progression of Acute Myeloid Leukemia via IL-1β Pathway. Front Immunol 2021; 12:661939.
- [21]. Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. Nat Med 2018; 24:541-550.
- [22]. Junttila MR, de Sauvage FJ. Influence of tumour micro-environment heterogeneity on therapeutic response. Nature 2013; 501:346-54.
- [23]. Jia Q, Zhu R, Tian Y, Chen B, Li R, Li L, et al. Salvia miltiorrhiza in diabetes: A review of its pharmacology, phytochemistry, and safety. Phytomedicine 2019; 58:152871.
- [24]. Zhang W, Shi C, Yao Z, Qian S. Bardoxolone methyl attenuates doxorubicin-induced cardiotoxicity by inhibiting the TXNIP-NLRP3 pathway through Nrf2 activation. Environ Toxicol 2024; 39:1936-1950.
- [25]. Rahman T, Nagar A, Duffy EB, Okuda K, Silverman N, Harton JA. NLRP3 sensing of diverse inflammatory stimuli requires distinct structural features. Front Immunol 2020; 11:1828.
- [26]. Jia Y, Zhang C, Hua M, Wang M, Chen P, Ma D. Aberrant NLRP3 inflammasome associated with aryl hydrocarbon receptor potentially contributes to the imbalance of T-helper cells in patients with acute myeloid leukemia. Oncol Lett 2017; 14:7031-7044.
- [27]. Basiorka AA, McGraw KL, Eksioglu EA, et al. The NLRP3 inflammasome functions as a driver of the myelodysplastic syndrome phenotype. Blood. 2016. 128(25): 2960-2975.
- [28].Sallman DA, Cluzeau T, Basiorka AA, List A. Unraveling the Pathogenesis of MDS: The NLRP3 Inflammasome and Pyroptosis Drive the MDS Phenotype. Front Oncol. 2016. 6: 151.
- [29]. Yoshihara E, Masaki S, Matsuo Y, Chen Z, Tian H, Yodoi J. Thioredoxin/Txnip: redoxisome, as a redox switch for the pathogenesis of diseases. Front Immunol 2014; 4:514.

- [30]. Zhou R, Tardivel A, Thorens B, Choi I, Tschopp J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. Nat Immunol 2010; 11:136-40.
- [31]. Ratajczak MZ, Bujko K, Cymer M, Thapa A, Adamiak M, Ratajczak J, et al. The Nlrp3 inflammasome as a "rising star" in studies of normal and malignant hematopoiesis. Leukemia 2020; 34:1512-1523.
- [32]. Liu N, Wu Y, Wen X, Li P, Lu F, Shang H. Chronic stress promotes acute myeloid leukemia progression through HMGB1/NLRP3/IL-1β signaling pathway. J Mol Med (Berl) 2021; 99:403-414.
- [33]. Moossavi M, Parsamanesh N, Bahrami A, Atkin SL, Sahebkar A. Role of the NLRP3 inflammasome in cancer. Mol Cancer 2018; 17:158.