

## Research Paper

# A Novel Model for Studying Voltage-Gated Ion Channel Gene Expression during Reversible Ischemic Stroke

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## Abstract

The dysfunction of voltage-gated ion channels contributes to the pathology of ischemic stroke. In this study, we developed rat models of transient ischemic attack (TIA) and reversible ischemic neurological deficit (RIND) that was induced via the injection of artificial embolic particles during full consciousness, that allow us to monitor the neurologic deficit and positron emission tomography (PET) scans in real-time. We then evaluated the infarction volume of brain tissue was confirmed by 2,3,5-triphenyl tetrazolium chloride (TTC) staining, and gene expressions were evaluated by quantitative real-time PCR (qPCR). We found that rats with TIA or RIND exhibited neurological deficits as determined by negative TTC and PET findings. However, the expression of voltage-gated sodium channels in the hippocampus was significantly up-regulated in the qPCR array study. Furthermore, an altered expression of sodium channel  $\beta$ -subunits and potassium channels, were observed in RIND compared to TIA groups. In conclusion, to our knowledge, this is the first report of the successful evaluation of voltage-gated ion channel gene expression in TIA and RIND animal models. This model will aid future studies in investigating pathophysiological mechanisms, and in developing new therapeutic compounds for the treatment of TIA and RIND.

Key words: animal model, embolic stroke, transient ischemic attack, reversible ischemic neurological deficit, voltage-gated ion channels

## Introduction

Stroke is the leading cause of disabilities in adults and the primary cause of death worldwide. Additionally, stroke-induced brain damage can often lead to seizures and epilepsy [1-3].

A TIA is a brief episode of neurological dysfunction caused by the loss of blood flow to the brain, spinal cord, or retina. This usually arises from a blood clot or a dislodged atherosclerotic plaque blocking a blood vessel in the brain. A TIA exhibits

the same underlying characteristics as a stroke, i.e., a disruption of cerebral blood flow, and is frequently referred to as a mini-stroke. Additionally, TIAs and strokes share similar symptoms, such as contralateral paralysis and epilepsy [4, 5]. However, unlike a stroke, the symptoms of a TIA resolve quickly ranging from a few minutes to 24 h [6]. Stroke symptoms that last longer than 24 h but fewer than 72 h are called RIND [4]. Importantly, the very early risk of stroke

after a first TIA is highlight in previous study [7]. Therefore, medical attention is sought more urgently. Currently, there is no suitable animal model for studying TIA and RIND, so the molecular mechanisms underlying these disorders remain unclear.

The hippocampus is a key brain structure for memory, learning, and cognition. It has been implicated in many aspects of the neuroendocrine responses during stress [8]. Lesion on the hippocampus could lead to the impaired persistent memory which was considered as the usual consequence of posterior cerebral artery ischemia. Moreover, stroke lesions in the hippocampus may become even more relevant in the presence of a second pathology like Alzheimer dementia [9]. Therefore, the hippocampus function may involve in the ischemia-induced neurologic deficit.

The Voltage-gated ion channels are responsible for maintaining neuronal homeostasis and function [10-12]. Consequently, while the influx of  $\text{Na}^+$  and/or  $\text{Ca}^{2+}$  ions modulate neuronal excitability, the sudden influx of these ions may also trigger apoptosis or give rise to cytotoxic edema [13]. Voltage-gated ion channels underlie the cell membrane conductance of  $\text{Ca}^{2+}$ ,  $\text{K}^+$ , and  $\text{Na}^+$  ions and directly prorogate, inhibit, or modify electrical signals. Since these channels are responsible for ion influx or efflux, it is evident that these are the channels that are key to neuronal action potential generation [10]. Ischemic brain damage, like TIA or RIND, may activate signal transduction cascades that result in changes in gene expression, including the genes responsible for encoding ion channels. The alteration of ion channel gene expression may affect neuronal function and could also result in stroke-like behavior. Recently, voltage-gated ion channel modulators proved as promising candidates for the treatment of stroke [14]. Studies such as these demonstrate the importance of

obtaining molecular mechanistic information about these disorders in order to apply it to translational channelopathy with the goal of identifying new therapeutic strategies for neuronal protection and repair of damage caused by TIA and RIND.

