

Research Paper

Tanshinone IIA Inhibits β -Catenin Nuclear Translocation and IGF-2R Activation via Estrogen Receptors to Suppress Angiotensin II-Induced H9c2 Cardiomyoblast Cell Apoptosis

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Abstract

Cardiomyopathy involves changes in the myocardial ultra-structure, hypertrophy, apoptosis, fibrosis and inflammation. Angiotensin II (AngII) stimulates the expression of insulin like-growth factors (IGF-2) and IGF-2 receptor (IGF-2R) in H9c2 cardiomyoblasts and subsequently leads to apoptosis. Estrogen receptors protect cardiomyocytes from apoptosis and fibrosis. Tanshinone IIA (TSN), a main active ingredient from Danshen, has been shown to protect cardiomyocytes from death caused by different stress signals. Estrogen receptor α (ER) is required for the rapid activation of the IGF-1R signaling cascade. This study aimed to investigate whether TSN protected H9c2 cardiomyocytes from AngII-induced activation of IGF-2R pathway and hypertrophy via ERs. We found that AngII caused the reduction in IGF-1R phosphorylation and the elevation of β -catenin and IGF-2R levels. This was reversed by increasing doses of TSN and of caspase-3 and ERK1/2 phosphorylation mediated by ERs. The phytoestrogen significantly attenuated AngII-induced apoptosis and suppressed the subsequent cardiac remodeling effect. Therefore, TSN reduced the AngII-induced activation of β -catenin and IGF-2R pathways, apoptosis and cardiac remodeling via ERs in H9c2 cardiomyoblasts.

Key words: Angiotensin II; β -Catenin; Estrogen receptors; H9c2 Cardiomyoblasts; Insulin-like Growth Factor-2 Receptor; Tanshinone IIA.

Introduction

Cardiac hypertrophy is a normal injury arising in different cardiovascular diseases, including hypertension, myocardial dead tissue, and inborn coronary illness, and is the main cause of heart failure and sudden death in patients with the above

maladies. The principal pathological changes in cardiovascular hypertrophy are introduced through the progression of numerous events, including myocardial interstitial cell hypertrophy, myocardial interstitial cell multiplication, and extracellular lattice

increment of myocardial cells, eventually leading to myocardial rebuilding. IGF-2 shares 47% homology with insulin [1, 2] and plays a role in mammalian postnatal and fetal growth [3-5]. However, its expression drops after birth and goes through a transition during the neonatal stage [4]. IGF-2 was found to behave as a rescuer in a brief coronary occlusion porcine model and in a sheep model with myocardial infarction [6]; however, it was later shown to cause Beckwith-Wiedemann syndrome, prenatal overgrowth, poly hydramnios and fetal and neonatal lethality [7]. IGF-2R binds IGF-2 on the cell surface and to mannose-6-phosphate (M6P)-tagged proteins in the trans-Golgi network [8]. It is a type I trans membrane receptor with a large extracellular domain, a relatively short intracellular tail and a trans membrane domain [9]. The binding of IGF-2 to IGF-2R causes the phosphorylation of phospholipase- β and subsequently apoptosis of H9c2 cardiomyoblasts [10]. In previous studies, AngII was shown to induce the expression of IGF-2 and IGF-2R in cardiomyocytes via MEK and JNK [11]. Subsequently, the binding of IGF-2 to IGF-2R leads to the activation of phosphatase 2B (PP2A or calcineurin), which dephosphorylates Bad and subsequently results in its translocation to the mitochondria [10]. This results in apoptosis of the cardiomyocytes via mitochondrial outer-membrane permeability. In cardiomyocytes, the expression β -catenin can be stimulated by various stress signals, such as endothelin-1, pressure overload and phenylephrine, which can stimulate β -catenin pathway [12]. Estrogen is a steroid hormone, existing in 3 different isoforms, estrone (E1), 17 β -estradiol (E2) and estriol (E3), among which E2 has the highest biological activity [13]. The hormone binds to its cell surface receptors, estrogen receptors α and β (ER α and ER β) [14]. E2 and ERs have been shown to protect cardiomyocytes from apoptosis and fibrosis [15]. Many phytoestrogens, such as genistein, tanshinone IIA and resveratrol, have been shown to exert a protective effect against cardiovascular diseases via the ERs [16-18].

TSN was discovered from *Salvia miltiorrhiza* Bunge (Danshen) and is used to treat cardiovascular disorders and hepatitis in Asia [19]. Phytochemicals from in Danshen, including TSN, exhibit protective effects including antibacterial activity, antioxidative activity, anti-inflammatory activity and anti-cytotoxicity and act as inhibitors of platelet aggregation. In different studies, TSN was shown to protect cardiomyocytes from cellular damages [20-22]. This study aimed to investigate the effects of TSN on estrogen receptors and determine whether it exerts a protective effect against AngII-induced activation of

β -catenin and IGF-2R, apoptosis and cardiac remodeling in cardiomyocytes.

