

Research Paper

## Improved Synthesis Strategy for Peptide Nucleic Acids (PNA) appropriate for Cell-specific Fluorescence Imaging

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### Abstract

Progress in genomics and proteomics attended to the door for better understanding the recent rapid expanding complex research field of metabolomics. This trend in biomedical research increasingly focuses to the development of patient-specific therapeutic approaches with higher efficiency and sustainability. Simultaneously undesired adverse reactions are avoided. In parallel, the development of molecules for molecular imaging is required not only for the imaging of morphological structures but also for the imaging of metabolic processes like the aberrant expression of the cysteine protease cathepsin B (*CtsB*) gene and the activity of the resulting product associated with metastasis and invasiveness of malign tumors. Finally the objective is to merge imaging and therapy at the same level. The design of molecules which fulfil these responsibilities is pivotal and requires proper chemical methodologies. In this context our modified solid phase peptide chemistry using temperature shifts during synthesis is considered as an appropriate technology. We generated highly variable conjugates which consist of molecules useful as diagnostically and therapeutically active molecules. As an example the modular PNA products with the complementary sequence to the *CtsB* mRNA and additionally with a cathepsin B cleavage site had been prepared as functional modules for distinction of cell lines with different *CtsB* gene expression. After ligation to the modular peptide-based BioShuttle carrier, which was utilized to facilitate the delivery of the functional modules into the cells' cytoplasm, the modules were scrutinized.

Key words: Click Chemistry; Diels Alder Reaction<sub>inverse</sub> (DAR<sub>inv</sub>); Fluorescence Imaging; Peptide Nucleic Acid (PNA); PNA building block functionalization

### Introduction

A search for the very first documentations of peptide nucleic acids (PNA) led to Miller's and Urey's experiments in the year 1953. They verified the hypothesis of the appearance of organic molecules under reducing atmospheric conditions [1]. The PNA's role as a pivotal prebiotic molecule in the RNA world which acts as a template for the polymerization of the

complementary nucleotide phosphorimidazoles was postulated [2-6]. The focus lies on Nielsen's successful pioneering work in the PNA syntheses [7]. Contributions were made by the introduction of the Merrifield's solid phase peptide synthesis [8] and Carpino's protecting group methodologies [9]. Additionally further parameters like the micro-

wave-assistance [10, 11], the choice of the solvents [12], the manual and automated procedures as well as the selection of resins and linkers [13, 14] ameliorated the SPPS methodology concerning the yields, purity, and reaction times are comprehensively documented by Martinez [15].

All this enabled the use of PNAs useful in many ways, for instance as antisense molecules in pro- as well as in eukaryotic organisms [16, 17], and as triplex forming oligonucleotides (TFO) for antigenic strategies [18-20]. From this point of view the use as therapeutic and diagnostic agents was documented [21]. Further, PNAs have applications in analysis of biosensor chips for identification of nucleic acids [22]. The structural and the physico-chemical properties of PNAs as well as the different synthesis methodologies are well documented [23-30]. They reveal similarities to the native nucleic acids whose phospho-ribose backbone is substituted by a backbone of poly-2-aminoethyl glycine with nucleobases connected via an acetate linker [31-34].

The use of PNA molecules is not restricted to its role as a DNA derivative. Derivatizations permit further functionalizations of the PNA polyamide backbone as demonstrated [35, 36]. Finally the nucleobases were substituted with functional molecules as reaction partners for chemical reactions, for instance as a ligation partner in the "click-chemistry" [37-40]. Manifold ligation reactions like the copper catalyzed alkyne-azide cycloadditions introduced by Sharpless [41] are well reviewed by El-Sagheer and Brown [42]. Here for instance, we used as ligation technology the Diels Alder Reaction with inverse electron demand (DAR<sub>inv</sub>) introduced by Lindsay [43] and comprehensively investigated by Sauer [44] and Bertozzi [45, 46]. The potential of the DAR<sub>inv</sub> is well documented, undoubted and underlines multifaceted applications in the medical science. It is an attractive platform for active agents in therapy [47-50] as well as for imaging components in diagnostics [51, 52]. The combination of different drugs and different diagnostic molecules in optimized ratios [52] allows the design of molecules ready to use for bi- tri- and multi-modal strategies. These can be considered as promising molecules in the personalized medicine and in the increasing field of theranostics [53]. Our variant, a heat-assisted approach of the solid phase peptide synthesis is considered as an indispensable methodology which realizes a proper chemistry of PNA-based backbones functionalized multifold with a great potential to contribute to the diagnostics' precision and to the better therapy's success.