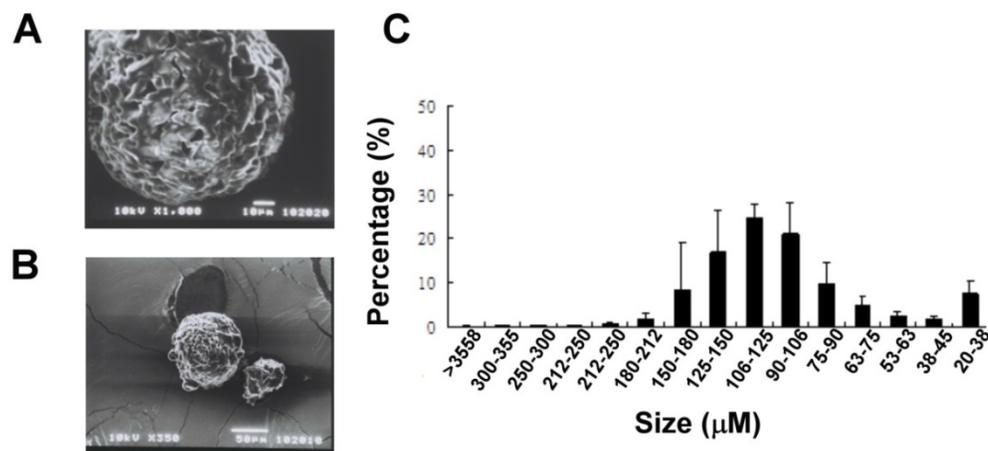
In this study, we used a novel animal model by using temperature-sensitive melted solid lipid microparticles to induce reversible stroke in order to mimic clinical TIA and RIND conditions, as we published previously [15]. Moreover, we utilized qPCR array analysis to understand how voltage-gated ion channel-encoded gene regulation plays a critical role in TIA and RIND. Understanding the role of these channels in stroke-associated pathologies is important for further new therapeutic strategy development.

## Materials and Methods

### Characterization of the melted solid lipid microparticles

The solid lipid microparticles were produced by emulsifying the oil phase, prepared from a mixture of different melting point of Gelucire, and the water phase, containing hydrophilic surfactant. We mixed Gelucire 33/01 and Gelucire 43/01 at a ratio of 9:1 to form solid lipid microparticles with a melting point of 34 and hydrophilic balances of 1 (34/01).

Gradation sieving of the melted solid lipid microparticles yielded 15 size fractions ranging from 20  $\mu\text{m}$  to >355  $\mu\text{m}$  (Fig. 1A). The distribution of the amount of each size fraction was showed; moreover half of the particles produced by our method were of sizes between 90 and 180  $\mu\text{m}$ . The blended Gelucire (34/01) solid lipid microparticles had uniform round shapes with a porous surface represented by SEM analysis (Fig 1B).



**Figure 1.** Physical characteristics of the lipid microparticles. (A) Representative SEM analysis showed the surface morphology of the particles. (Upper panel, bar = 10  $\mu\text{m}$ ; lower panel, bar = 50  $\mu\text{m}$ ) (C) Distribution of the sizes of lipid microparticles. Particles size was classified using different sizes of mesh. The distribution tends to have a skewed type (n = 3).

## Lipid particle preparation

The detailed procedure to prepare lipid microparticles has been described previously [16]. The oil phase was prepared by mixing Gelucire 43/01 and Gelucire 33/01. A ratio of 1:9 of Gelucire 43/01 and Gelucire 33/01 was prepared for the blending of solid lipid microparticles (a melting point of about 34°C was estimated by using a melting point apparatus Mel-Temp II, Laboratory Devices, INC, USA). To obtain the blends, the oil phase was homogeneously mixed by heating to a temperature that was over the melting point of the Gelucire lipid.

The water phase was prepared by adding sodium dodecyl sulfate in purified water to get a final concentration of 1%. Both the oil and water phase were heated up to 70°C. The oil phase (3 ml) was kept at 70°C and was mixed with the water phase (300 ml) by slowly dropping the oil into the water phase while stirring at a speed of 300 rpm for 5 min. The water phase was then slowly brought down to room temperature. Gelled lipid microparticles were then allowed to harden in the cool water phase while stirring at a speed of 300 rpm for 24 h. After hardening, the microparticles were filtered, rinsed with deionized water and air dried overnight before sieving the particles using standard U.S. size meshes (from 40 to 400 mesh). Finished microparticles were kept at 4°C and final particles sizes ranging from 75 to 125 µm were used as emboli for inducing transient brain ischemia.