Materials and Methods

Cell Culture

H9c2 cardiomyoblasts from American Type Culture Collection (ATCC) (Rockville, MD) were cultured in 10-cm dishes in DMEM (St. Louis, Sigma, MO) with 10% cosmic calf serum (CCS; HyClone, South Logan, UT) in humidified air (5% CO₂) at 37°C. The medium was replaced every 48 h. The cells were washed with 3 to 5 ml 1 \times Dulbecco's phosphate-buffered saline (1 \times PBS; Gibco, Auckland, New Zealand). The H9c2 cells were administered with AngII (St. Louis, Sigma, MO), ICI 182780 (ICI; ER inhibitor; Tocris Bioscience, Bristol, UK) or TSN (St. Louis, Sigma, MO) with appropriate concentrations.

Immunofluorescence Microscopy

H9c2 cells were seeded in 12-well plates (1 \times 10⁵ cells per well). Twenty-four hours after cell seeding, 10⁻⁸ M AngII was added, followed by 40 μ M TSN after 1 h and then 10⁻⁶ M ICI after another 2 h. Twenty-four hours after the administration of ICI, cells were fixed with 4% paraformaldehyde in 1 \times PBS at RT for 15 min. The cells were permeabilized with 0.1% Triton X-100 in 0.1% sodium citrate for 20 min. Non-specific binding of the fixed cells was blocked with 10% CCS at room temperature (RT) for 1 h and the cells were incubated with diluted primary antibody solutions in 1 \times PBS for 48 h at 4°C. Then, the cells were incubated with Alexa Fluor 488-conjugated donkey anti-goat IgG secondary antibody (Invitrogen Corp., Carlsbad, CA) in the dark for 1h at 37°C. Finally, the cells were stained with 1 μ g/ml DAPI for 5 min to detect the cell nuclei (blue staining). Fluorescence was visualized using a fluorescence microscope coupled with an image analysis system.

Nuclear and Cytoplasmic Fractionation

The cells were harvested in PBS and centrifuged at 1,000 g for 10 min at 4°C. After carefully aspirating the supernatant, the cells were resuspended in 200 μ L ice-cold BUFFER-I (10 mM Hepes (pH 8.0), 1.5 mM MgCl₂, 10 mM KCl, 1 mM dithiothreitol, and proteinase inhibitor cocktail (Roche Molecular Biochemicals)) and incubated for 15 min on ice to allow the cells to be lysed, followed by adding 20 μ L IGEPAL-CA630. After vigorously vortexing for 10 s and centrifuging at 12,000 g for 5 min at 4 °C, the supernatant (cytoplasmic fraction) was carefully aspirated and the pellet was resuspended with ice-cold BUFFER-II (20 mM Hepes (pH 8.0), 1.5 mM MgCl₂, 25% glycerol, 420 mM NaCl, 0.2 mM EDTA,

1 mM dithiothreitol and proteinase inhibitor cocktail (Roche Molecular Biochemicals)) and vigorously vortexed. After vortexing, the suspension was placed on ice for 30 min before centrifuging at 15,000 g for 15 min at 4°C. The supernatant (nuclear extracts) was stored in aliquots at -80°C. These samples contained the nuclear proteins. Protein concentration in each sample was determined by the Lowry assay, and Western blot analysis was conducted in order to determine the protein expression.

TUNEL Assay

H9c2 cells were seeded in 12-well plates (1×10^5 cells per well) containing DMEM(10% CCS) to 80-90% confluency and incubated for 24 h in humidified air with 5%CO₂ at 37°C. Twenty-four hours after cell seeding, 10^{-8} M AngII was added, followed by 40 μ M TSN after 1 h and then 10^{-6} M ICI after another 2 h. Twenty-four hours after the administration of ICI, each well was washed with 1×PBS (1 ml/well) and then incubated with the fixative (1 ml/well) (4% paraformaldehyde in 1×PBS) at RT for 1 h. The blocking buffer (3% H₂O₂ in 100% methanol) was added (1 ml/well) and the plate was incubated for 10 min at RT. The permeabilization solution (0.1% Triton X-100 in 0.1% sodium citrate) was added (0.5 mL/well) and the 12-well plate was placed on ice for 2 min without shaking. The diluted 10× TUNEL reagent (Roche Diagnostics GmbH, Mannheim,Germany) was added (200-250 μ L/well) and the plate was placed in a humidified incubator at 37 °C in the dark for 1 h for the reagent to react with cell nuclei. Diluted 10000× DAPI was added (200 μ L/well) and the plate was covered with tin foil and rested for 25 min. Finally, the cells were observed under fluorescence microscope. The number of TUNEL-positive cardiac myocytes and the number of apoptotic bodies were determined by counting 3×10^5 cardiac myocytes. All morphometric measurements were performed by at least two independent individuals in a blinded manner.

