

Research Paper

Proteomic analysis of Exosomes derived from the Aqueous Humor of Myopia Patients

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Abstract

Objectives: Myopia is the most common refractive vision disorder. In recent years, several studies have suggested that the alteration of the exosomal protein levels in the aqueous humor (AH) is associated with the development of several eye diseases. Therefore, we aimed to explore the exosomal protein profile of the AH from myopia patients.

Methods: Exosomes were isolated from the AH. The quality, concentration, and size distribution of exosomes for each patient were measured using nanoparticle tracking analysis system. Then, the exosomal proteins were purified and digested by trypsin for liquid chromatography-tandem mass spectrometry (LC-MS/MS) analysis.

Results: There was no significant difference observed between the myopia and control when comparing the concentration and size distribution of exosomes in the AH for each sample. Based on LC-MS/MS analysis, myopia patients had higher and more complex exosomal peptide content. We found two proteins that were common in AH exosomes and eight proteins that were highly expressed in the myopia group.

Conclusions: Our results provide pioneering findings for the exploration of the exosomal protein profile in myopia development. Further studies may provide significant information for the diagnosis, clinical treatment, and prognosis of myopia.

Key words: aqueous humor; exosome; myopia

Introduction

Myopia is the most common refractive vision disorder where light focuses in front of the retina [1]. The severity of myopia is usually measured in diopters: low, -3.00 diopters or less; moderate, -3.00 to -6.00 diopters; and high, more than -6.00 diopters [2]. Genetic and environmental factors have been shown to influence the occurrence and development of myopia, but the detailed mechanism is still largely unknown [3]. Myopia is known as a milder disease for the optical error can be corrected by wearing glasses,

contact lenses, or refractive surgery [2, 4]. However, it is becoming an ever-increasing global health concern as the prevalence of myopia has risen sharply in the past 50 years [5]. In Taiwan, the prevalence and severity of myopia among the younger generation have increased rapidly over the past two decades [6]. The rupture rate is as high as 70.3% among individuals 12-19 years old but only 5.6% among individuals older than 65 years [7]. With a higher degree of myopia, there is greater risk for

complications such as macular degeneration, retinal detachment, cataracts, and glaucoma [8]. Myopia prevention and treatment is an urgent public health issue in Taiwan.

The aqueous humor (AH) is the clear liquid filling in the anterior and posterior chamber of the eye [9]. It is secreted by the pigment-free epithelial cells of the ciliary body and serves for many important functions [9]. First, the dynamic balance of AH production and discharge is important to maintain the shape and optical properties of the eye [10]. AH fluid is also responsible for supplying nutrients and removing cellular waste for intraocular tissues [9, 10]. Recently, several studies have suggested that the alteration of protein levels in AH associated with the development of several eye diseases [11-16]. For example, the severity of corneal edema in eyes with bullous keratopathy has been associated with the levels of specific cytokines in the AH [17]. Elevated TGF β 2 (transforming growth factor β 2) levels in AH of cataract patients is a potential risk factor for capsular contraction syndrome [18, 19]. Therefore, studying the composition of the AH will help to understand the process of disease development and to explore the clinical treatments.

Exosomes are composed of a lipid bilayer containing transmembrane proteins, enclosing cytosolic proteins, and RNAs [20, 21]. They are secreted from cells and are released into body fluids (i.e., blood, urine, tears, and spinal fluid), which can be transported between adjacent or distant cells via the circulatory system [21]. They are considered as important messengers for cell-to-cell communication [21]. An increasing number of studies have reported that proteins or miRNAs in exosomes derived from the AH may be associated with ophthalmic diseases [22-25]. For example, trabecular meshwork cells released exosomes containing myocilin under the stimulation of the external environment [26, 27]. Different expression profiles of exosomal miRNAs were found to be associated with glaucoma and myopia [28, 29]. To analyze the alteration of exosomal proteins that occur with myopia, we aimed to compare the individual protein profile from nine high myopia patients with nine control patients by using liquid chromatography-tandem mass spectrometry (LC-MS/MS).

Materials and Methods

AH sample collection

The study protocol was approved by the Research Ethics Committee of Taipei City Hospital (TCHIRB-10604109) and conducted according to the tenets of the Declaration of Helsinki. All study

participants provided written informed consent before their enrollment. Human AH samples were collected from patients undergoing cataract surgery at the Zhongxing branch of Taipei City Hospital. The diagnostic criterion for myopia was defined as axial length of more than 26 mm. The control eyes were collected from senile cataract patients who were free from other ocular or systemic diseases. Approximately 100 μ L of AH was collected from each patient by anterior chamber paracentesis, using a needle inserted through the peripheral cornea at the beginning of the procedure. Undiluted AH samples were collected in sterile tubes and stored at -80 °C until further study.