## Scanning electron microscopy (SEM)

Lipid microparticles were attached onto double-sided adhesive tape, fixed to a stage, and coated with gold so that the size and morphology of the microparticles could be observed and photographed using JEOL JSM-5300 scanning electron microscopy (Tokyo, Japan).

## Animal model and surgical preparation

All animals of experimental protocols were reviewed and approved by the Institutional Animal Care and Use Committee of Kaohsiung Medical University. Three-month-old male Wistar rats weighing 300 -350 g were used for this study. Male rats were housed in a well-controlled animal care facility under diurnal lighting cycles, and given free access to food and water.

All animals were food-restricted overnight and were anesthetized the next day by an intra-peritoneal injection of 300 mg/kg chloral hydrate. Body temperature was monitored by a rectal thermometer and kept at 37°C by an automated temperature regulator system. The rat was then fixed in the supine position on an operation plate and a midline excision

of the ventral neck was used to explore the bifurcation of the right carotid artery. A PE-10 catheter (Becton Dickinson, NJ, USA) was inserted through the excised external carotid artery at the bifurcation of the common carotid artery concomitantly with ligation of the ipsilateral four collateral cerebral arteries. The PE-10 catheter was preserved by being carefully fixed to the external carotid artery and exposed on the neck as an indwelling injection tube after the closure of the surgical wound; heparin was used for preventing clot on the PE-10 tubing. In order to prevent infections, surgical wounds were carefully cleared, and all rats received postoperative neurologic evaluations. If any focal neurologic deficits were found, rats were excluded from further studies.

The stroke model induced by melted solid lipid in conscious animals was established in our previous study [15]. Briefly, we injected solid lipid microparticles within the range of 75 -125 µm via the indwelling catheter tube, injections were made into the ipsilateral bifurcation of the common carotid artery without anesthesia. Neurologic deficit was evaluated immediately after injecting solid lipid microparticles, and the total duration of neurologic deficit induced by transient ischemia was recorded.

In order to induce RIND, we injected solid lipid microparticles 12 h after recovery from the transiently induced ischemic episode using the same process described above. Neurological deficit was again evaluated immediately after injecting the solid lipid microparticles until the symptoms of the stroke disappeared.

Following the transient ischemic episode, TTC staining and quantitative real-time PCR (qPCR) were used to measure the infarct volume in the brain.

## Neurologic deficit evaluation

All rats received neurological testing every 2 min in the first hour, then at 1.5, 2, 2.5, 3, 3.5, 4, and 24 h after operation. The neurologic deficits were scored as 0 (no neurologic defects), 1 (one paw clumsiness), 2 (tilt), 3 (rounding in only a unilateral circle), 4 (akinesia), 5 (seizure), 6 (absence of any spontaneous movement), and 7 (death). All neurologic behaviors were evaluated by the same investigator.

## Calculation of infarction volume

After neurological evaluations, rats were deeply anesthetized, and brains were removed and then positioned in a rat brain matrix (Activational Systems, Ann Arbor, MI). In cases where rats died prior to the end of evaluations, brains were removed within 30 min of death. Each brain was cut into twelve 2-mm-thick coronal sections. We then used TTC staining to measure infarct size by incubating sections

in a 0.05% TTC solution for 30 min at 30°C before being fixed in a 4% buffered formaldehyde solution for 24 h.

### MicroPET study

A high-resolution PET scanner (Concord Microsystems, Knoxville, TN) was used for brain imaging according to previous study [17]. Briefly, rats were anesthetized with isoflurane, administered 37 MBq  $^{18}\text{F}$ -FDG via the tail vein, and were allowed 30 min for uptake. Within 10 min of  $^{18}\text{F}$ -FDG administration, images were acquired in the PET scanner and reconstructed with a 3D ordered subset expectation-maximization (OSEM) iterative algorithm. In transverse brain sections, regions of interest (ROIs) in the lesion area were identified by changes in metabolism induced by artery occlusion. Following calibration of both ipsi- and contralateral insulated areas, the uptake of  $^{18}\text{F}$ -FDG in the ROIs was calculated as the average nCi/cc. Radioactivity in the contralateral area was used as a reference to normalize data obtained in the ipsilateral area of the same image.

