

Research Paper

A Nonsense Mutation of γ D-crystallin Associated with Congenital Nuclear and Posterior Polar Cataract in a Chinese Family

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Abstract

Objective: The goal of this study was to characterize the disease-causing mutations in a Chinese family with congenital nuclear and posterior polar cataracts. **Methods:** Clinical data of patients in the family were recorded using slit-lamp photography and high definition video. Genomic DNA samples were extracted from the peripheral blood of the pedigree members and 100 healthy controls. Mutation screening was performed in the candidate genes by bi-directional sequencing of the amplified products. **Results:** The congenital cataract phenotype of the pedigree was identified by slit-lamp examinations and observation during surgery as nuclear and posterior polar cataracts. Through the sequencing of the candidate genes, a heterozygous c. 418C>T change was detected in the coding region of the γ D-crystallin gene (CRYGD). As a result of this change, a highly conserved arginine residue was replaced by a stop codon (p. R140X). This change was discovered among all of the affected individuals with cataracts, but not among the unaffected family members or the 100 ethnically matched controls. **Conclusions:** This study identified a novel congenital nuclear and posterior polar cataract phenotype caused by the recurrent mutation p. R140X in CRYGD.

Key words: congenital cataract, γ D-crystallin, mutation, stop codon, Greek key motif

Introduction

Cataracts can be defined as complete or partial lens opacification, either congenital or acquired. A congenital cataract is especially severe, as it potentially impairs visual development, and is one of the leading causes of childhood blindness. The prevalence of congenital cataracts is approximately 6.31/100,000 [1], 27-39% of which are believed to be inherited [2]. Autosomal dominant (AD) inheritance is commonly observed among hereditary cataracts, while autosomal recessive and X-linked patterns have also been reported [3].

To date, scientists have identified more than 35 loci, including over 20 genes which are associated with isolated congenital cataracts [4]. Among the

congenital cataract causative mutations discovered thus far, approximately half of the mutations are in the crystallin family genes, and one quarter of them are in gap junction genes. Other mutations have been identified in heat shock transcription factor-4 (HSF4), aquaporin-0 (AQP0, MIP), v-maf avian musculoaponeurotic fibrosarcoma oncogene homolog (MAF), and beaded filament structural protein-2 (BFSP2), as well as other genes [5].

In this study, we investigated a three-generation family with congenital nuclear and posterior polar cataracts, and detected a R140X nonsense mutation in γ D-crystallin, which cosegregated with the disease in this family.

Methods

The research protocols of this study adhered to the guidelines of the Declaration of Helsinki and were approved by the Medical Ethics Committees of the Second Affiliated Hospital, College of Medicine, Zhejiang University (Hangzhou, China). Appropriate informed consent from each participant was obtained.

Clinical Evaluation

A three-generation Chinese pedigree consisting of eleven individuals, including two affected individuals, formed the basis for this study. All eleven family members participated in the study (two affected and nine unaffected individuals; Figure 1). The proband (III:1) was a 1 year-old girl who visited our eye center in June of 2013 for a cataract extraction surgery. Her father (II:1), diagnosed with congenital cataracts 30 years previously (at birth), had his cataract extraction and intraocular lens implantation 16 years previously at our eye center as well. All of the patients and family members underwent ophthalmic examinations that included visual function, slit-lamp examination, intraocular pressure measurement, and fundus evaluation with dilated pupils. In addition, 100 unrelated healthy subjects were recruited as a control group.

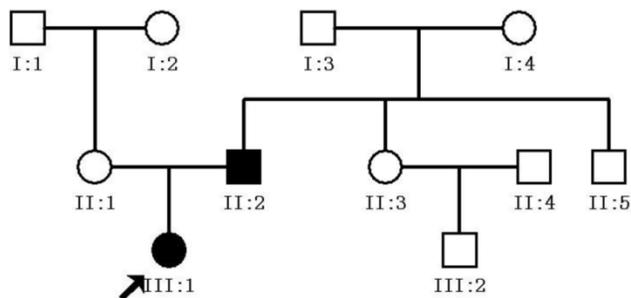


Figure 1. Pedigree of Chinese family with congenital cataracts. The proband is marked with an arrow. Squares and circles indicate males and females, respectively. Black and white symbols represent affected and unaffected individuals, respectively.