We tested the synthesized PNAs implemented into the drug delivery and targeting system called

BioShuttle [54] as functional modules for hybridization in the cells with target sequences in the cathepsin B (CtsB) mRNA reacting in case of the existence of the activated CtsB enzyme in the cells [55]. After enzymatic cleavage in the cytoplasm the fluorescent dye Rhod110 is transported and detectable in the cell nucleus.

## Chemical Procedures & Results

### PNA synthesis

For the PNA synthesis of the sequences (I) cagcgtgcag-C, (II) ctgcagcgtg-C, and (III) agcgtgagct-C (listed in the column D of Table 1) we used a manual synthesizer, PetiSyzer® from HiPep Laboratories, Kyoto, Japan. On this instrument we performed low cost simultaneous synthesis in disposable polypropylene reactors. An assembly of five disposable reactors is connected to a polytetrafluoroethylene tube (PTFE) manifold via PTFE two-way valves and are fixed on to a rack that can be shaken by a vortex-like mixer. The five reactors make contact with an aluminium block whose temperature is controlled by either an electric heater or cooled by the circulation of chilled liquid. In peptide synthesis it is known that elevated temperatures in the coupling reaction can overcome some coupling difficulties and shorten the coupling time. However it is also known that Fmoc-removal with piperidine at elevated temperatures results in racemization. Thus temperature control is required for such reactions. For the present PNA synthesis we intended to minimize aggregation and sterical hindrance by raising the temperature during the coupling time as it is known from peptide synthesis [56]. We used 9-fluorenylmethoxycarbonyl chloride (Fmoc)-building blocks with blocked side chains of Adenine A, Cytosine C, and Guanine G by benzhydroxycarbonyl (Bhoc) groups. The syntheses were performed in an 2 μmol scale on a H-Cys(Trt)-HMPB-ChemMatrix® resin loading 0.19 mmol/g. In the first step the Fmoc-group was cleaved with 20% piperidine in dimethylformamide (DMF) for 5 min at 20°C. After that, the resin was washed 5 times with DMF. For the coupling reaction the temperature was raised to 80°C and the coupling reaction was performed with 2-(1H-7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyl uronium hexafluorophosphate (HATU) and diisopropylethylamine (DIPEA) for 40 min. After the coupling all free amino groups were capped with acetic acid anhydride in DMF and the resin was washed with DMF. Before the next Fmoc-deprotection step the reaction vessel was cooled to 20°C and the resin was treated with 20% piperidine in DMF. At the end of the PNA synthesis, the last step

was a cleavage of the N-terminal protecting group with piperidine.