Western Blot Analysis

H9c2 cells were seeded in 10-cm culture plates (1×10^5 cells per well) containing DMEM (10% CCS) to 80-90% confluency and incubated for 24 h in humidified air with 5% CO₂ at 37°C. Twenty-four hours after cell seeding, 10^{-8} M AngII was added, followed by 10, 20 and 40 μ M TSN after 1 h and then 10^{-6} M ICI after another 2 h. Twenty-four hours after the administration of ICI, each plate was washed twice with 3-4 ml 1×PBS and the remaining fluid in each plate was discarded. Then, 100 μ L cell lysis buffer (50 mM Tris-base (pH 7.5), 0.5 M NaCl, 1 mM EDTA (pH 8.0), 1 mM β -mercaptoethanol, 1% NP-40, 1% glycerol and protease inhibitor cocktail tablets)

was added to lyse cells in each plate. The cells were scraped down and collected in appropriate 1.5-ml micro centrifuge tubes on ice, which were then vortexed three times, once every 10 min and centrifuged for 20 min at 12000 rpm at 4°C. The supernatants were transferred to another set of 1.5-ml micro centrifuge tubes. These were the total protein samples. The protein concentration in each sample was determined by the Lowry assay, and Western blot analysis was conducted in order to determine the protein expression.

Statistical Analysis

Each sample was analyzed based on the results from experiments that were repeated at least three times, and Sigma Plot 10.0 software and ANOVA was used to analyze the numerical data. The values were expressed as the means \pm SD and statistical significance was set at $P < 0.05$. The statistical data significant TSN treatment group compared to ANGLI.

Results

TSN Up-Regulated the Expression of Endogenous ERs and Survival Pathway while Reducing AngII-Induced β -Catenin and IGF-2 Pathways in H9c2 Cardiomyoblasts in a Dose-Dependent Manner

To determine whether TSN was able to increase the expression of ERs and survival proteins and reduce β -catenin and IGF-2 pathways under the influence of AngII in the H9c2 cells, effective doses of the AngII were tested. AngII significantly increased the levels of β -catenin, IGF-2R, LEF-1, NFAT-c3 p-GATA-4, cleaved caspase-3 protein and reduced the level of p-IGF-1R protein (Fig. 1A), the difference is statistically significant (Fig 1B). In previous publications from the PI lab, 10^{-7} M and 10^{-8} M doses of AngII were shown to induce significant increases in the apoptosis of H9c2 cells at the 24h time point. Therefore, the 10^{-8} M dose of AngII was chosen for subsequent experiments. The administration of AngII alone reduced ER α , ER β , IGF-1R and p-IGF-1R and elevated β -catenin, p-ERK1/2, IGF-2R and cleaved caspase-3, (Fig. 1C). The administration of 40 μ M TSN up-regulated the expression of ERs and cell survival pathway, and decreased AngII-induced β -catenin, p-ERK1/2, IGF-2R and cleaved caspase-3 in the H9c2 cells. Therefore, TSN was able to up-regulate ERs and IGF-1R/Akt survival pathway and reduce the AngII-induced activation of β -catenin and IGF-2 pathways in the H9c2 cardiomyoblasts in a dose-dependent manner and the difference is statistically significant showed Fig 1D.

