

Research Paper

The inhibition of indoleamine 2, 3-dioxygenase 1 by connexin 43

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Abstract

Upregulation of connexin 43 (Cx43) showed potential in enhancing immune surveillance that was suppressed in the tumor microenvironment. The expression of indoleamine 2, 3-dioxygenase (IDO) is one of the crucial factors contributing to tumor immune tolerance by depletion of tryptophan and IDO-mediated tryptophan metabolites. Here, we aim to investigate the role of Cx43 in IDO production in murine tumor by using Cx43 inducers. Resveratrol (trans-3, 5, 4'-trihydroxystilbene) is a natural plant-derived polyphenol possessing positive effect against cancer. *Salmonella enterica* serovar choleraesuis (S.C.) was proved to target and inhibit tumor growth. Both of them regulated Cx43 expression in tumor cells and led to either chemosensitizing or immune-activating. In this study, the correlation between Cx43 and IDO were determined by the treatment of resveratrol and S.C. Our data showed an increase in Cx43 while IDO protein and IDO-mediated inhibited effects on T cell decreased after tumor cells are given with resveratrol and S.C. treatments. All of which could be inhibited once the expression of Cx43 was blocked. Cx43 involved in IDO regulation might be useful in developing IDO-targeted cancer immune therapy.

Key words: connexin 43, indoleamine 2, 3-dioxygenase 1, resveratrol, *Salmonella*

Introduction

The impaired ability of antigen presentation of cancer cells can be restored by enhancement of gap-junction-forming molecule, connexin 43 (Cx43) [1, 2]. Consisting two connexons and each composed of six connexins, gap junction is responsible for cell-cell molecule transferring and homeostasis maintenance [3]. Cx43, one of the widely-studied connexin isoform, is considered to have correlation with immunity, disease formation and progression [4, 5]. The lack of Cx43 induced by hypoxia renders melanoma more resistant to natural kill cell mediated lysis whereas its accumulation can restore the phenomenon by the formation of immune synapse and also involves

inactivation of T cells [6-8]. Indoleamine 2, 3-dioxygenase 1 (IDO) involves in one mechanism by which tumor cells avoid immune surveillance. As an enzyme initiating first and rate-limiting step of the tryptophan metabolic pathway, IDO mediates depletion of tryptophan and accumulation of tryptophan catabolites, thus leading to effector T cell apoptosis and activation of regulatory T cells (Treg) [9]. The inhibition of IDO production is able to activate host immunity [10, 11]. The combination of chemotherapy drugs and the IDO inhibitor were evaluated the beneficial effect in ongoing clinical studies, indicating IDO might be promising in

developing cancer immunotherapeutic treatment [12, 13].

In this study, we investigated the signaling pathway responsible for this process after two immune-activating agent, resveratrol and *Salmonella enterica* serovar choleraesuis (*S. Choleraesuis*, S.C.). Resveratrol (trans-3, 5, 4'-trihydroxystilbene) is a phytoalexin found in several plant species in response to environmental stress [14]. It was exerted in cancer prevention and inhibition contributes to antioxidant and anti-proliferative properties [14-16]. Moreover, several researches showed the beneficial effects of resveratrol on inhibition of cancer by inactivating tumor-induced tolerance and modulation of cytokine production [17-20].

On the other hand, bacteria were observed by a sarcoma surgeon to inhibit tumor growth in the 19th century [21]. As a facultative anaerobe with motility, *Salmonella* is more preferentially reaching, accumulating and proliferating in the tumor region than other strains or drugs [22-23]. *Salmonella* could inhibit the immune tolerance of tumor and activate the host immunity [7, 24, 25]. Herein, the treatment of S.C. or resveratrol in melanoma cells were demonstrated the ability of reducing IDO production through upregulating Cx43.