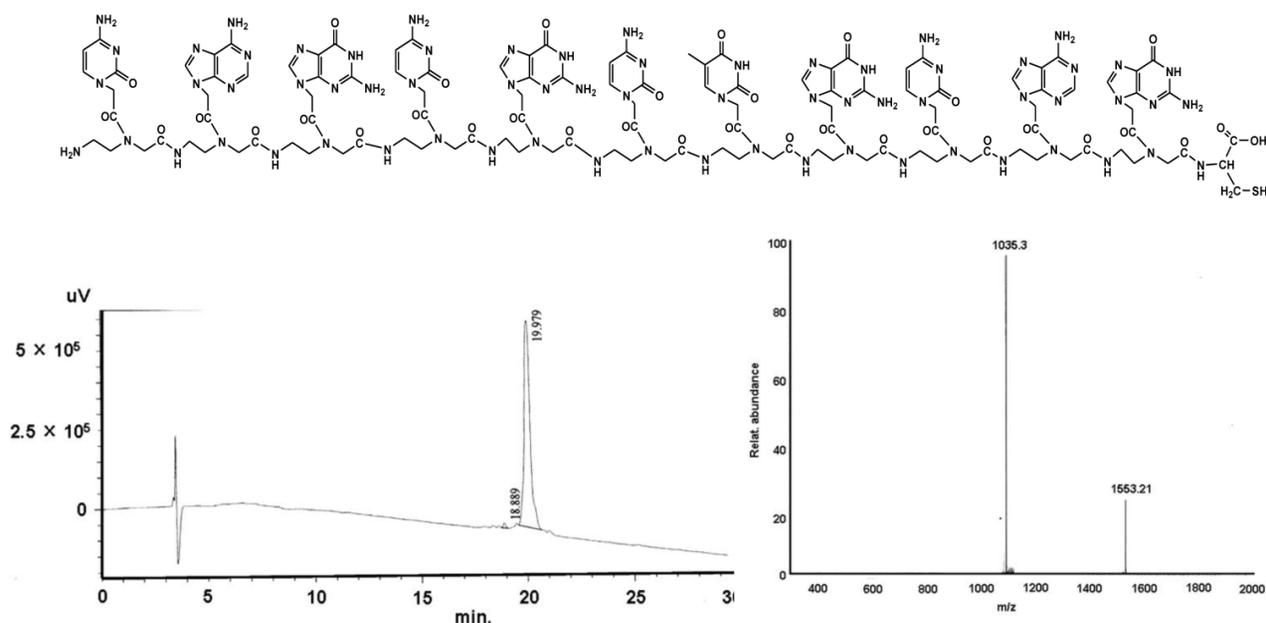
A small sample of each PNA was cleaved with TFA (90%) and scavenger triethylsilan/water (2.5%/2.5%) for 2.5 h at room temperature. The PNA was precipitated in ether and lyophilized. Analysis

was carried out with HPLC and MS. By using different temperatures (20°C for Fmoc-protection (avoid racemization) and 80°C for the coupling reaction we could improve coupling efficiency compared to use of elevated temperature for the whole synthesis.

**Table I.** The schematic structure of the CtsB-BioShuttle (upper part) accentuates the modular construction which consists of the following functional units: CPP module (red) is responsible for the passage across cell membranes (the amino acid sequence is shown in the column F). The PNA module (yellow) harbours the sequences for the hybridization (column D, line **1** – antisense; line **2** – sense; line **3** – random) with the target-sequence inside of the CtsB mRNA's Exon I. After the enzymatic cleavage ✂ (red) of the disulfide bridge (column E) inside of the cytosol, the CtsB cleavage module (column C, highlighted in grey) ✂ covalently coupled to the NLS module (column B, red/blue) is cut. This in turn is connected to the Rhod110 (column A) fluorescent dye as a cargo (green) is illustrated. In the lines **1-3** the BioShuttle conjugates used for experiments are described.

	A	B	C	D	E	F	G
<b>1</b>	Rhod110	VKRKKP	GFRK	cagcgctgcag-C-	S <sup>∩</sup> S	-C-RQIKIWFQNRRMKWKK	Calc. MW: 6006.21
<b>2</b>	Rhod110	VKRKKP	GFRK	ctgcagcgctg-C-	S <sup>∩</sup> S	-C-RQIKIWFQNRRMKWKK	Calc. MW: 5997.21
<b>3</b>	Rhod110	VKRKKP	GFRK	agcgctgagct-C-	S <sup>∩</sup> S	-C-RQIKIWFQNRRMKWKK	Calc. MW: 6021.31

### PNA I - cag cg c tg c ag-C



**Figure 1.** shows the structural formula of the PNA I (upper part). The lower part demonstrates the graphs of HPLC (left) and mass (right).