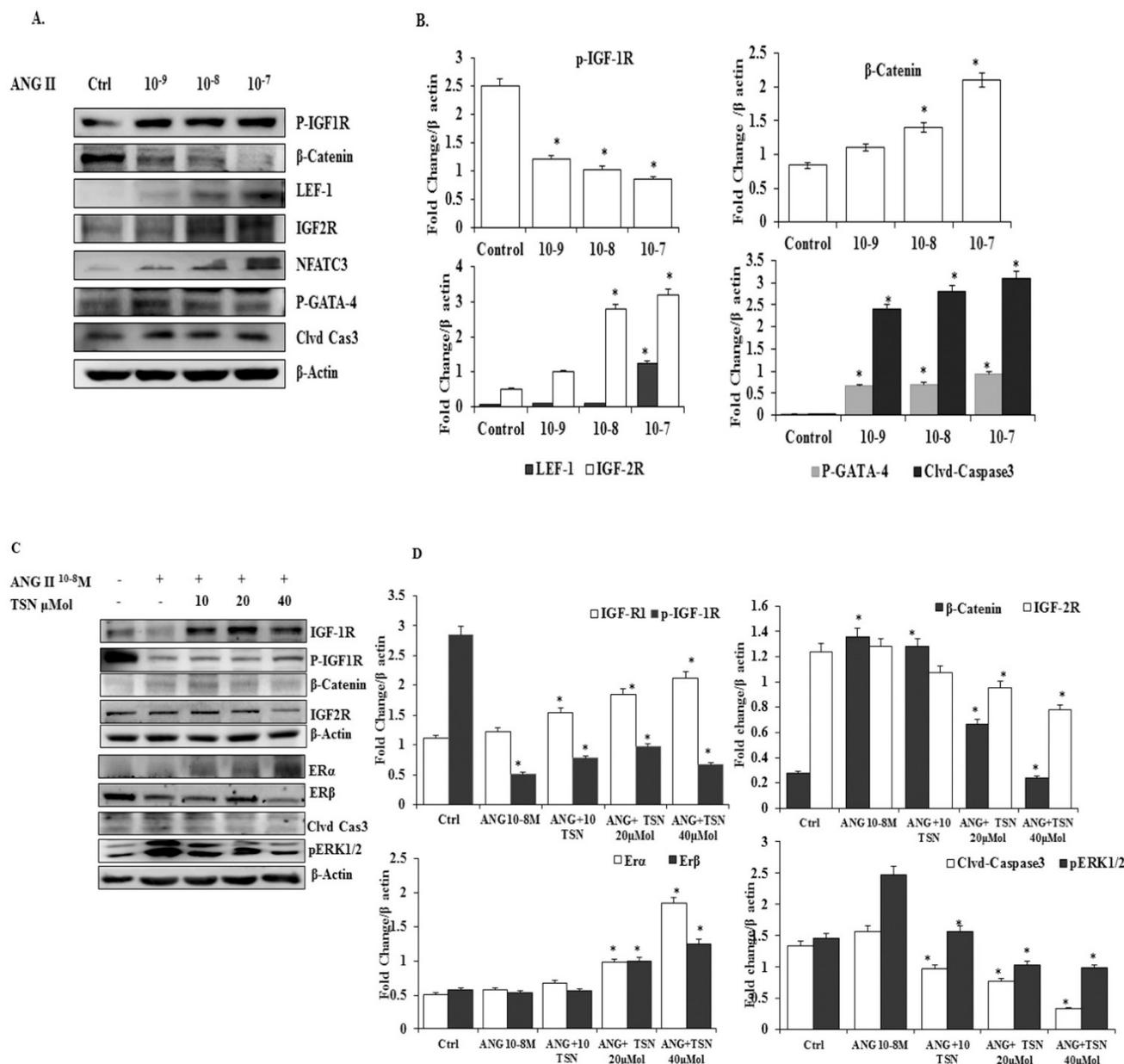


Figure 1. TSN suppressed the AngII-induced activation of β-catenin and IGF-2pathways and increased the expression of endogenous ERs and survival pathway in H9c2 cardiomyoblasts in a dose-dependent manner; all the experiments were repeated at least three times. (A) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, different doses of AngII were added. Western blot analysis was conducted 24 h after the administration of AngII. (B) The difference is statistically significant. (C) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10⁻⁸ M AngII was added, followed by different doses of TSN. Western blot analysis was conducted 24 h after the administration of TSN. (D) Difference is statistically significant. Data are expressed as the fold change relative to the control and presented as the mean ± SD, representing the results of three independent experiments (n = 3, *p < 0.05 was considered significant).

AngII Increased the Expression and Nuclear Localization of β-Catenin, which were reversed by TSN via the Activation of ERs

In different studies, AngII was found to activate the β-catenin pathway in a mouse model and in podocytes [23]. Hence, the possibility of induction of β-catenin by AngII in the H9c2 was determined first. Then, effect of TSN on the activity of the β-catenin pathway was investigated. AngII at 10⁻⁸ M alone increased β-catenin and LEF-1 and reduced ERα and

ERβ (Fig. 2A) and the difference is statistically significant (2B). In contrast, 40μM TSN decreased AngII-induced β-catenin and LEF-1 and increased ERα and ERβ, and this was reversed by 10⁻⁶M ICI. Moreover, 10⁻⁸ M AngII also led to an increase in the nuclear localization of β-catenin (Fig. 2C and 2E). However, this was down-regulated by the addition of 40μM TSN. Nevertheless, the addition of 10⁻⁶ M ICI restored nuclear level of β-catenin. The difference is statistically significant showed in figure 2D. Therefore, TSN was able to attenuate AngII-induced

activation of β -catenin pathway via ERs in the H9c2 cardiomyoblasts.