Materials and Methods

Bacteria, reagents and cell line

Vaccine strain *Salmonella enterica* serovar choleraesuis (*S. Choleraesuis*; S.C.) (ATCC 15480) was obtained from Bioresources Collection and Research Center (Hsinchu, Taiwan). Bacteria were maintained in L.B. plate and propagate in L.B. broth for using. Resveratrol (0-6 μ g/ml.), and DMSO were purchased from Sigma Aldrich (Sigma Aldrich, St. Louis, MO, USA). Murine B16F10 cells were maintained in culture dish with Dulbecco's Modified Eagle Medium (DMEM) containing 1% Penicillin-Streptomycin (100 units/mL penicillin and 100 μ g/mL streptomycin), 2mM glutamine and 10% heat-inactivated fetal bovine serum. Jurkat cell line (human T lymphocyte; (ATCC TIB-152™)) was a kind gift from Professor Wen-Wei Chang (Chung Shan Medical University) and maintained in HyClone RPMI 1640 medium containing 10% FBS. All cells were passaged every two to three days and incubated at 37°C, 5% CO₂.

Knockdown of Cx43

The specific shRNA of Cx43 plasmids were purchased from Santa Cruz Biotechnology (sc-35091-SH, Santa Cruz Bio-technology, Santa Cruz, CA, USA). B16F10 were transfected with Cx43 shRNA plasmids (5 μ g) by lipofectamine 2000 (Thermo

Scientific, Rockford, IL, USA) in Opti-MEM I reduced serum media (Thermo Scientific). Medium would be replaced with 10% DMEM after 16 h. The siRNA oligos of Cx43 designed to knockdown gene expression and the target sequences were listed below: Cx43 siRNA73 CCTGCTGATCCAGTGGTACA-TCTAT and Cx43 siRNA94 GCGTGAAGGGAAG-AAGCGATCCTTA. Lipofectamine RNAiMax reagent (Invitrogen, Carlsbad, CA, USA) was used for siRNA transfection following the manufacturer's protocol.

Analysis of Cx43 transcriptional activity

Cells grown in 24-well plates were cotransfected with luciferase reporter plasmids driven by Cx43 promoters (0.66 μ g) and pTCYLacZ (0.34 μ g), a β -galactosidase (β -gal) expression plasmid driven by the β -actin promoter, by lipofectamine 2000 (Invitrogen). At 6 h post-transfection, cells were treated with resveratrol or S.C. and cell lysates were harvested at different concentrations. The cell lysates were assessed for their luciferase activities determined by a dual-light luciferase and β -gal reporter gene assay system (Promega, Madison, WI, USA) using a luminometer (Minilumate LB9506, Bad Wildbad). Relative luciferase activity was measured as luciferase activity divided by β -gal activity to normalize transfection efficiency per microgram protein. The protein content in each sample was determined by the BCA protein assay (Pierce).

Western blot analysis

The protein content in each sample was determined by bicinchoninic acid (BCA) protein assay (Pierce Biotechnology, Rockford, IL, USA). Quantified each sample concentration to 60-80 μ g and add 4 \times SDS sample dye and then sample were denatured for 10 min at 95°C. Proteins were fractionated on SDS-PAGE, transferred onto Hybond enhanced chemiluminescence nitrocellulose membranes (Amersham, Little Chalfont, UK) and detected with antibodies against IDO (Thermo Scientific), Cx43 (Sigma-Aldrich) and β -actin (Sigma-Aldrich). Rabbit anti-mouse IgG-peroxidase antibody (Sigma-Aldrich) and goat anti-rabbit IgG-peroxidase antibody (Sigma-Aldrich) were used as the secondary antibody and protein-antibody complexes were visualized by enhanced chemiluminescence system (Amersham). The signals were quantified with ImageJ software (rsbweb.nih.gov/ij).

IDO functional assay

All cells incubations were at 37°C, 5% CO₂. B16F10 cells were plated in 12 well waited for attachment, and incubated with either different the multiplicity of infection (MOI) of *Salmonella* for 1.5 h

or various concentrations of resveratrol for 24 h. The medium of cells treated with *Salmonella* would be replaced with gentamicin-contained medium after bacteria incubation for 24 h. 160 µl of supernatants were harvested with 10 µl 30% TCA (Sigma-Aldrich) at 50°C, 30 min and then centrifuged mixture at 3000 rpm for 20 min at RT. The 125 µl mixture were mixed in an equal amount of 125 µl Erchlich's reagent (Sigma-Aldrich) in acetic acid in 96-well plates for 10 min. The absorbances of kynurenine were detected by spectrophotometer at a wavelength of A490. The data were performed as percentage of control. All of the IDO functional assays were done in quadruplicate.