Mutation Screening

We used the functional candidate gene analysis approach for screening, and the detailed strategy was described in our previous study [6]. The experimental procedure is briefly described here. Peripheral blood was collected by venipuncture in EDTA-coated Becton-Dickinson Vacutainer tubes (BD, New Jersey, USA) and stored at -20°C . Genomic DNA was extracted from peripheral lymphocytes using the QIAamp Blood kit (Qiagen, Dusseldorf, Germany). Polymerase chain reaction (PCR) was used to amplify the exons and the flanking intronic regions of the candidate genes: crystallin alpha A (CRYAA), crystal-

lin alpha B (CRYAB), crystallin beta A3/A1 (CRYBA3/1), crystallin beta B1 (CRYBB1), crystallin beta B2 (CRYBB2), crystallin gamma C (CRYGC), crystallin gamma D (CRYGD), gap junction protein alpha 3 (GJA3), gap junction protein alpha 8 (GJA8), and major intrinsic protein (MIP). The primer sequences were described in our previous study [7], and the specific primer sequences for the CRYGD are listed below (Table 1). Bi-directional direct sequencing of the PCR products was performed afterwards.

Bioinformatics Analysis

The γ D-crystallin amino acid sequences were obtained from the NCBI Gene Database (<http://www.ncbi.nlm.nih.gov/gene/>). We obtained the structure of wide-type human γ D-crystallin from the Protein Data Bank (PDB) database (ID:1HK0), and visualized the protein structure using PyMOL software (DeLano Scientific LLC, San Francisco, CA, USA). We also used the BEST/COREX web server (<http://best.bio.jhu.edu/BEST/index.php>) to predict the stability of the native and mutant proteins.

Results

Clinical Evaluation

We identified isolated bilateral congenital nuclear and posterior polar cataracts in the Chinese family, and the two patients showed the same clinical symptoms: lens opacification, horizontal nystagmus and amblyopia. Individual II:2 was diagnosed with congenital cataracts 30 years previously, at his birth, and had phacoemulsification and intraocular lens implantation 16 years previously. According to his medical records, his lens opacities were located mainly at the lens nuclei and posterior poles. Individual III:1 was diagnosed with congenital cataracts at the age of 3 months. Slit-lamp examination revealed that III:1 had nuclear and posterior polar opacities with posterior lenticonus. Individual III:1 underwent surgeries for both eyes at one year of age. According to our observation, during the surgeries, both eyes of individual III:1 had nuclear and posterior polar opacities, with a posterior capsular thinning or defect, which are secondary complications to the posterior lenticonus [8]. However, each eye had a different appearance. Her right nuclear opacities had been reabsorbed, which made the remnants drop into the vitreous body through the posterior capsular defect. Her left eye had intact nuclear cataracts and posterior lenticonus with round posterior capsular thinning (Figure 2). We performed irrigation-aspiration combined with anterior vitrectomy on the right eye, and standard phacoemulsification on the left (high-definition video can be provided upon request). The postoperative best-corrected visual acuity (BCVA) of affected

member II:2 was 0.1/0.1 due to amblyopia, and he still exhibited severe horizontal nystagmus. Individual III:1's BCVA was unable to be measured since she was still preverbal; however, the postoperative visual

stimuli could be fixed, centered and followed. Intra-ocular lenses will be implanted when she is two years of age.

Table 1. Primers used in PCR of CRYGD.

Name	Prime Sequence (5'-3')	Product length (bp)
Exon-1.2F	5' CCTCGCCTTGTCCCGC 3'	340
Exon-1.2R	5' TTAACITTTTGCTTGAAACCATCCA 3'	
Exon-3F	5' TGCITTTTCTCTCTTTTATTTCGGGTCC 3'	400
Exon-3R	5' AGTAAAGAAAGACACAAGCAAATCAGTGCC 3'	