Isolation of Exosomes from the AH

Exosomes were isolated from the AH using the ExoQuick precipitation solution (System Biosciences, Inc., Mountain View, CA) according to its recommended procedure. Approximately 100 μ L of AH was centrifuged at 3000 \times g for 15 minutes to remove cellular debris, after which the supernatants were collected. Phosphate-buffered saline (PBS) buffer was added to the supernatants to achieve a final volume of 250 μ L. Then, 63 μ L of precipitation solution was added to the supernatants and incubated overnight. The exosome pellets were collected by centrifugation at 12,000 \times g for 90 minutes and then suspended in 100 μ L PBS. The concentration and size distribution of the vesicles were measured by NanoSight NS10 (Malvern Instruments, Rancho Cucamonga, CA).

Proteins purification and LC-MS/MS analysis

Exosomes were lysed by radioimmuno-precipitation assay (RIPA) buffer to extract the exosomal proteins. Then, the proteins were digested by trypsin (SMART Digest Trypsin Kit, Thermo Fisher Scientific; Waltham, MA), desalted (Millipore® Ziptips Micro-C18; Sigma-Aldrich, Milwaukee, WI), purified (SOLA μ ™ SPE Plates; Thermo Fisher Scientific; Waltham, MA), and dissolved in 0.1% formic acid for LC-MS/MS analysis (LTQ Orbitrap Velos, Thermo Scientific; Waltham, MA) (service provided by the Mass Core Facility of Genomics Research Center, Academia Sinica). The acquired proteomic raw data files were then applied to search against a UniProt human protein database by using PEAKS Studio 7.5 (Bioinformatics Solutions, Waterloo, Ontario, Canada). The settings in PEAKS Studio 7.5 combined with UniProt for searching the protein database were as follows: enzyme set as trypsin with a maximum of two missed cleavage site precursor and fragment mass tolerance of 20 ppm and 0.8 Da, respectively. Finally, spectral counts obtained

from each peptide were normalized to the total spectral counts recorded for all peptides in a sample.

Statistical analyses

Data are expressed as the mean ± standard error of the mean. The measurements of axial lengths and exosome concentration were analyzed using a paired sample t-test. All data analysis was performed using SPSS version 24 (SPSS, Inc; Chicago, IL). A P value of less than 0.05 was considered to indicate a statistically significant difference. The principal component analysis (PCA) plot of all samples was generated using Partek Genomics Suite 7.18 (Partek Inc., St. Louis, MO).

Results

Samples collection and exosome isolation

The AH samples were collected from 18 patients undergoing cataract surgery at Taipei City Hospital. The patients were assigned to the myopia group if their axial length was more than 26 mm. There were no significant differences in the clinical characteristics between myopia and control group, except the axial length (Table 1). Exosomes were isolated from the AH by using exosome purification kit for each individual patient. The concentration and the size distribution of the purified small vesicles were determined by nanoparticle tracking analysis system. Calculating the diameter of these small vesicles, most were found to be in the expected range of exosomes (30-150 nm) (Fig. 1). Comparing the concentration of exosomes in AH for each sample, there was no significant difference observed between myopia and control groups (Table 2).

Table 1. Information of patients with and without myopia

Group	Number	Gender	Age	Axial length (mm)
Myopia	M1	M	65	27.27
	M2	M	73	27.56
	M3	M	60	28.26
	M4	F	64	30.05
	M5	M	49	27.80
	M6	F	60	27.27
	M7	M	73	27.56
	M8	M	59	28.64
	M9	M	68	26.78
			63.44 ± 7.57	27.91 ± 0.97
Control	N1	F	53	23.18
	N2	F	77	23.10
	N3	M	71	24.96
	N4	F	61	22.79
	N5	M	86	25.59
	N6	F	90	23.31
	N7	M	58	23.30
	N8	M	72	23.29
	N9	F	82	22.34
			72.22 ± 12.86	23.54 ± 1.04