Cell viability assay

Tumor cells were plated in 6 well culture plates and treated with various MOI of *Salmonella* for 1.5 h or various concentrations of resveratrol for 24 h after cells had been attached. The supernatants of *Salmonella*-treated B16F10 or 4T1 were added to Jurkat cells mixed with equal amount of RPMI medium. After 3 days, cell survival was assessed using the trypan blue exclusion assay. All of the viability assays were done in quadruplicate or sextuplicate.

Statistical analysis

All data were expressed as mean ± standard deviation (SD). The unpaired, two-tailed Student's *t* test was used to determine differences between groups. Any *P* value less than 0.05 is regarded statistically significant.

Results

The correlation between Cx43 and IDO

Cx43 has been demonstrated as a tumor suppressor, and interact with various molecules [26]. This study aims to identify the relationship between Cx43 and IDO in tumor cells. First, we used Western blotting to measure the expression of Cx43 and IDO in a variety of murine tumor cells (Fig 1A). The expression of Cx43 in CT26 (murine colon cancer) and 4T1 (murine breast cancer) cells was higher than that in B16F10 (murine melanoma) cells, but the expression of IDO in CT26 and 4T1 cells was not significantly different between that in B16F10 cells. The expression of Cx43 in B16F10 cells was lower than that in K1735 (murine melanoma) cells. The K1735 cell with higher Cx43 expressed lower IDO relatively. The B16F10 cells with the lowest Cx43 and higher IDO expression was chosen in further experiments. The correlation between Cx43 and IDO were identified in Cx43-knockdown B16F10 cell lines. Both Cx43 knockdown B16F10 cell lines #2 and #3 showed higher IDO expression (Fig 1B). To confirm the results,

IDO-mediated tryptophan metabolites, kynurenine was evaluated. The accumulation of kynurenine and starvation of tryptophan lead to T cell apoptosis, causing immune suppression within the tumor microenvironment [27]. A higher production of kynurenine in Cx43 knockdown B16F10 cell lines #2 and #3 was observed in Fig 1C. These results suggested Cx43 might be related to IDO and IDO-mediated activity.

Resveratrol and *Salmonella* upregulated Cx43 and inhibited IDO expression in melanoma cells

Previously, other and our studies found that *Salmonella* (*Salmonella enterica* serovar choleraesuis (*S. Choleraesuis*; S.C.) (ATCC 15480)) and resveratrol could enhance the expression of Cx43 [2, 27]. Hererin, the resveratrol and *Salmonella* as Cx43 activators were used to further evaluate the correlation between Cx43 and IDO. The results performed the upregulation of Cx43 by resveratrol (Fig. 2) and *Salmonella* (Fig. 3) accompanying the reduction in IDO and kynurenine production. Both of them possessed the ability of increasing Cx43 expression (Fig. 2A and Fig. 3A). Meanwhile, we examined the effect of resveratrol or S.C. on the Cx43 transcriptional activity by the luciferase reporter assay (Fig. 2B and Fig. 3B). The extent of Cx43 transcriptional activity of resveratrol or S.C. treatment in melanoma cells varied, ranging from 1.2 to 2.4 folds. A dose-dependently increase in Cx43 expression was regulated by resveratrol or *Salmonella*. On the contrary, IDO was downregulated both by resveratrol and *Salmonella* in dose-dependent manner. A low level of kynurenine production in resveratrol or *Salmonella*-treated groups (Fig. 2C and Fig. 3C). Next, the effect of kynurenine on T cell viability was performed in Fig. 2D and Fig. 3D. The conditioned medium from tumor cells treated with resveratrol or *Salmonella* increased the proliferation of T cell compared with control group (Fig. 2D and Fig. 3D). The results demonstrated that Cx43 activators reduced the expression and function of IDO in tumor cells.