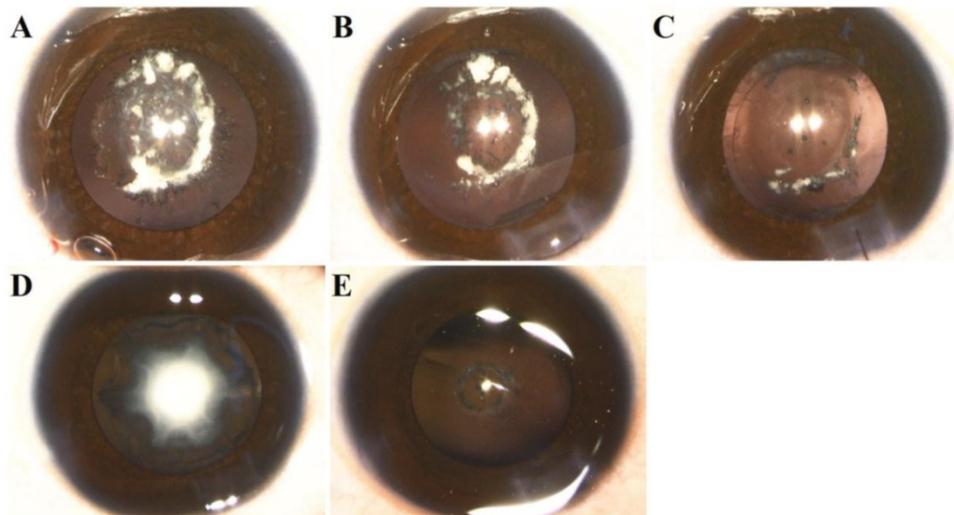


Figure 2. Phenotype of individual III:1. A: Photograph of right eye before surgery showing that lens was partially reabsorbed, with some dropping into the vitreous body through the posterior capsular hole. B: Photograph of right eye after irrigation/aspiration showing posterior capsular punctate opacities with central round hole; lens remnants remaining in the anterior segment of the vitreous. C: Photograph of right eye after anterior vitrectomy. D: Photograph of left eye before surgery showing nuclear opacities. E: Photograph of left eye after phacoemulsification and irrigation/aspiration showing round thinning of posterior capsule without rupture, protruding into the vitreous, and punctate opacities around the posterior pole.

Mutation Screening

Mutation screenings were performed for all candidate genes, and a heterozygous change C>T at position 418 (c. 418C>T) was identified in exon 3 of the CRYGD gene (Figure 3). This mutation resulted in the substitution of a stop codon for a phylogenetically conserved arginine residue (p. R140X), and was only identified in the patients. It was not found in either the healthy family members or the 100 controls with same ethnic background. Moreover, this mutation was not present in either the 1000 Genomes Project database (<http://browser.1000genomes.org/index.html>) or the NHLBI Exome Variant Server (EVS, <http://evs.gs.washington.edu/EVS/>). With the exception of several nonpathogenic single nucleotide polymorphisms, no other mutations were detected.

Bioinformatics Analysis

The mutant protein is 35 amino acids shorter than the wild-type one, and the fourth Greek key motif (GKM) at the C-terminus was partially absent

(Figure 4). The BEST/COREX generated a coarse-grained energy evaluation at the residue level by calculating the structural thermodynamic ensembles from native and mutant γ D-crystallin (Table 2). Compared to the mutant, the native γ D-crystallin has a positive delta G for unfolding, while the mutant favors the C-terminal domain in the unfolded state by a negative delta G [9]. Moreover, the stability constant at the per residue level showed that the C-terminal domain of the mutant protein favors the unfolded states (Figure 5).

Table 2. COREX/BEST scores

	Lowest energy state in ensemble	
	Native	Mutant
Fraction Folded	0.9769	0.5797
Predicted delta G (Kcal/mol)	2.053	-9.209
% total ensemble simulated	95%	83%
Residue States (F=Folded; U=Unfolded)	F(1-169)U(170-174)	F(1-80)U(81-139)

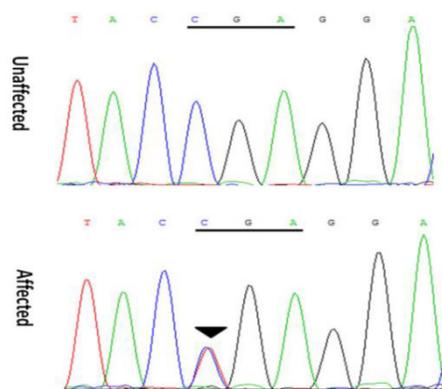


Figure 3. Forward sequence analysis of the affected and unaffected individuals with congenital cataracts in this Chinese family, showing a heterozygous mutation (c. 418C>T) in exon 3 of CRYGD (black triangles).