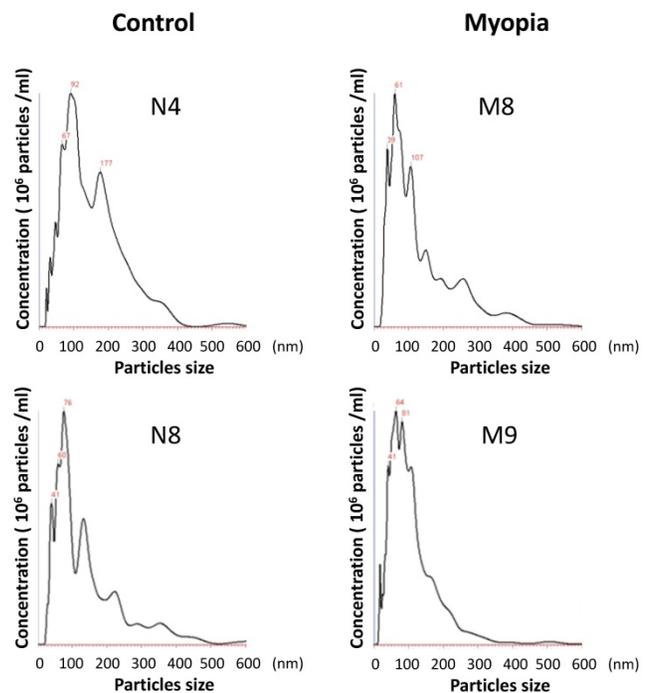


Figure 1. The size distribution of AH exosomes. Representative nanoparticle tracking analysis of exosomes isolated from the AH sample of patients with myopia (M) and controls (N).

Table 2. Concentration of exosomes in AH

		10 ⁵ particles/μl
Myopia	M1	2.35
	M2	1.87
	M3	3.25
	M4	2.83
	M5	2.03
	M6	3.66
	M7	2.3
	M8	8.48
	M9	13.77
	AVE.	4.29±4.01
Control	N1	0.3
	N2	7.7
	N3	4.6
	N4	3.36
	N5	5.76
	N6	0.82
	N7	2.47
	N8	4.45
	N9	7.83
	AVE.	4.14±2.87

Exosomal protein isolation and LC-MS/MS analysis

Proteins isolated from the exosomes were purified and digested by trypsin for LC-MS/MS quantitation. As shown in Figure 2A, the number of peptides identified from the myopia group was more than that in the control group. PCA was applied to further demonstrate similarities and differences in the exosomal peptides content. On a PCA plot, samples with similar peptide profiles can be positioned in proximity to each other. The position of each sample

was plotted against the PC1, PC2, and PC3 axes in a three-dimensional space. The PCA resulted in the delineated of two distinct positions in the three-dimensional space for myopia and control groups (Fig. 2B). In addition, the myopia group presented more diversity than the control group (Fig. 2B).

Potential protein makers for AH exosomes

In summary, 164 peptides were identified from myopia and control exosomes. By searching the peptide sequences in the UniProt peptide database (www.uniprot.org), we obtained their protein names. Comparing our results with previous proteomic

analysis of exosome from the AH, there were 15 proteins in our results have been reported in the studies of age-related macular degeneration patients [25]. More interestingly, we found that two of them, apolipoprotein A1 (APOA1) and opticon (OPTC), commonly had high expression in either myopia or control patients (Fig. 3A). To confirm our finding, we next test their protein expression in exosome from retinal pigmented epithelium cell lines (ARPE19). Western blot results indicated both APOA1 and OPTC were present in exosomes (Fig. 3B). APOA1 protein has been found in exosomes derived from saliva and urine [30, 31], but there are no reports