Inhibited IDO expression by upregulation of Cx43

To confirm the role of Cx43 in regulation of IDO, Cx43 RNA interference (RNAi) were transfected to B16F10 to block Cx43 expression prior to the treatment of resveratrol or *Salmonella*. Since the Cx43 expression reach the highest level in 4 µg/ml of resveratrol and 100 MOI of *Salmonella*, the next experiments were all followed by the two concentrations. Cells transfected with Cx43 RNAi #73 showed better ability in reduction Cx43 expression whereas Cx43 RNAi #94 had less affect. After administrated with resveratrol (4

µg/ml) or *Salmonella* (MOI=100) with lower expression of Cx43 showed little change of IDO while IDO in control groups was still inhibited after the administration (Fig. 4A and 5A), and either was the production of kynurenine in RNAi transfected-groups (Fig. 4C and 5B). The Cx43 RNAi did not influence the

promoter activity (Fig. 4B). The blockade of Cx43 also prevented T cell proliferation being restored by resveratrol and *Salmonella* treatment (Fig. 4D and 5C). These data proved that Cx43 might be crucial in resveratrol and *Salmonella*-controlling IDO and tryptophan metabolites production.

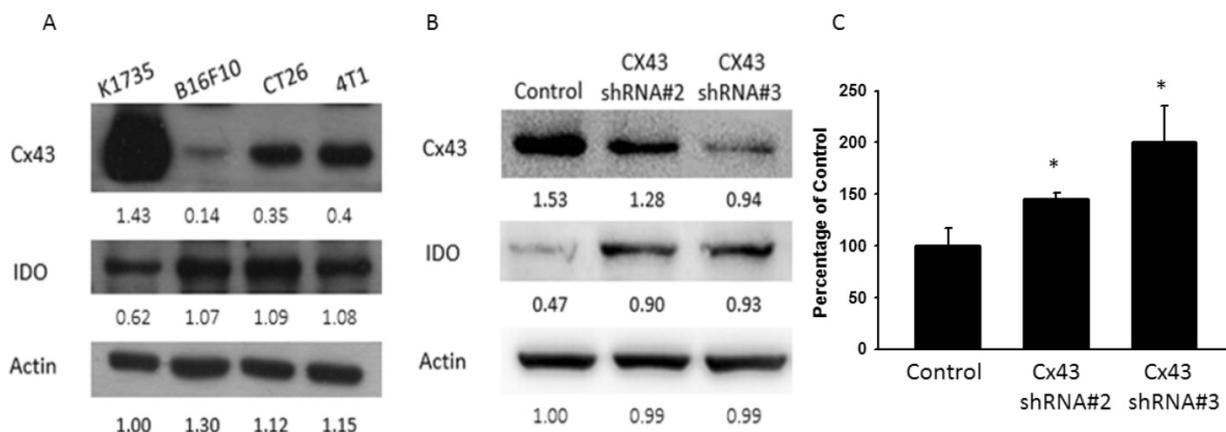


Figure 1. Cx43, IDO and kynurenine expression varies in different cancer cell lines. (A) The IDO and Cx43 protein level of different cancer cell lines, K1735, B16F10, CT26 and 4T1 were determined by Western blotting. (B) The IDO and Cx43 protein levels in Cx43-knockdown B16F10 cell lines were determined (C) The kynurenine production in Cx43-knockdown B16F10 cell lines were measured by kynurenine assay. (n = 3, data are mean± SD. *P< 0.05, **P < 0.01, ***P < 0.001).