TSN Reduced AngII-Induced Apoptosis and Promoted Survival in H9c2 Cardiomyocytes

In other studies, AngII was found to cause apoptosis in cardiomyocytes [24]. TSN was found to protect the cardiomyocytes from apoptosis [25]. To investigate the effect of TSN on AngII-induced apoptosis, Western blot analysis was performed.

Administration of 10^{-8} M AngII alone resulted in the significant increase in the levels of apoptotic proteins, cleaved caspase-3, and a sharp reduction in survival pathway proteins (Fig. 3A) difference is statistically significant as showed in figure 3B. This was counteracted by the addition of $40 \mu\text{M}$ TSN. However, the anti-apoptotic effect of the phytoestrogen was reversed by the addition of 10^{-6} M ICI. Furthermore, AngII alone caused a significant increase in the total number of TUNEL-positive cells (Fig. 3C). TSN

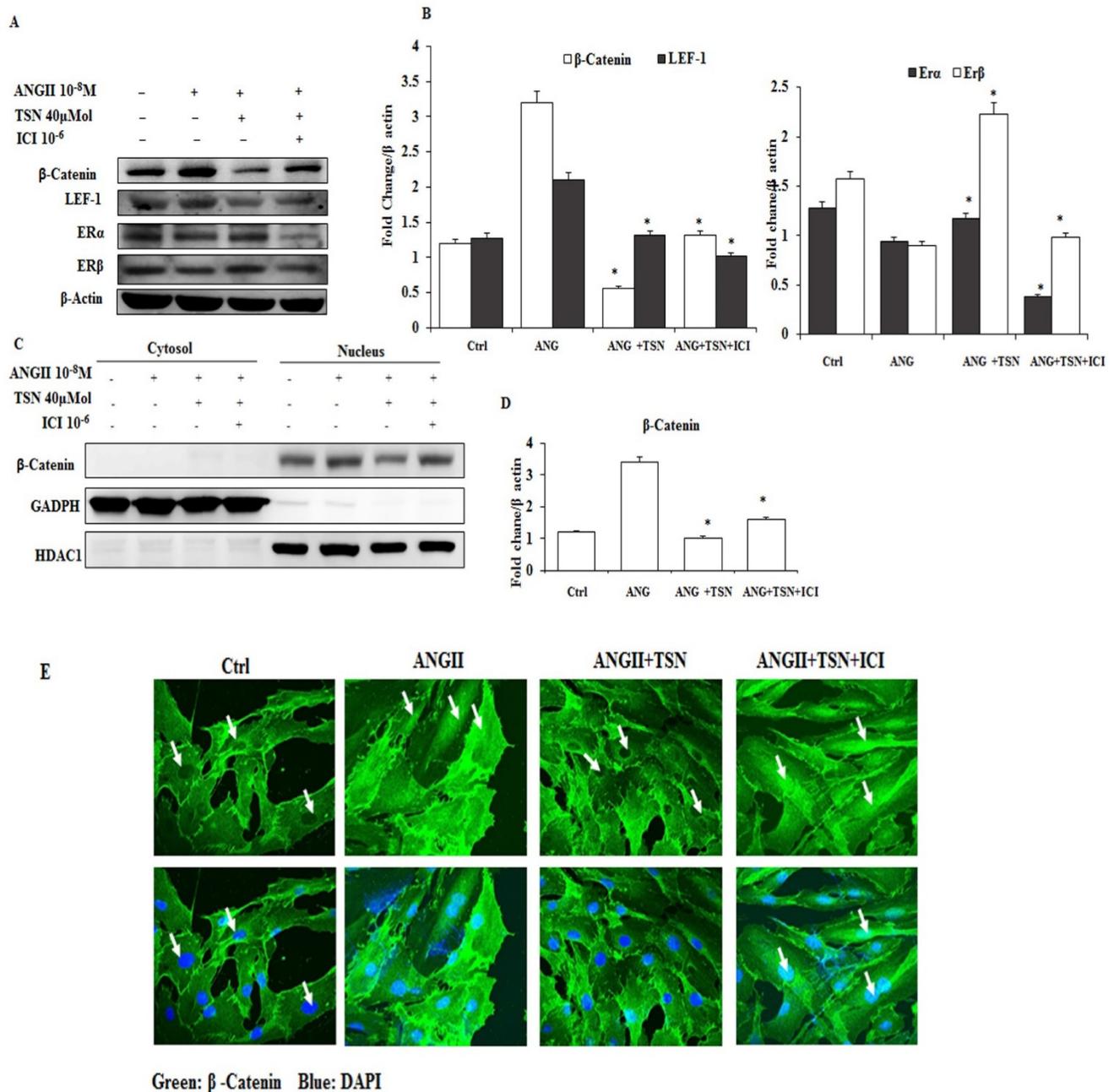


Figure 2. TSN reduced AngII-induced β -catenin expression and nuclear localization through ERs; all the experiments were repeated at least three times. (A) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10^{-8} M AngII was added, followed by $40\mu\text{M}$ TSN and 10^{-6} M ICI. Western blot analysis was conducted 24 h after the administration of ICI. (B) fold change were calculated and are shown. (C) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10^{-8} M AngII was added, followed by $40\mu\text{M}$ TSN and 10^{-6} M ICI. The nuclear and cytoplasmic fractions were separated 24 h after the administration of ICI. (D) Difference is statistically significant. (E) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10^{-8} M AngII was added, followed by $40\mu\text{M}$ TSN and 10^{-6} M ICI. Confocal microscopy was conducted 24 h after the administration of ICI. Data are expressed as the fold change relative to the control and presented as the mean \pm SD, representing the results of three independent experiments ($n = 3$, $*p < 0.05$ was considered significant).