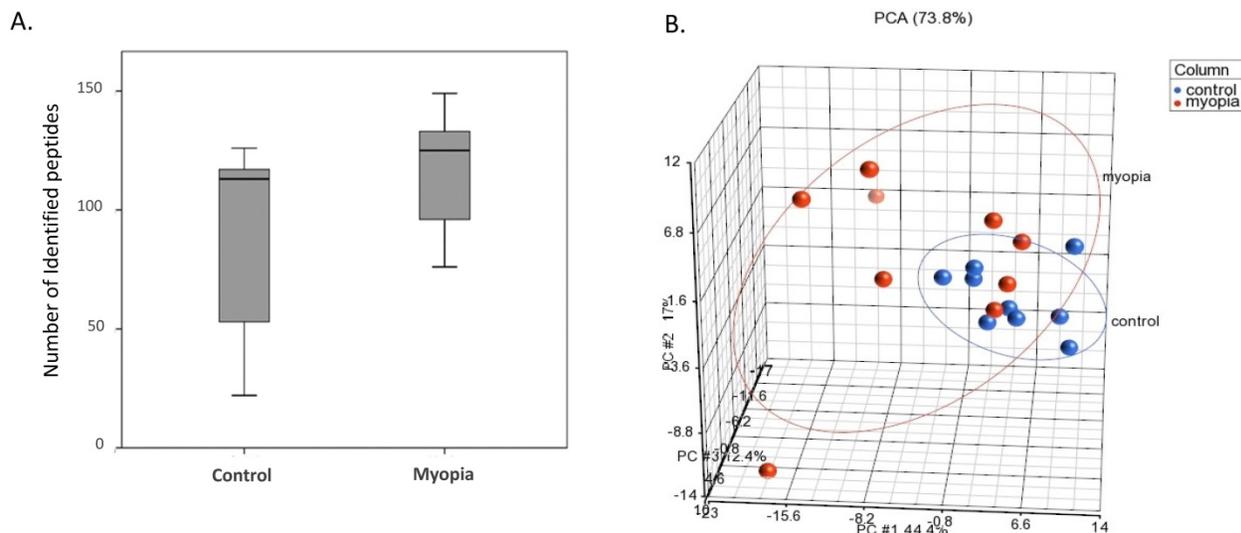


Figure 2. LC-MS/MS-identified peptides from myopia and control groups. A) Box plot of identified peptides from individual myopia and control patients. B) The PCA plot of all samples was generated to assess the variability of peptide expression in myopia and control patients. (red, myopia; blue, control).

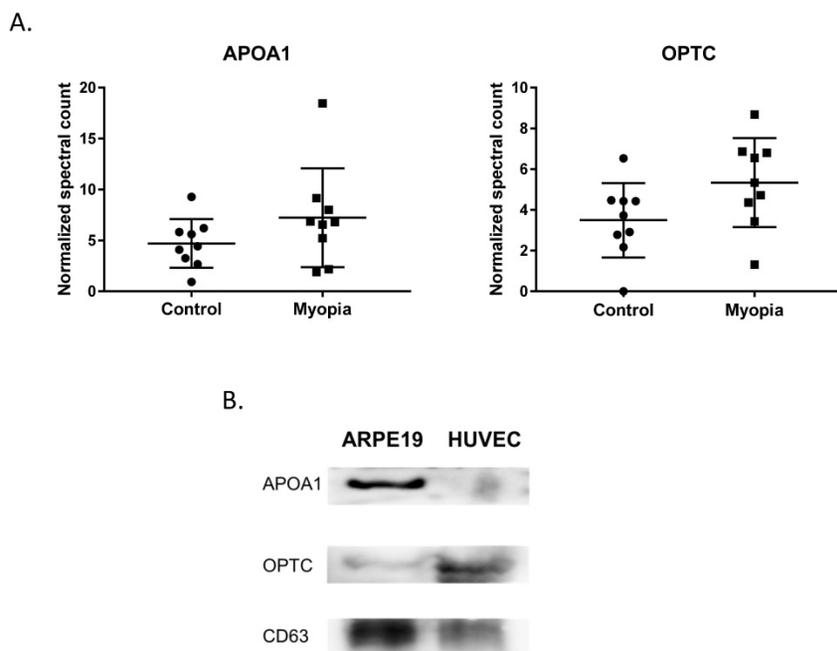


Figure 3. Common proteins in AH exosome. A) Normalized spectral count (NSC) of AH common proteins detected by LC-MS/MS. Scatterplot presenting the mean \pm SD for the NSC of proteins detected in each patient from myopia or control. B) Exosomes proteins from ARPE19 (retinal pigmented epithelial cell) and HUVEC (umbilical vein endothelial cells) were analyzed by western blotting. CD63 is a well-known common exosome marker.

suggesting OPTC as an exosomal protein. Because OPTC expression is common in various eye tissues [32], we suggested that OPTC could be a potential AH-specific exosomal marker.

Myopia specific exosomal proteins

Comparing the exosomal peptides between myopia and control patients, 35 peptides were significantly different between myopia and control groups (t- test, $P < 0.05$) (Table 3). These peptides belong to 8 known (Fig. 4) and 17 unidentified proteins. More interestingly, all were significantly increased in myopia patients, suggesting that myopia patients secreted more exosomal proteins to the AH. In previous studies, transthyretin (TTR) and hemopexin (HPX) have been detected in the serum of high myopia patients [33, 34]. In addition, TTR has also been found in the AH of high myopia patients [35]. In summary, these results support our finding that these are myopia-specific exosomal proteins. Further studies are necessary to explore their roles in myopia.