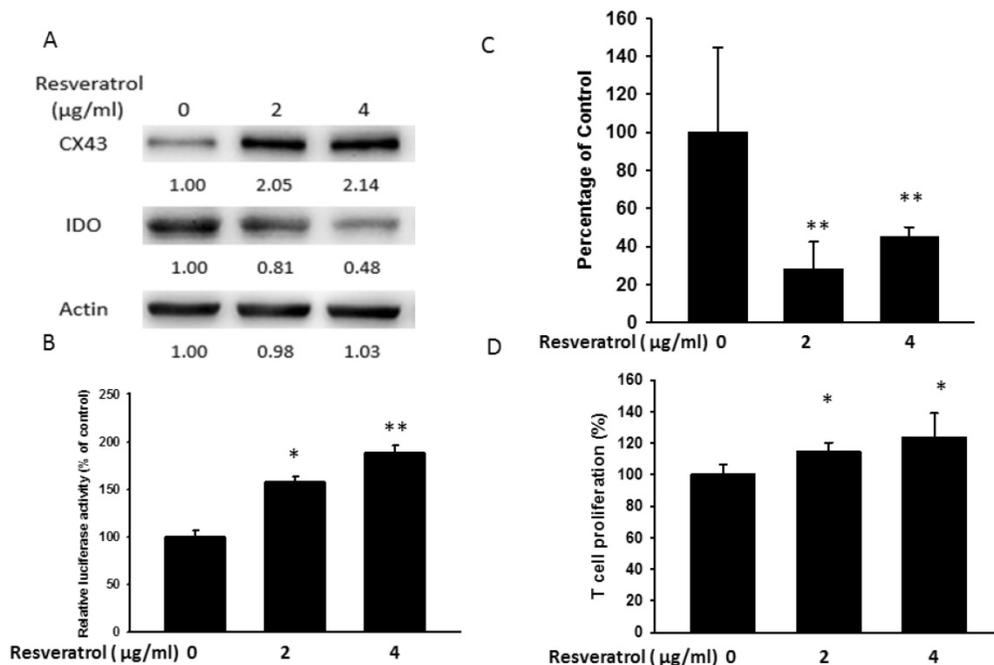


Figure 2. Resveratrol inhibited IDO expression dose dependently. B16F10 cells (10⁶) were treated with different concentrations of resveratrol (0-4 µg/ml) for 24 h. (A) The IDO and Cx43 protein levels of resveratrol were determined by Western blotting. (B) B16F10 cells transfected with luciferase gene under the control of Cx43 promoter were treated with resveratrol (0-4 µg/ml) for 24 h. The transcriptional activity of Cx43 was determined by the luciferase reporter assay and is expressed as the fold of the relative luciferase activity relative to that in the control tumor cells. (C) The kynurenine production were measured by kynurenine assay. (D) Supernatants were collected after 24 h resveratrol incubation, and harvested Jurkat cells with equal amount of RPMI. 3 days later, Jurkat cells were stained with trypan blue to measure proliferation rate. (n = 4, data are mean± SD. *P < 0.05, **P < 0.01).