$C_{120}H_{151}N_{67}O_{34}S$ :

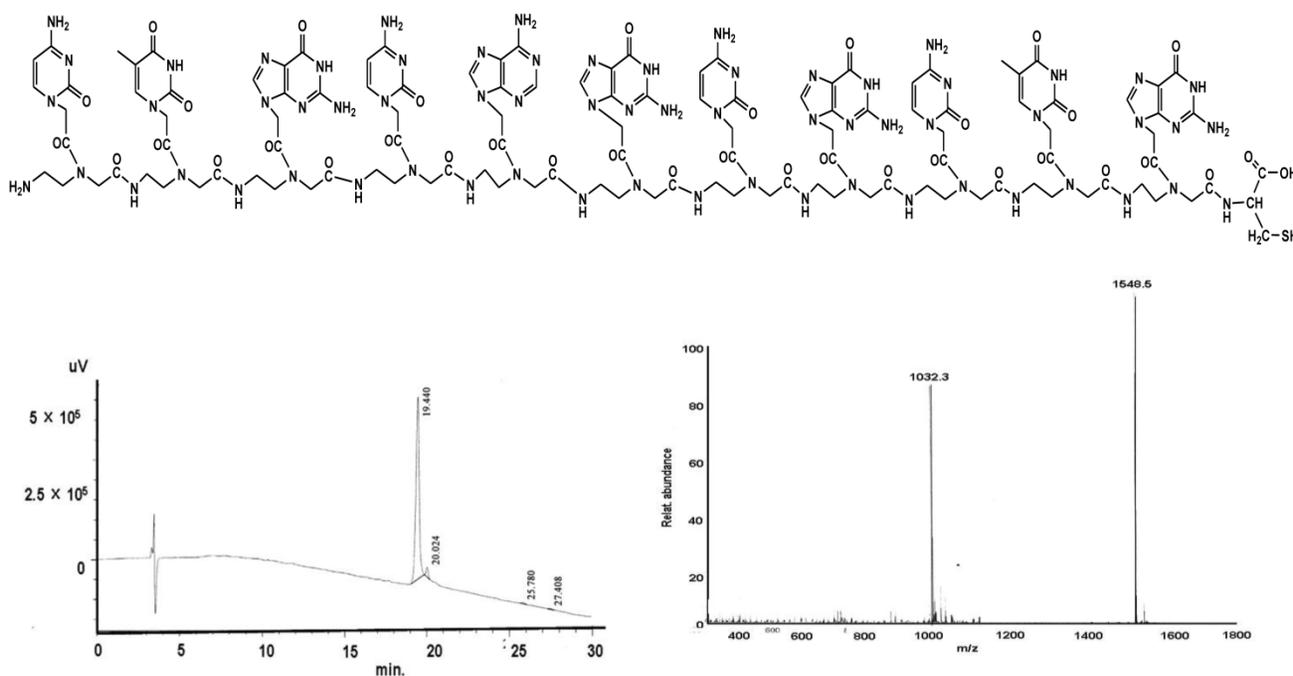
Found mass: 3106.2;  $(M-2H)^{2+}$  1553.21;  $(M+3H)^{3+}$  1035.3

Exact mass: 3106.19 (Mol. Wt.: 3107.98);

m/e: 3107.19 (100.0%), 3108.19 (94.4%), 3106.19 (74.8%), 3109.19 (29.6%), 3109.20 (28.7%), 3107.18 (18.5%), 3110.20 (14.5%), 3110.19 (14.0%), 3111.20 (6.9%), 3108.18 (5.8%), 3111.19 (4.6%), 3109.18 (4.0%), 3112.20 (2.3%), 3108.20 (1.7%), 3110.18 (1.5%), 3112.19 (1.1%)

C, 46.37; H, 4.90; N, 30.19; O, 17.50; S, 1.03

### PNA II - ctg cag cgc tg-C



**Figure 2.** shows the structural formula of the PNA II (upper part). The lower part demonstrates the graphs of HPLC (left) and mass (right).