Discussion

In this study, we analyzed the exosomal protein content from the AH of the high myopia and control by LC-MS/MS. By analyzing the exosomal protein

profile, we found that two proteins, APOA1 and OPTC, were common in the AH exosomes and could be potential exosomal protein markers for AH. Comparing the protein profile between myopia and control groups, 25 proteins showed significant enrichment in myopia patients. Two of these myopia-specific exosomal proteins, TTR and HPX have been found in previous myopia associated research. Further studies exploring their role in the development of myopia, would be beneficial. To our knowledge, we are the first to analyze the individual exosomal protein profile of the AH.

We found two proteins that were commonly had high expression in both myopia and control groups. APOA1 encodes apolipoprotein A I, which is the major protein component of high-density lipoprotein (HDL) in the plasma [36]. APOA1 protein has been found in exosomes derived from the saliva, urine, platelets, and hepatocytes [30, 31, 37, 38]. OPTC is a member of the small leucine-rich repeat protein (SLRP) family and is present in various eye tissues such as vitreous, cornea, iris, ciliary body, optic nerve, choroid, and retina [32]. There are no previous reports showing OPTC as an exosomal protein. We suggest OPTC could be an AH-specific exosomal marker.

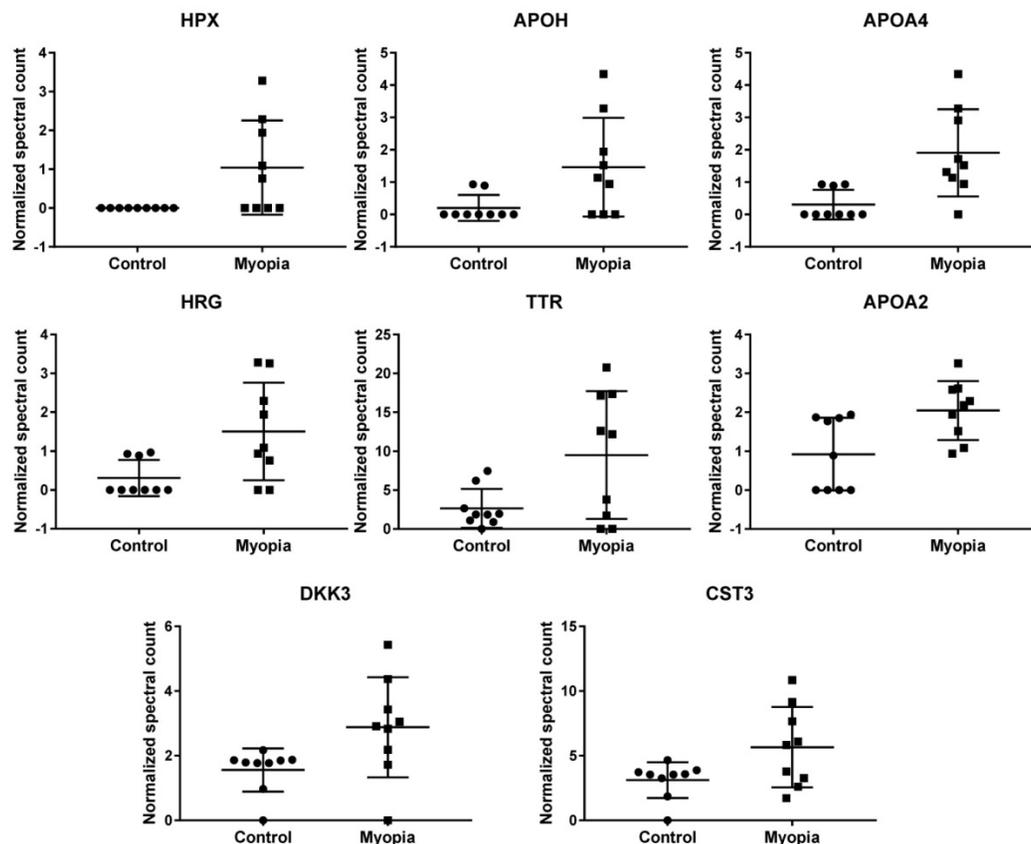


Figure 4. Normalized spectral count (NSC) of myopia-specific proteins were detected from LC-MS/MS. Scatterplot presenting the mean \pm SD for the NSC of proteins detected in each patient from myopia or control. P values were calculated by paired two-sample t tests.