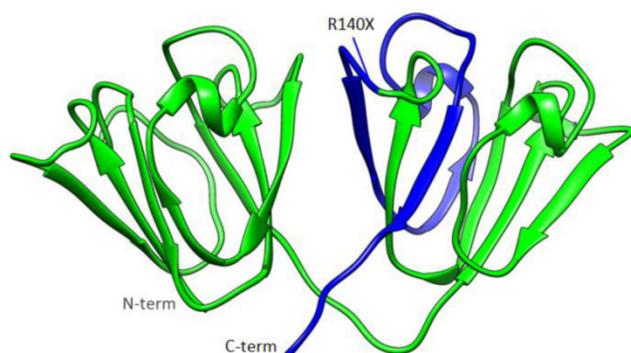


Figure 4. The structural model of the γ D-crystallin mutation. A structural model of γ D-crystallin is shown in this illustration, in which the truncated portion resulting from the R140X mutation is highlighted in blue, while the rest of the protein is depicted in green.

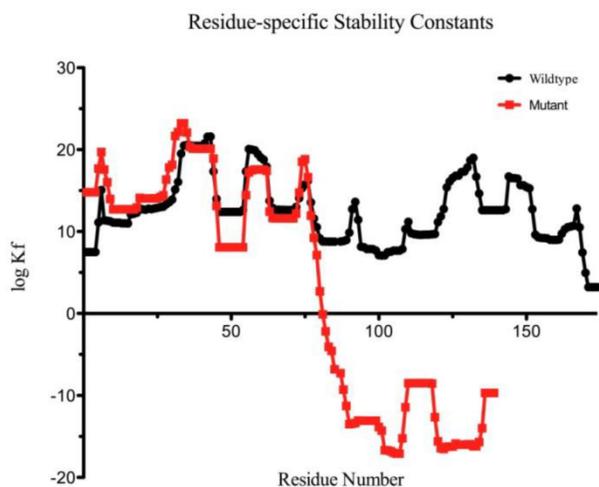


Figure 5. Residue-specific Stability Constant for each residue of the protein as predicted by the BEST/COREX server. The $\log K_f$ stability constant is the ratio of the summed probability of the residue in folded conformation to the summed probability of the residue in unfolded conformation.

Discussion

In this study, we report a nonsense mutation (c. 418C>T) in exon 3 of the CRYGD gene which caused congenital nuclear and posterior polar cataracts in a

Chinese family. This mutation cosegregated with affected individuals, and was not detected in either the healthy family members or 100 ethnically matched controls.

Crystallins are structural proteins that play essential roles in lens development and the maintenance of transparency. Mammalian lens crystallins have a core set of three protein classes, the α -, β - and γ -crystallins, which are defined by the sizes of the oligomers that they form [10]. With the lowest molecular mass (20 kDa), the γ -crystallin family proteins are strictly monomeric [11]. The cDNA analyses show that most of the transcripts arising from this family among humans come from γ C-crystallin and γ D-crystallin, with the latter being the predominantly expressed species [12].

γ D-crystallin is a member of the β/γ -crystallins super family, which shares the common feature of antiparallel β -sheets in the protein, called the "Greek key motif" [11]. In the γ D-crystallin, the N-terminal and C-terminal halves of the sequence comprise two individual domains that each consist of two GKM, which interact via the GKM2 and GKM4 [13]. Each GKM is composed of four antiparallel β -strands, and the R140X mutation in the present study is located in GKM4.

The transition of C>T at c. 418 in exon 3 of CRYGD is predicted to cause a premature stop codon, which leads to the deletion of the C-terminal residues 140–174. The overall γ D-crystallin structure is depicted where the truncated portion from the R140X mutation is colored in blue (Figure 4).

As shown in the diagram, this deletion mutant loses its two β -strands of the fourth GKM, which destabilizes the C-terminal domain. Additionally, the missing residues may not only influence the folding of the domain, but also affect the aggregation state of the protein. A previous mutant investigation indicated that the R140X mutant exposed several buried residues to the surface, and displayed lower solubility and structural stability [14]. Venu Talla and co-workers reported that the loss of the C-terminal fragment in human γ D-crystallin would lead to substantial intermolecular aggregates, which was expected to generate light-scattering particles, compromising the transparency of the cells and their assemblies [15]. The other two previously reported nonsense mutations in CRYGD, Y134X and W157X, also affected the fourth GKM and caused congenital cataracts.