$C_{120}H_{152}N_{64}O_{36}S$ :

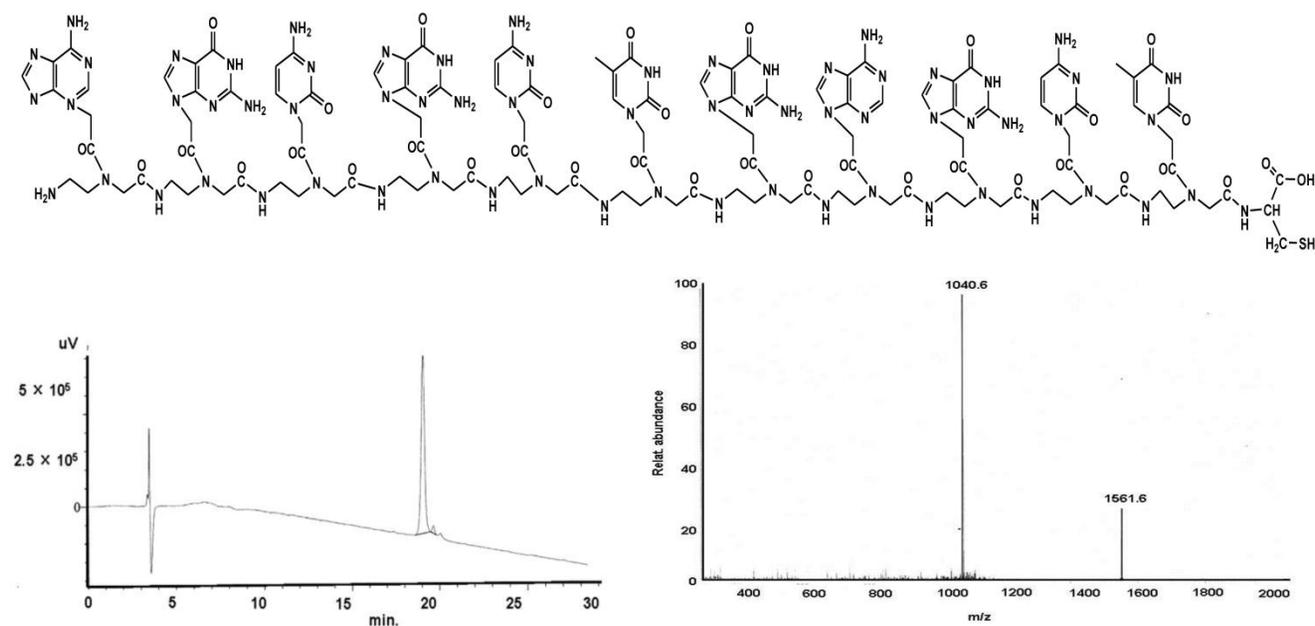
Found mass: 3097.0;  $(M-2H)^{2+}$  1548.5;  $(M+3H)^{3+}$  1032.3

Exact mass: 3097.18; (Mol. Wt.: 3098.96)

m/e: 3098.18 (100.0%), 3099.18 (95.9%), 3097.18 (75.2%), 3100.19 (28.8%), 3100.18 (24.9%), 3098.17 (18.4%), 3101.19 (14.5%), 3101.18 (13.9%), 3100.17 (8.3%), 3102.19 (7.1%), 3099.17 (5.6%), 3102.18 (3.9%), 3103.19 (1.9%), 3101.17 (1.6%), 3103.18 (1.5%), 3102.17 (1.0%)

C, 46.51; H, 4.94; N, 28.93; O, 18.59; S, 1.03

## PNA III – agc gct gag ct-C



**Figure 3.** shows the structural formula of the PNA III (upper part). The lower part demonstrates the graphs of HPLC (left) and mass (right).