treatment reduced the number of TUNEL-positive H9c2 cells and the addition of 10^{-6} M ICI reversed this effect. Therefore, TSN protected the H9c2 cells from AngII-induced apoptosis through ERs.

TSN Attenuated AngII-Induced Fibrosis in H9c2 Cardiomyoblasts via ERs

AngII has been shown to induce fibrosis [26]. To investigate effect of TSN on AngII-induced fibrosis, Western blot analysis was performed. AngII up-regulated the levels of MMP-9, MMP-2, TGF- β 1, p-Smad2/3, SP-1 and CTGF proteins and decreased the expression of TIMP-1 and TIMP-2 (Fig. 4A). TSN, on the other hand, reduced the AngII-induced protein expression of fibrosis markers at $40 \mu\text{M}$ and 10^{-6} M ICI reversed this situation. The difference is statistically significant showed in figure 4B.

Discussion

Cardiomyopathy involves the alteration of the myocardial ultra structure, apoptosis, cardiac fibrosis and modifications of cardiac gene expressions [27, 28]. AngII signals through AT1R, MEK and JNK to stimulate the expression of IGF-2 and IGF-2R and cause the subsequent activation of G α_q in H9c2 cardiomyoblasts, leading to apoptosis [11, 29]. E2 binds to its receptors, estrogen receptor α and β (ER α and ER β) [14]. E2 and ERs were shown to protect cardiomyocytes from apoptosis [30] and fibrosis [15]. Finally, phytochemicals found in Danshen, including TSN, display diverse protective effects such as antibacterial activity, antioxidative activity, anti-inflammatory activity, cytotoxicity and act as inhibitors of platelet aggregation. In previous studies, TSN has been shown to protect cardiomyocytes from