Since γ D-crystallin expression remains at a relatively high level in the human lens throughout one's lifetime, the CRYGD gene mutation has been associated with not only congenital cataracts, but also juvenile-onset cataracts. Mutations in the human CRYGD

gene that cause childhood cataracts have been identified in more than 23 families thus far (Table 3) [16-37]. The R140X mutation was observed in our study as well as in another previously described family with congenital cataracts [35]. Our study indicates that the R140X is the actual causative mutation, and although the two studies revealed the same R140X amino acid

alternation, the phenotypes were different. The previous study presented an exclusively nuclear congenital cataract family, and did not provide photographs of the affected individuals. Our study, however, suggests that the R140X mutation could present a novel nuclear and posterior polar cataract phenotype, and provides the photographic records of the patients.

Table 3. Human CRYGD mutations associated with childhood cataracts.

Year	Mutation	Amino acid change	Origin of family	Phenotype	Reference
1999	c.43C>T	R15C	American	Juvenile-onset punctate cataracts	[16]
2006	c.43C>T	R15C	Chinese	Congenital coralliform cataract	[17]
2009	c.43C>A	R15S	Chinese	Congenital coralliform cataract	[18]
2007	c.70C>T	P24S	Russian	Polymorphic congenital cataract	[19]
2003	c.70C>A	P24T	French	Congenital cerulean cataract	[20]
2004	c.70C>A	P24T	American	Congenital coral-like cataract	[21]
2004	c.70A>C	P24T	Chinese	Congenital fasciculiform cataract	[22]
2011	c.106G>C	A36P	Chinese	Congenital nuclear cataract	[23]
2000	c.109C>A	R37S	Czechs	Congenital crystal-like cataract	[24]
2011	c.109C>A	R37S	American	Congenital crystal-like cataract	[25]
2005	c.109C>A	R37S	Chinese	Congenital nuclear cataract	[26]
2011	c.110G>C	R37P	Chinese	Congenital nuclear cataract	[27]
2011	c.127T>C	W43R	Chinese	Congenital nuclear cataract	[28]
2009	c.168C>G	Y56X	Brazilian	Congenital nuclear cataract	[29]
2005	c.173G>A	R59H	Mexican	Congenital aculeiform cataract	[30]
2008	c.181G>T	G61C	Chinese	Congenital coralliform cataract	[31]
2010	c.229C>A	R77S	Indian	Juvenile-onset anterior polar cataract	[32]
2006	c.320A>C	E107A	Mexican	Congenital nuclear cataract	[33]
2009	c.418C>A	Y134X	Danish	Not mentioned	[34]
2008	c.418C>T	R140X	Indian	Congenital nuclear cataract	[35]
2013	c.418C>T	R140X	Chinese	Congenital nuclear and posterior polar cataract	Present study
2002	c.470G>A	W157X	German	Congenital nuclear cataract	[36]
2007	c.494delG	165 new amino acid	Chinese	Congenital nuclear cataract	[37]

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Competing Interests

The authors have declared that no competing interest exists.