Table 3. Significantly changed exosomal proteins in myopia patients

Protein IDs	Protein Name	Gene Name	Control	Myopia	<i>p</i> -value
			Mean NSC ± SD	Mean NSC ± SD	
P02790	Hemopexin	HPX	0 ± 0	1.039 ± 1.213	3.32E-02
P02749	Beta-2-glycoprotein 1	APOH	0.202 ± 0.401	1.464 ± 1.525	3.96E-02
P06727	Apolipoprotein A-IV	APOA4	0.305 ± 0.458	1.908 ± 1.346	7.17E-03
B4DPN0	cDNA FLJ51265		0.202 ± 0.401	1.137 ± 1.134	4.20E-02
B4E1B2	cDNA FLJ53691		0.617 ± 1.546	3.384 ± 3.434	4.94E-02
P04196	Histidine-rich glycoprotein	HRG	0.31 ± 0.465	1.506 ± 1.254	2.26E-02
B2R8I2	cDNA FLJ93914		0.31 ± 0.465	1.506 ± 1.254	2.26E-02
A6XMH1	Transthyretin	TTR	2.662 ± 2.503	9.51 ± 8.2	3.88E-02
A0A087WV45	Transthyretin	TTR	2.662 ± 2.503	9.51 ± 8.2	3.88E-02
A0A087WT59	Transthyretin	TTR	3.618 ± 2.145	10.355 ± 7.589	2.99E-02
E9KL36	Transthyretin	TTR	4.641 ± 2.917	13.16 ± 9.822	3.31E-02
P02766	Transthyretin	TTR	4.641 ± 2.917	13.16 ± 9.822	3.31E-02
A6XGL1	Transthyretin	TTR	4.641 ± 2.917	13.16 ± 9.822	3.31E-02
Q6DHW4	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q5CZ94	DKFZp781M0386		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q6PJG0	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q6P2J1	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q7Z2U7	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q6GMV8	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
A0A5E4	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q8NEJ1	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
Q6P5S3	Uncharacterized protein		0.93 ± 0.819	2.084 ± 0.875	1.07E-02
P02652	Apolipoprotein A-II	APOA2	0.924 ± 0.93	2.046 ± 0.754	1.29E-02
V9GYM3	Apolipoprotein A-II	APOA2	0.924 ± 0.93	2.046 ± 0.754	1.29E-02
O43532	RIG-like 7-1		1.562 ± 0.668	2.88 ± 1.548	3.90E-02
Q8N294	cDNA FLJ33633		1.562 ± 0.668	2.88 ± 1.548	3.90E-02
B3KS70	cDNA FLJ35660		1.562 ± 0.668	2.88 ± 1.548	3.90E-02
B4DI69	cDNA FLJ59893		1.562 ± 0.668	2.88 ± 1.548	3.90E-02
Q9UBP4	Dickkopf-related protein 3	DKK3	1.562 ± 0.668	2.88 ± 1.548	3.90E-02
F6SYF8	Dickkopf-related protein 3	DKK3	1.562 ± 0.668	2.88 ± 1.548	3.90E-02
B4DID6	cDNA FLJ52545		1.562 ± 0.668	2.88 ± 1.548	3.90E-02
P01034	cystatin-C	CST3	3.116 ± 1.381	5.661 ± 3.109	4.63E-02
A0A0K0K1J1	cystatin C	CST3	3.116 ± 1.381	5.661 ± 3.109	4.63E-02

#NSC: Normalized spectral count.

By comparing the exosomal protein profile between myopia and control groups, we identified 25 myopia-specific exosomal proteins (Table 3). In previous studies, Shao et al. showed that the expression of HPX and TTR in serum were positively correlated with myopia patients [33]. Higher TTR levels have also been detected in the vitreous of myopia, macular detachment, and macular hole patients [39]. Recently, exosomes have been indicated to pass through the blood-brain barrier [40], it would be possible for the AH exosomes carrying HPX or TTR to also pass through the blood-retinal barrier. In the eye development of zebrafish model, the expression of DKK3 was suggested to be associated with eye size [41].

Our results provide pioneer findings for the exploration of the exosomal protein profile in myopia development. Further studies identifying the target tissues of these exosomes and understanding the function of these myopia-specific proteins may provide significant information for the diagnosis, clinical treatment, and prognosis of myopia.

Abbreviations

AH: aqueous humor; APOA1: apolipoprotein A1; HPX: hemopexin; LC-MS/MS: liquid chromatography-tandem mass spectrometry; OPTC: optician; PCA: principal component analysis; TTR: transthyretin.

Acknowledgements

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Competing Interests

The authors have declared that no competing interest exists.

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