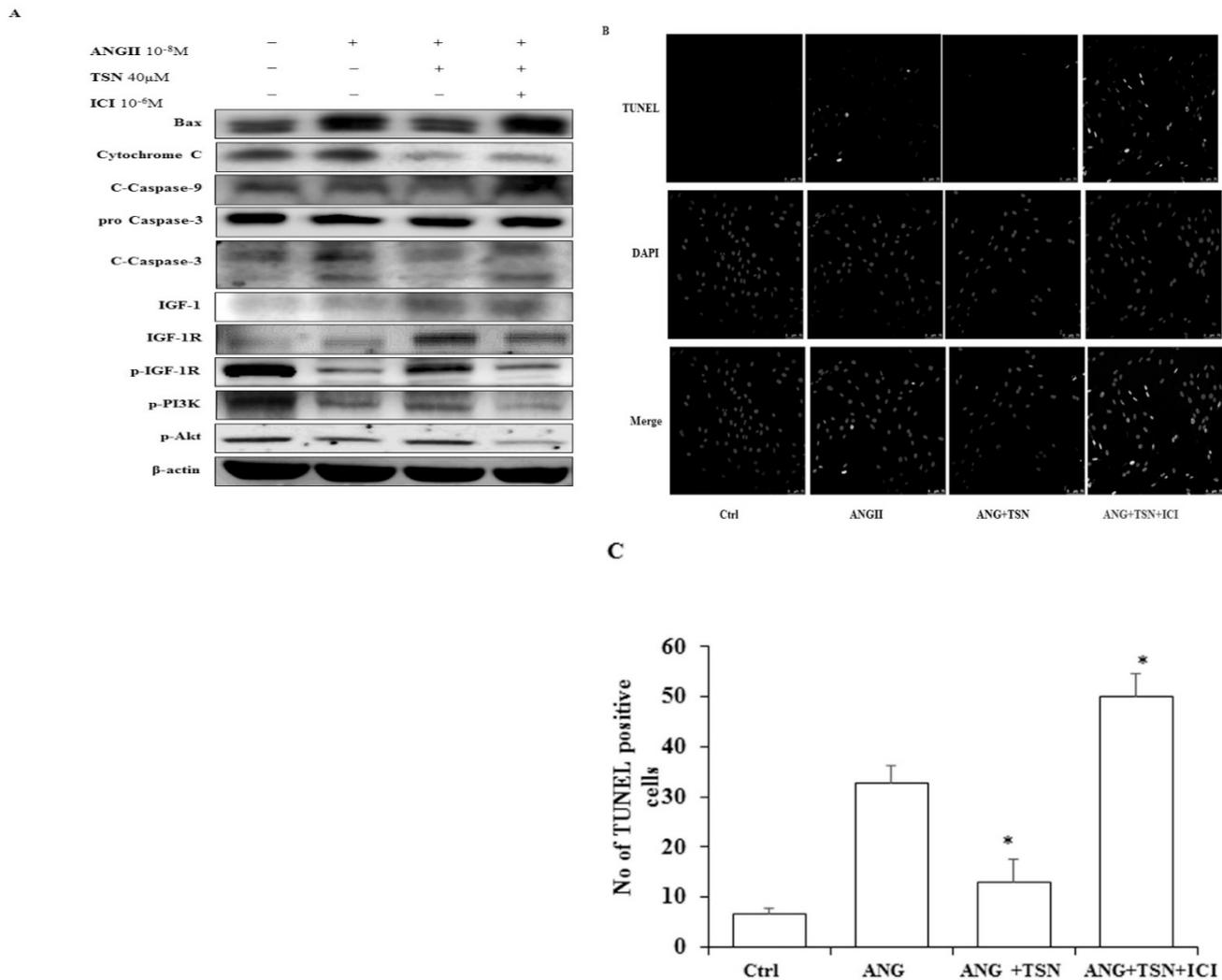


Figure 3. TSN reduced AngII-induced apoptosis and promoted survival via estrogen receptors in H9c2 cardiocytes. (A) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10^{-8} M AngII was added, followed by $40 \mu\text{M}$ TSN and 10^{-6} M ICI. Western blot analysis was conducted 24 h after the administration of ICI. (B) Fold change were calculated and are shown. (C) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10^{-8} M AngII was added, followed by $40 \mu\text{M}$ TSN and 10^{-6} M ICI. TUNEL staining was conducted 24 h after the administration of ICI (400X magnification). Data are expressed as the fold change relative to the control and presented as the mean \pm SD, representing the results of three independent experiments ($n = 3$, $*p < 0.05$ was considered significant).

death caused by different stress signals [18]. AngII was previously found to be capable of inducing the expression of IGF-2 and IGF-2R via MEK and JNK, resulting in the apoptosis of cardiomyocytes [11]. In this particular study, AngII was shown to reduce the activities of the cell survival pathway and ER signaling while promoting the expression of the β -catenin pathway and IGF-2R, ERK1/2 phosphorylation, NFAT-c3 activation and apoptosis in H9c2 cardiomyoblasts. However, the addition of TSN promoted IGF-1R phosphorylation and the activation of estrogen receptors and down-regulated the expression of β -catenin and IGF-2R and reduced the activation of caspase-3 and phosphorylation of ERK1/2. In different study, AngII was shown to not only trigger the expression of IGF-2 and IGF-2R [31] but also cause cardiac hypertrophy [32]. In addition, IGF-2R is also able of inducing cardiac hypertrophy [18]. In this study, AngII stimulated the activities of MAPKs (ERK1/2, JNK and p38) and the expression of IGF-2R and hypertrophic proteins. In other words, AngII caused an IGF-2R-induced hypertrophic response in H9c2 cardiomyoblasts. TSN significantly attenuated the phosphorylation of MAPKs and GATA-4 and reduced the protein levels of IGF-2R and hypertrophic markers. These effects were reversed by ICI, an inhibitor of the ERs. Therefore, the phytochemical TSN signaled through the ERs to lessen AngII-induced hypertrophy in H9c2 cells.