References

- Bermejo E, Martinez-Frias ML: Congenital eye malformations: clinical-epidemiological analysis of 1,124,654 consecutive births in Spain. *American journal of medical genetics* 1998, 75(5):497-504.
- Haargaard B, Wohlfahrt J, Fledelius HC, Rosenberg T, Melbye M: A nationwide Danish study of 1027 cases of congenital/infantile cataracts: etiological and clinical classifications. *Ophthalmology* 2004, 111(12):2292-2298.
- Hejtmancik JF: Congenital cataracts and their molecular genetics. *Seminars in cell & developmental biology* 2008, 19(2):134-149.
- Shiels A, Bennett TM, Hejtmancik JF: Cat-Map: putting cataract on the map. *Molecular vision* 2010, 16:2007-2015.
- Huang B, He W: Molecular characteristics of inherited congenital cataracts. *European journal of medical genetics* 2010, 53(6):347-357.
- Zhu Y, Shentu X, Wang W, Li J, Jin C, Yao K: A Chinese family with progressive childhood cataracts and IVS3+1G>A CRYBA3/A1 mutations. *Molecular vision* 2010, 16:2347-2353.
- Li J, Wang Q, Fu Q, Zhu Y, Zhai Y, Yu Y, Zhang K, Yao K: A novel connexin 50 gene (gap junction protein, alpha 8) mutation associated with congenital nuclear and zonular pulverulent cataract. *Molecular vision* 2013, 19:767-774.
- Amaya L, Taylor D, Russell-Eggitt I, Nischal KK, Lengyel D: The morphology and natural history of childhood cataracts. *Survey of ophthalmology* 2003, 48(2):125-144.
- Vertrees J, Barritt P, Whitten S and Hilser VJ. COREX/BEST server: a web browser-based program that calculates regional stability variations within protein structures. *Bioinformatics* 2005, 21:3318-3319.
- Wistow G: The human crystallin gene families. *Human genomics* 2012, 6:26.
- Graw J: Genetics of crystallins: cataract and beyond. *Experimental eye research* 2009, 88(2):173-189.
- Wistow G, Bernstein SL, Wyatt MK, Behal A, Touchman JW, Bouffard G, Smith D, Peterson K: Expressed sequence tag analysis of adult human lens for the NEIBank Project: over 2000 non-redundant transcripts, novel genes and splice variants. *Molecular vision* 2002, 8:171-184.
- Blundell T, Lindley P, Miller L, Moss D, Slingsby C, Tickle I, Turnell B, Wistow G: The molecular structure and stability of the eye lens: x-ray analysis of gamma-crystallin II. *Nature* 1981, 289(5800):771-777.
- Vendra VP, Agarwal G, Chandani S, Talla V, Srinivasan N, Balasubramanian D: Structural Integrity of the Greek Key Motif in betagamma-Crystallins Is Vital for Central Eye Lens Transparency. *PLoS one* 2013, 8(8):e70336.
- Talla V, Srinivasan N, Balasubramanian D: Visualization of in situ intracellular aggregation of two cataract-associated human gamma-crystallin mutants: lose