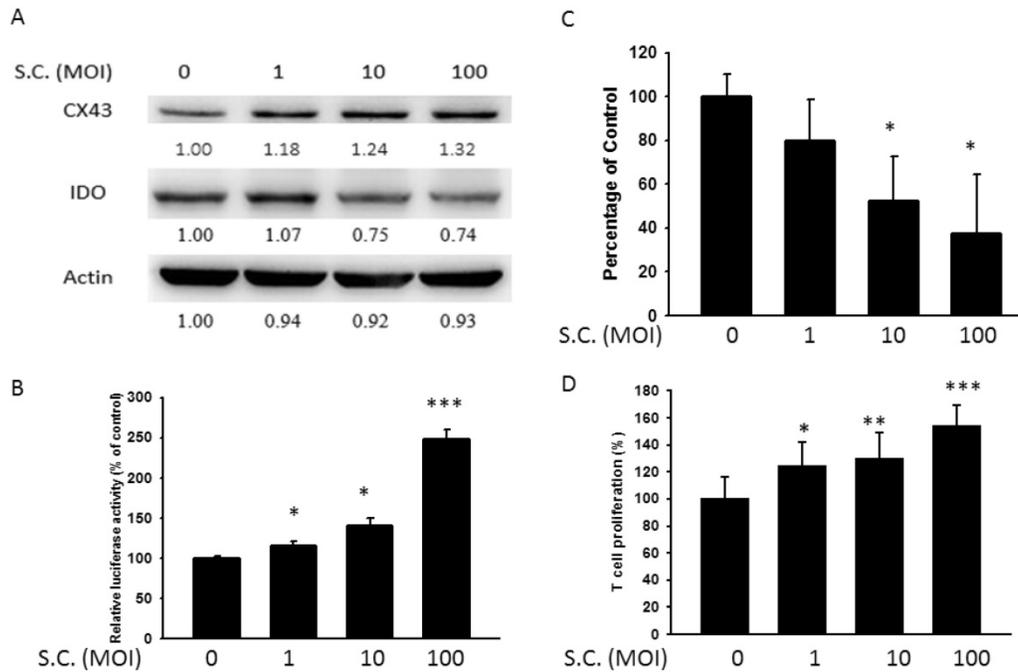


Figure 3. *Salmonella* (S.C.) inhibited IDO expression. B16F10 cells were treated with various MOI of S.C. (0-100) for 1.5 h and replaced medium with gentimicin-containing medium to kill bacteria. (A) The protein levels of IDO and Cx43 after resveratrol treatment were determined by Western blotting. (B) B16F10 cells transfected with luciferase gene under the control of Cx43 promoter were treated with S.C. (MOI = 0-100) for 24 h. The transcriptional activity of Cx43 was determined by the luciferase reporter assay and is expressed as the fold of the relative luciferase activity relative to that in the control tumor cells. (C) The kynurenine production were measured by kynurenine assay. (D) Supernatants were collected after 24 h resveratrol incubation, and harvested Jurkat cells with equal amount of RPMI. 3 days later, Jurkat cells were stained with trypan blue to measure proliferation rate. (n = 4, data are mean± SD. *P < 0.05, **P < 0.01, ***P < 0.001).

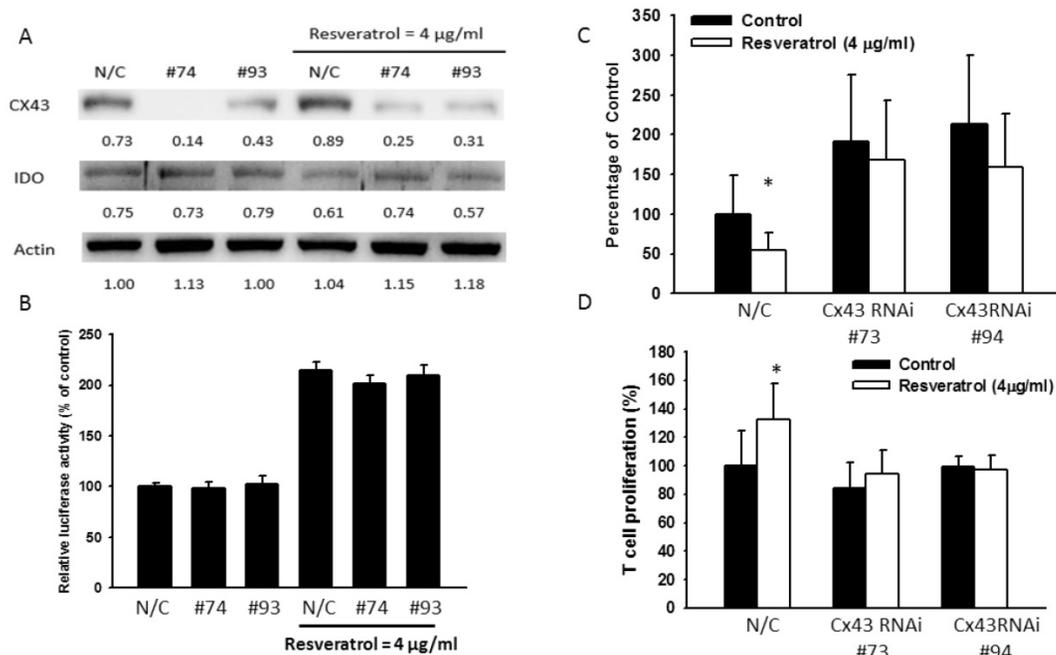


Figure 4. Resveratrol inhibited IDO expression through upregulating Cx43. B16F10 cells were transfected with Cx43 RNAi. Medium were replaced with 10% FBS contained DMEM. Transfected cells were treated with 4µg/ml resveratrol for 24 h. (A) The IDO and Cx43 protein level of resveratrol were determined by Western blotting. (B) B16F10 or Cx43 knockdown cells transfected with luciferase gene under the control of Cx43 promoter were treated with resveratrol (4 µg/ml) for 24 h. The transcriptional activity of Cx43 was determined by the luciferase reporter assay and is expressed as the fold of the relative luciferase activity relative to that in the control tumor cells. (D) The kynurenine production were measured by kynurenine assay. (C) Supernatants were collected after 24 h resveratrol incubation, and harvested Jurkat cells with equal amount of RPMI. 3 days later, Jurkat cells were stained with trypan blue to measure proliferation rate. (n = 6, data are mean± SD. *P < 0.05).