In previous studies, AngII was found to induce

the expression of β -catenin to cause cellular damages [33]. In the present study, the increase in the levels of β -catenin and LEF-1 proteins and the nuclear localization of β -catenin were results of the AngII treatment, which also caused a reduction in ER α and ER β . TSN, in contrast, exhibited inhibitory effects on β -catenin activation and restored the estrogen receptors in the H9c2 cells. In other words, TSN suppressed AngII-induced β -catenin signaling through ERs in H9c2 cardiomyoblasts. This phenomenon was observed in another study where TSN exerted a protective effect against AngII-induced damages in H9c2 cells through the activation of ERs [18]. In this study, AngII alone caused a significant increase in the levels of Bax, cytochrome c, caspase-9 and caspase-3 proteins in H9c2 cells. TSN not only decreased AngII-triggered apoptosis but also up-regulated the IGF-1 survival pathway. However, this event was blocked by the use of ICI. Similarly, TSN reduced Leu27 IGF-2-induced apoptosis in cardiomyocytes through the estrogen receptors [18]. In other studies, the activation of ERs was shown to down-regulated apoptosis in cardiomyocyte caused by different stress stimuli. AngII was previously shown to causes cardiac fibrosis [34]. Furthermore, AngII was also found to stimulate inflammation of the heart [35]. In this particular study, the administration of AngII alone resulted in an increase in the expressions of fibrosis markers and inflammation to some degree. However, these were reduced by TSN

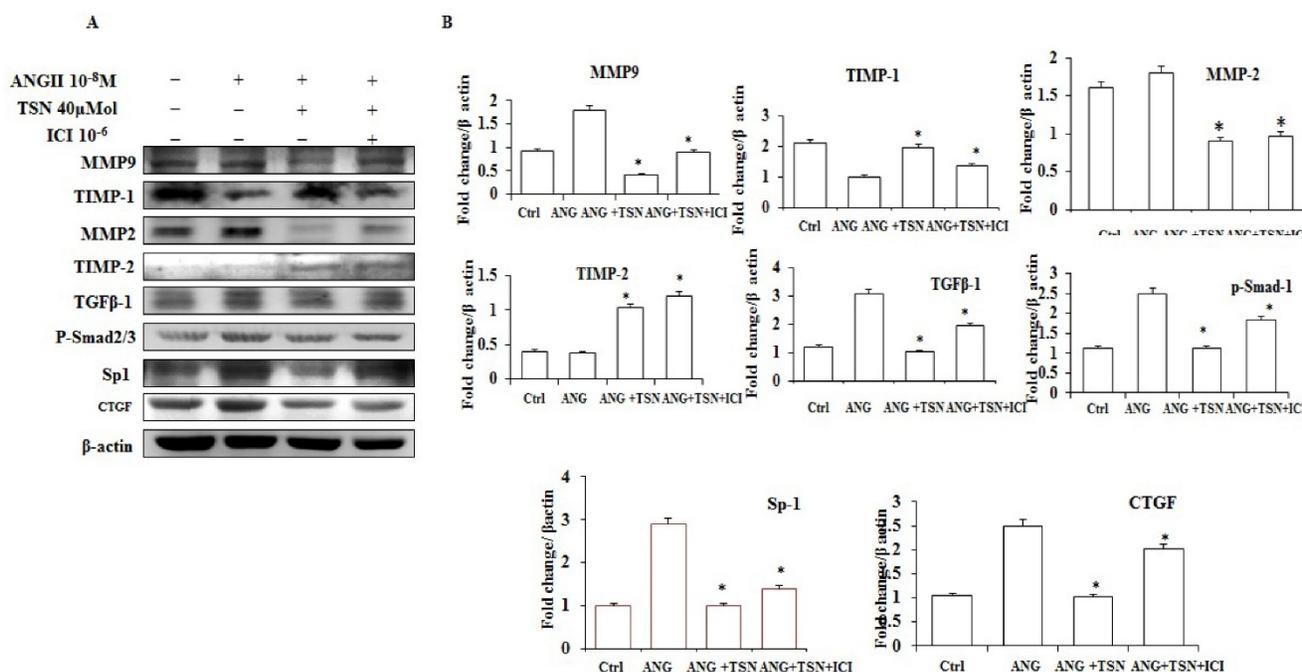


Figure 4. TSN reduced AngII-induced fibrosis via ERs in H9c2 cardiomyocytes. (A) Twenty-four hours after seeding the H9c2 cells in 10-cm culture plates, 10⁻⁸M AngII was added, followed by 40μM TSN and 10⁻⁶ M ICI. Western blot analysis was conducted 24 h after the administration of ICI. (B) Fold change were calculated and are shown.

and restored by ICI. Similarly, estrogen receptors were shown to prevent or suppress cardiac fibrosis [36] and inflammation [37].

In conclusion, AngII was found to significantly increase the expression and nuclear localization of β -catenin and to increase the expression of IGF-2R, apoptotic proteins and fibrosis while increasing the localization of the IGF-2R molecules to the cell membrane. These effects were sharply reduced by TSN via estrogen receptors, as they were activated by the phytoestrogen. Therefore, tanshinone IIA attenuates β -catenin and IGF-2R pathways and reduces subsequent apoptosis and remodeling while increasing survival proteins in H9c2 cardiomyoblasts.

Acknowledgments

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Competing Interests

The authors have declared that no competing interest exists.

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