- a tail, lose transparency. *Investigative ophthalmology & visual science* 2008, 49(8):3483-3490.
16. Stephan DA, Gillanders E, Vanderveen D, Freas-Lutz D, Wistow G, Baxeavanis AD, Robbins CM, VanAuken A, Quesenberry MI, Bailey-Wilson J *et al*: Progressive juvenile-onset punctate cataracts caused by mutation of the gammaD-crystallin gene. *Proceedings of the National Academy of Sciences of the United States of America* 1999, 96(3):1008-1012.
 17. Gu F, Li R, Ma XX, Shi LS, Huang SZ, Ma X: A missense mutation in the gammaD-crystallin gene CRYGD associated with autosomal dominant congenital cataract in a Chinese family. *Molecular vision* 2006, 12:26-31.
 18. Zhang LY, Gong B, Tong JP, Fan DS, Chiang SW, Lou D, Lam DS, Yam GH, Pang CP: A novel gammaD-crystallin mutation causes mild changes in protein properties but leads to congenital coralliform cataract. *Molecular vision* 2009, 15:1521-1529.
 19. Plotnikova OV, Kondrashov FA, Vlasov PK, Grigorenko AP, Ginter EK, Rogaev EI: Conversion and compensatory evolution of the gamma-crystallin genes and identification of a cataractogenic mutation that reverses the sequence of the human CRYGD gene to an ancestral state. *American journal of human genetics* 2007, 81(1):32-43.
 20. Nandrot E, Slingsby C, Basak A, Cherif-Chefchaoui M, Benazzou B, Hajaji Y, Boutayeb S, Gribouval O, Arbogast L, Berraho A *et al*: Gamma-D crystallin gene (CRYGD) mutation causes autosomal dominant congenital cerulean cataracts. *Journal of medical genetics* 2003, 40(4):262-267.
 21. Mackay DS, Andley UP, Shiels A: A missense mutation in the gammaD crystallin gene (CRYGD) associated with autosomal dominant "coral-like" cataract linked to chromosome 2q. *Molecular vision* 2004, 10:155-162.
 22. Shentu X, Yao K, Xu W, Zheng S, Hu S, Gong X: Special fasciculiform cataract caused by a mutation in the gammaD-crystallin gene. *Molecular vision* 2004, 10:233-239.
 23. Sun W, Xiao X, Li S, Guo X, Zhang Q: Mutation analysis of 12 genes in Chinese families with congenital cataracts. *Molecular vision* 2011, 17:2197-2206.
 24. Kmoch S, Brynda J, Asfaw B, Bezouska K, Novak P, Rezacova P, Ondrova L, Filipec M, Sedlacek J, Elleder M: Link between a novel human gammaD-crystallin allele and a unique cataract phenotype explained by protein crystallography. *Human molecular genetics* 2000, 9(12):1779-1786.
 25. VanderVeen DK, Andrews C, Nihalani BR, Engle EC: Crystalline cataract caused by a heterozygous missense mutation in gammaD-crystallin (CRYGD). *Molecular vision* 2011, 17:3333-3338.
 26. Gu J, Qi Y, Wang L, Wang J, Shi L, Lin H, Li X, Su H, Huang S: A new congenital nuclear cataract caused by a missense mutation in the gammaD-crystallin gene (CRYGD) in a Chinese family. *Molecular vision* 2005, 11:971-976.
 27. Wang L, Chen X, Lu Y, Wu J, Yang B, Sun X: A novel mutation in gammaD-crystallin associated with autosomal dominant congenital cataract in a Chinese family. *Molecular vision* 2011, 17:804-809.
 28. Wang B, Yu C, Xi YB, Cai HC, Wang J, Zhou S, Zhou S, Wu Y, Yan YB, Ma X *et al*: A novel CRYGD mutation (p.Trp43Arg) causing autosomal dominant congenital cataract in a Chinese family. *Human mutation* 2011, 32(1):E1939-1947.
 29. Santana A, Waiswol M, Arcieri ES, Cabral de Vasconcellos JP, Barbosa de Melo M: Mutation analysis of CRYAA, CRYGC, and CRYGD associated with autosomal dominant congenital cataract in Brazilian families. *Molecular vision* 2009, 15:793-800.
 30. Zenteno JC, Morales ME, Moran-Barroso V, Sanchez-Navarro A: CRYGD gene analysis in a family with autosomal dominant congenital cataract: evidence for molecular homogeneity and intrafamilial clinical heterogeneity in aculeiform cataract. *Molecular vision* 2005, 11:438-442.
 31. Li F, Wang S, Gao C, Liu S, Zhao B, Zhang M, Huang S, Zhu S, Ma X: Mutation G61C in the CRYGD gene causing autosomal dominant congenital coralliform cataracts. *Molecular vision* 2008, 14:378-386.
 32. Roshan M, Vijaya PH, Lavanya GR, Shama PK, Santhiya ST, Graw J, Gopinath PM, Satyamoorthy K: A novel human CRYGD mutation in a juvenile autosomal dominant cataract. *Molecular vision* 2010, 16:887-896.
 33. Messina-Baas OM, Gonzalez-Huerta LM, Cuevas-Covarrubias SA: Two affected siblings with nuclear cataract associated with a novel missense mutation in the CRYGD gene. *Molecular vision* 2006, 12:995-1000.
 34. Hansen L, Mikkelsen A, Nurnberg P, Nurnberg G, Anjum I, Eiberg H, Rosenberg T: Comprehensive mutational screening in a cohort of Danish families with hereditary congenital cataract. *Investigative ophthalmology & visual science* 2009, 50(7):3291-3303.
 35. Devi RR, Yao W, Vijayalakshmi P, Sergeev YV, Sundaresan P, Hejtmančík JF: Crystallin gene mutations in Indian families with inherited pediatric cataract. *Molecular vision* 2008, 14:1157-1170.
 36. Santhiya ST, Shyam Manohar M, Rawley D, Vijayalakshmi P, Namperumalsamy P, Gopinath PM, Loster J, Graw J: Novel mutations in the gamma-crystallin genes cause autosomal dominant congenital cataracts. *Journal of medical genetics* 2002, 39(5):352-358.
 37. Zhang LY, Yam GH, Fan DS, Tam PO, Lam DS, Pang CP: A novel deletion variant of gammaD-crystallin responsible for congenital nuclear cataract. *Molecular vision* 2007, 13:2096-2104.