### RNA isolation and cDNA synthesis

Total RNA from the rat hippocampus was extracted using the TRIzol plus RNA purification kit (Invitrogen Life Technology, Carlsbad, CA), and was converted to cDNA using the SuperScript® III first-strand synthesis system (Invitrogen Life Technology, Carlsbad, CA).

### Quantitative real-time PCR array

A customized qPCR Array (Invitrogen Life Technology, Carlsbad, CA) was performed according to the manufacturer's protocol to detect voltage-gated ion channels expressions. This array was developed to profile the expression of a panel of 96 genes encoding brain-related ion channels and transporters. The genes represented on the array were grouped according to their functional and structural features, and included  $\text{Ca}^{2+}$ ,  $\text{K}^{+}$ ,  $\text{Na}^{+}$ , and  $\text{Cl}^{-}$  channels. Expression of mRNA in the rat hippocampus was then quantified by real-time RT-PCR using Applied Biosystems 7500 Real-Time PCR System (Applied Biosystems) to verify the mRNA targets.

### Statistics

Statistical analysis was performed using a one-way ANOVA followed by Bonferroni's post-hoc comparison test and a student t-test. A *p* value of < 0.05 was considered statistically significant.

## Results

### Characterization of lipid microparticles

A SEM analysis was performed to reveal the

morphology (Fig 1A) and size of the lipid microparticles. The size of the particles was determined to be around 100  $\mu\text{m}$  (Fig 1B).

### Neurological and histology studies

Rats that received the microparticle injections successfully displayed neurological deficits (Fig 2A). As shown in Fig 2A, TIA was considered to be induced when behavioral deficits (neurological score >1) lasted from a few minutes to 24 h. Importantly, neurological behavior deficits lasting from 24 to 72 h were categorized as RIND. As showed in Fig 2A, rats received the microparticles injection again after we inducing rats TIA successfully, the neurological behavior deficits of rats lasted more than 24h, and were categorized as RIND. The infarction area during RIND was then determined by TTC staining. However, no obvious region was demarcated suggesting that there was no significant neuronal death during RIND (Fig 2B). The irreversible stroke rats displayed as a positive control for the TTC staining.

### MicroPET studies

PET has been widely used in clinical diagnostics and treatment monitoring. Normal brain tissue has prominent uptake of  $^{18}\text{F}$ -FDG due to the constant need for glucose in the brain. Using the contralateral side as baseline, RIND-induced metabolic changes over the ipsilateral side of the rat brain were detected by PET scan after RIND (Fig. 2C). Radioactive quantification of the ROIs in transverse coronal brain sections indicated no significant changes in glucose uptake in the ipsi- and contralateral areas of the hippocampus (Fig. 2D).