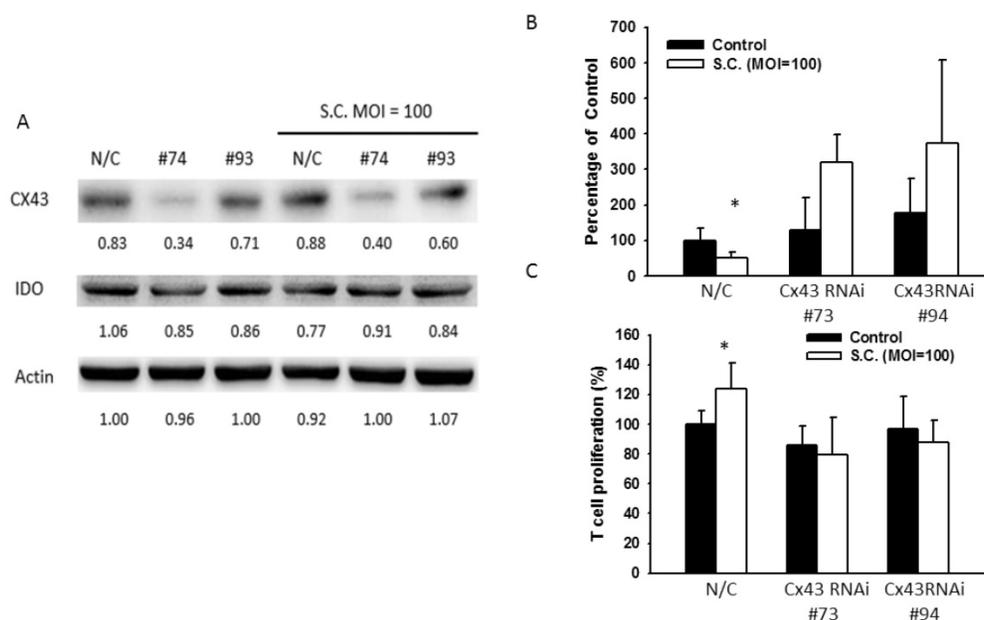


Figure 5. *Salmonella* (S.C.) inhibited IDO expression through upregulating Cx43. B16F10 cells were transfected with Cx43 RNAi. Medium were replaced with 10% FBS contained DMEM. Transfected cells were treated with *Salmonella* (MOI=100). (a) The IDO and Cx43 protein level were determined by Western blotting after *Salmonella* treatment. (B) The kynurenine production were measured by kynurenine assay. (C) Supernatants were collected after 24 h resveratrol incubation, and harvested Jurkat cells with equal amount of RPMI. 3 days later, Jurkat cells were stained with trypan blue to measure proliferation rate. (n = 6, data are mean \pm SD. *P < 0.05).

Discussion

In this study, we investigated the molecular mechanism that Cx43 involves in IDO regulation. The expression of Cx43 varies in different cancer cell lines and cells with lower Cx43 exhibited a higher IDO level, indicating Cx43 might correlate with IDO expression. Then, resveratrol and *Salmonella* which have been reported to upregulate Cx43 were exerted to evaluate the Cx43-IDO signaling pathway. The administration of resveratrol and *Salmonella* in B16F10 increased Cx43 expression with reduced protein level of IDO and kynurenine content significantly. Furthermore, knockdown of Cx43 inhibited the resveratrol and *Salmonella*-mediated IDO reduction, indicating Cx43 might be crucial in regulation of tumor-releasing IDO. Moreover, the treatment of well-known IDO inducer, interferon- γ (IFN- γ) [28], was not used in our experiment because IDO and Cx43 protein expressed the same pattern after IFN- γ treatment, indicating Cx43 might not be the main regulator in IFN- γ -induced IDO expression (data not shown). In spite of the success *in vivo* experiment, the utilization of resveratrol and *Salmonella* in clinical studies still encountered several problems. Rapid metabolism made resveratrol hard to maintain therapeutic effect in blood level [29]. Administration of resveratrol did not achieve the expected results on established cancer and safety concerns were observed in multiple myeloma patients with kidney problems [30, 31]. Gene modified strain *Salmonella*, VNP20009,