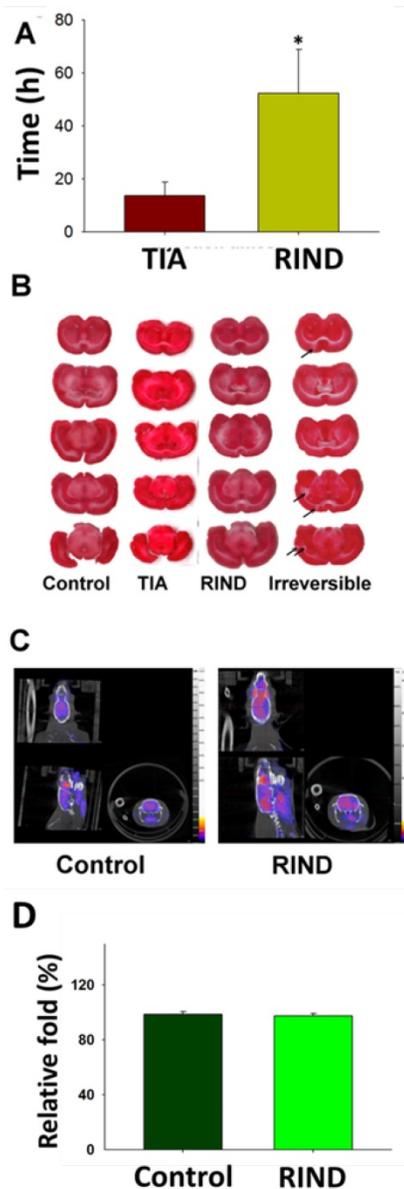
### Q-PCR array analysis

To examine the transcriptional response to TIA or RIND, we performed a qPCR array to compare the mRNA profiles between the hippocampus in control, TIA, and RIND rats. The genes that encode voltage-gated ion channels that were included in the custom array are listed in supplemental table 1. Expression of voltage-gated ion channel mRNA was considered significantly different when values for all samples from the hippocampus of experimental rats were more than double those for the controls ( $n=3$  for each subgroup). The hierarchical cluster analysis of all significant mRNAs is shown in Fig 3A, which illustrates mRNAs differentially expressed in the hippocampus during TIA and RIND, compared to control conditions. Interestingly, the voltage-gated potassium channels, *Kcnj4*, were down-regulated in both TIA and RIND, as shown in Fig 3B. However, *Cang3* and *Kcnq1* gene expression decreased during

RIND, but not in the TIA condition. The up-regulated mRNA targets that were more than those of the control, including significant changes in *Scn7a* and *Kcna7*, are shown in Fig 3B. Moreover, the genes exhibiting a greater changes during RIND compared to TIA were *kcna5*, *kcna6*, *kcnj13*, *kcnk16*, and the sodium channel modulatory  $\beta$ -subunit, *scn4b*.

### Statistical analysis

All results were presented as mean  $\pm$  standard error of the mean, and the Student *t* test was used to evaluate inter-group differences. A *p*-value of  $<0.05$  was considered statistically significant.



**Figure 2.** (A) The duration of neurologic behaviors was repeatedly evaluated after injection of the particles. (B) The infarction area of rat brains after TIA, RIND, and irreversible ischemic damage was determined by TTC staining. Arrows indicate the infarction. (C) The metabolic changes in the brains after RIND were detected by PET scan. (D) The experiments were repeated three times and similar results were observed. All *p*-values are represented as ( $p < 0.05$ ).

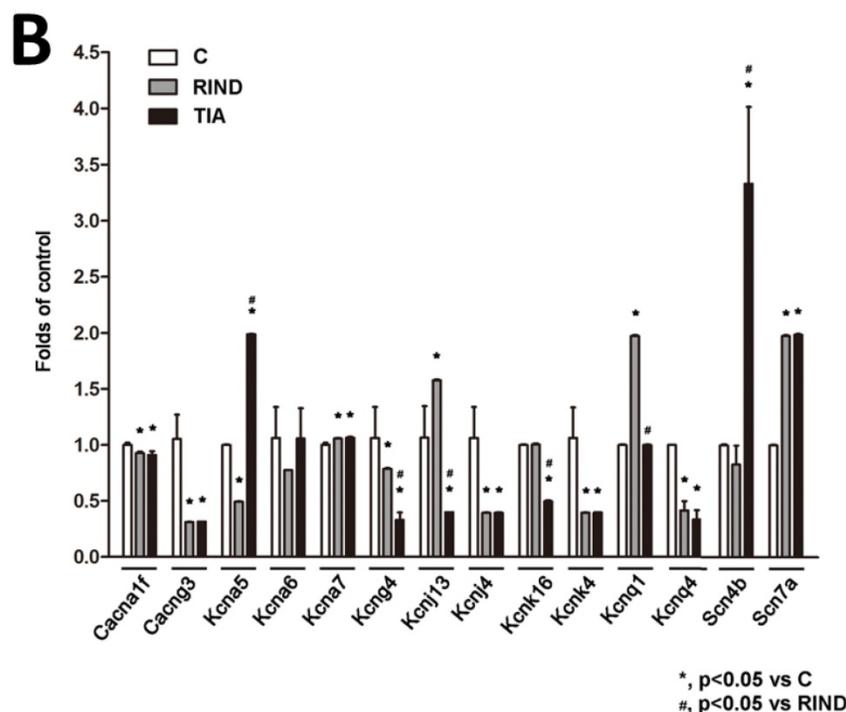
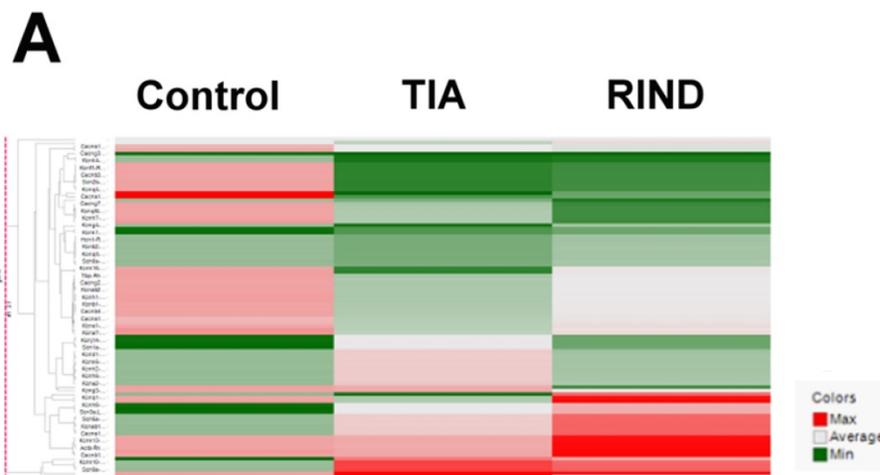
### Discussion

TIA and RIND are transient episodes of neurological dysfunction caused by ischemia. These types of neurological abnormalities are reversible and recover in less than 72 h. However, suitable animal models of TIA and RIND are not currently available. Most of the currently available animal models have been created by inducing irreversible-ischemic damage and show obvious infarctions in the brain. In our previous study, we used artificial microparticles to successfully establish an animal model of embolic stroke, including a novel model that mainly expressed lacunar infarction [16]. Therefore, in our current study we used this technique with the aim of creating a more suitable animal model of TIA and RIND. Interestingly, while stroke-related neurological dysfunctions were observed in our TIA and RIND animal models, we obtained negative results in TTC-staining (Fig 2A). These results suggest that there was only minor cell death during our microparticle-induced TIA or RIND indicating that our model might be more suitable for studying reversible ischemic stroke.

During an ischemic episode, neurons exhibit a transient, early rise in intracellular  $Ca^{2+}$  and modulate calcium-dependent gene transcription.[18, 19] Calcium-response factor or sodium-calcium exchanger isoform 1 may responsible for this calcium-dependent gene transcription, and may be play a relevant role under pathophysiological conditions of stroke by regulating intraneuronal  $Ca^{2+}$  and  $Na^+$  homeostasis [20, 21]. In this study, we identified a novel group of genes that encode proteins that underlie the transmission of nerve impulses. We also expanded the analysis to profile the expression of all 90 genes listed as putative neuronal targets of stroke-related neurological dysfunction in the hippocampus. Consistent with earlier reports, we found that the expression of 14 of the 90 previously reported genes were statistically different, in the current study. These results may implicate the involvement of voltage-gated ion channel transcriptional activity in hippocampal pathologies, such as epilepsy. Indeed, previous studies have shown that epilepsy can arise because of stroke [2, 3]. Interestingly, a vast number of channelopathies, related to recurrent seizures have been associated with increased expression of genes responsible for encoding a component of voltage-gated ion channels, as well as the increased expression of many other genes that directly or indirectly control brain development, neuronal function, and neuronal activity [1]. Therefore, alterations in the expression of these genes may contribute to the pathophysiology of recurrent seizures.

Accumulating evidence suggests that voltage-gated ion channels are either up or down-regulated in ischemic stroke and that these alterations may play a direct role in the modulation of sodium or potassium-mediated neuronal function [11]. Our qPCR array analysis demonstrated that *Scn7a*, which encodes an atypical voltage-gated sodium channel, up-regulated during TIA and RIND. This is in line with previous findings of increased expression of *Scn7a* in the human and rat hippocampus during epileptogenic processes in temporal lobe epilepsy. The *Scn7a* gene, which encodes  $Na_v2.x$ , has also been shown to be persistently up-regulated during a chronic epileptic phase in neurons of the

hippocampus [22, 23]. The  $\beta$ -subunit of sodium channels functions as a modulator to control channel ( $\alpha$ -subunit) gating. One previous study showed that functional deletion of the  $\beta$ -subunit in sodium channels resulted in Dravet Syndrome, a severe and intractable pediatric epileptic encephalopathy [24]. Therefore, the up-regulation of *Scn7a* may result in contralateral paralysis, sudden weakness or numbness, and/or epilepsy during TIA and RIND. And down-regulation of sodium channel modulatory  $\beta$ -subunit, *Scn4b*, may be an important transition point from TIA to RIND.