showed low colonization in phase I melanoma clinical trial [32]. The unexpected outcome might contribute to lack of lipid A and impaired ability of producing purine which was known to be expressed poorly in melanoma [33, 34]. The alternation of biosynthesis based on safety concerns made gene-modified strain applied to limited cancer type. Hence, current researches focused on supportive role of *Salmonella* or resveratrol in chemoresistance, anti-angiogenesis and immune modulation etc. rather than their direct toxicity to tumor cells [24, 26, 35-38]. Here, we identified the potential of resveratrol and *Salmonella* that regulating Cx43 and IDO. Ino and colleagues demonstrated the high expression of IDO in endometrial cancer clinical surgical specimen has correlation with disease progression and poor prognosis [39]. Being sensitive to tryptophan, T cells sense IDO-mediated tryptophan depletion by uncharged tRNAs and trigger kinase general control non-depressible 2 (GCN2) resulting in cell cycle arrest and CD8+ T cells anergy [40]. The binding of tryptophan metabolites and aryl hydrocarbon receptor (AHR) involves in the differentiation of naive helper T cells (Th) to Treg phenotype. Moreover, IDO serves as a signaling protein triggered by transforming growth factor- β (TGF- β) is responsible for maintain long-term tolerance [41]. 1-methyl-tryptophan (1-MT) is now a widely studied tryptophan analog with higher affinity to IDO thus abrogating the IDO-mediated immune tolerance. One of 1-MT isoform, 1-methyl-D-tryptophan was

conducted in several ongoing clinical trials [5]. However, one still cast doubts on IDO-inhibited ability of 1-MT since an upregulation of IDO was seen in 1-MT-treated cancer cells [42]. The other IDO selective inhibitor, Epacadostat, also possesses higher affinity to IDO than tryptophan without affecting other IDO-related enzyme, was also exerted in several ongoing clinical trial accompanied with other chemotherapeutic drugs, while the correlation between results and biomarkers still being evaluating [43]. Cx43 used to be classified as tumor suppressor gene, however, new evidence indicated the role of Cx43 in tumor progression depends on cancer stage [44]. Evidence showed that Cx43 and Cx43-mediated gap junction is important in forming immune synapse and activating T cells by regulating Ca²⁺ signals. The data also performed Cx43 and Cx43-mediated gap junction involved in the regulation of IFN- γ since the inhibition of which abrogated secretion of IFN- γ in T cells [6]. The upregulation of Cx43 and its mediated functional gap junction in tumor cells facilitated antigen transferring between immune cells and tumor cells, thus contributing to antitumor response [1, 2]. The other studies also pointed out that the silencing Cx43 suppressed synovial fibroblast-releasing pro-inflammatory cytokines such as IFN- γ , tumor necrosis factor- α (TNF- α) which should have been stimulated by lipopolysaccharides [5]. These findings inferred the importance of Cx43 in regulating immunity. In this article, our results identified the underlying molecular mechanism that Cx43 controlled IDO and IDO-mediated metabolites. Our findings might raise further investigation of Cx43-related tumor immune tolerance and highlight the potential of Cx43 and IDO in developing anticancer treatment.

Abbreviations

IDO: Indoleamine 2, 3-dioxygenase 1; Treg: regulatory T cells; Cx43: Connexin43; S.C.: *Salmonella enterica* serovar choleraesuis; IFN- γ : interferon- γ ; GCN2: kinase general control non- derepressible 2; AHR: aryl hydrocarbon receptor; Th: helper T cells; TGF- β : transforming growth factor- β ; 1-MT: 1-methyl-tryptophan; IFN- γ : interferon- γ ; TNF- α : tumor necrosis factor- α ; DMEM: Dulbecco's Modified Eagle Medium.

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Author Contributions

H.C.L and C.H.L conceived and designed the experiments; C.J.Y, W.K.W, Y.D.K. and H.C.L. performed the experiments; C.J.Y. and C.H.L analyzed the data; C.H.L contributed reagents/materials /analysis tools; Y.D.K. and C.H.L wrote the paper.

Competing Interests

The authors have declared that no competing interest exists.

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