**Figure 3.** (A) Hierarchical cluster analysis of significant expression of rat hippocampal voltage-gated ion channel mRNA in control, TIA and RIND conditions. (B) Genes exhibiting greater change following TIA or RIND. The experiments were repeated in triplicate on one occasion. All *p*-values are represented as (*p* < 0.05).

In our qPCR array study, the calcium and potassium channel genes were also found to have altered expression patterns during TIA and RIND. Calcium overload may cause neuronal cell death during ischemic damage [25, 26]. Therefore, the reduction expression of the voltage-gated calcium channels *Cacna1f* and *Cacng3* in our study may also explain why we got negative TTC staining in TIA and RIND groups. Interestingly, a down-regulation of *Cacna1f* and *Cacng3* channel genes has been reported to be associated with childhood absence epilepsy [27, 28].

Along with sodium and calcium channels, we also found an alteration in potassium channel expression in our animal models of TIA and RIND. The potassium channels, *Kcna* and *Kcnj* encodes the delayed rectifier K<sup>+</sup> channels responsible for repolarization after an action potential [1, 29], and *Kcnj* and *Kcnq* encode inward rectifier K<sup>+</sup> channel which play a role in controlling neuron cell membrane excitability [30]. Functional studies of the physiological role of these voltage-gated potassium channels in healthy cerebral vessels show that they contribute to vascular tone by regulating resting membrane potential, and limiting depolarization and Ca<sup>2+</sup> influx by promoting K<sup>+</sup> entry into the cells. *Kv1.5* (encoded by *Kcna5*) has been reported to be down-regulated in the seizure-sensitive gerbil hippocampus [29]. Moreover, a recent study has suggested that the down-regulation of *Kir2.3* channel expression, which is encoded by *Kcnj4*, may contribute to the pathogenesis of temporal lobe epilepsy [31]. Therefore, a disturbance of these voltage-gated channels could be a contributing factor to complex cases of neurological disease and disorders and may specifically result in seizures, a disruption in autonomic control, and can even lead to sudden death [1, 32].

Recently, some studies have suggested that the transcription factors, *Pyk2* and *REST*, are activated following transient brain ischemia and reperfusion. These studies propose that these factors might play an important role in mediating post-ischemic signal transduction [33, 34]. Therefore, *Pyk2* or *REST*-dependent remodeling may also modulate channel gene expression, such as those of *Scn7a*, *Kcna* or *Kcnq* in the hippocampus, leading to increased neuronal excitability and epileptiform activity during TIA or RIND. The mechanisms underlying the modulation of the expression of these genes still needs further investigation.

In conclusion, we performed a novel animal model of TIA and RIND, which resulted in a transient neurologic deficit without cell death in the brain. This animal model has indicated that specific ion channels may serve as potential biomarkers in neurological

disorders such as stroke. Additionally, the data collected from qPCR screening, suggest that the voltage-gated sodium channels, *SCN7A*; the sodium channel regulatory subunit, *SCN4B*; the voltage-gated calcium channels, *Cacna1f*, *Cacng3*; the voltage-gated potassium channels, *Kcna5*, *Kcna7*, *Kcng4*, *Kcnj13*, *Kcnj4*, *Kcnk4*, *Kcnq1* and *Kcnq4*, might be important molecular targets for brain repair after transient ischemic stroke. Profiling the array of genes that are dysregulated in response to neuronal disease may advance our understanding of the mechanisms underlying the pathophysiology of these disorders. Furthermore, it may lead to the identification of novel therapeutic strategies in TIA and RIND.

## Supplementary Material

Supplementary information.

<http://www.medsci.org/v16p0060s1.zip>

## Acknowledgements

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## Competing Interests

The authors have declared that no competing interest exists.

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