$C_{121}H_{152}N_{66}O_{35}S$ ;

Found mass: 3123.2;  $(M-2H)^{2+}$  1561.6;  $(M+3H)^{3+}$  1040.6

Exact mass: 3121.19; Mol. Wt.: 3122.99

m/e: 3122.19 (100.0%), 3123.19 (94.7%), 3121.19 (74.2%), 3124.19 (29.7%), 3124.20 (29.1%), 3122.18 (18.1%), 3125.20 (14.7%), 3125.19 (14.0%), 3126.20 (7.1%), 3123.18 (5.7%), 3126.19 (4.6%), 3124.18 (3.9%), 3127.20 (2.4%), 3123.20 (1.7%), 3125.18 (1.6%), 3127.19 (1.1%)

C, 46.54; H, 4.91; N, 29.60; O, 17.93; S, 1.03

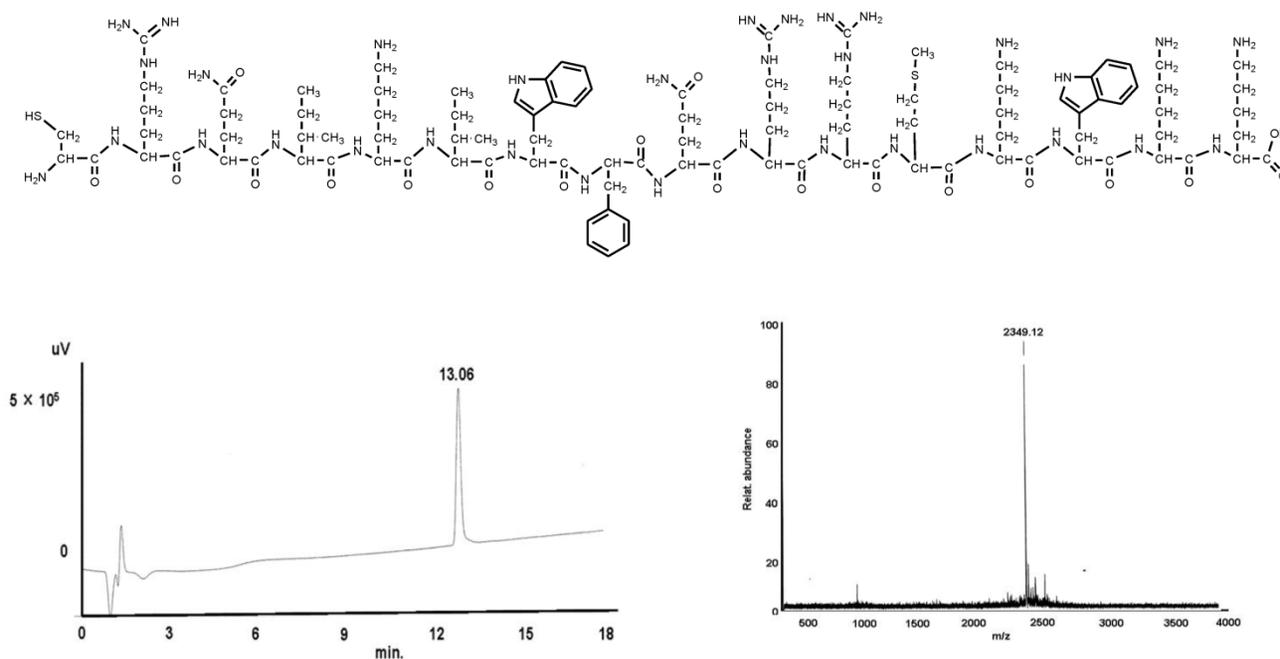
### Peptide synthesis

The peptides C-RQIKIWFQNRMMKWKK [pAnt43-58], VKRKKP-GFGRK-cagcgctgcag-C, VKRKKP-GFGRK-ctgcagcgctg-C, and VKRKKP-GFGRK-agcgcctgagct-C were synthesized in an automated multiple synthesizer Syro II (MultiSyn Tech, Germany) using Fmoc chemistry under atmospheric conditions at room temperature using the Fmoc-Lys(Boc)-HMPB-ChemMatrix® resin loading 0.50 mmol/g. For amino acid coupling a fivefold excess of the Fmoc-protected amino acid was activated *in situ* with 5 equivalents 2-(1H-Benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU) and DIPEA (0.5M) in DMF. The coupling time was 40 min. The Fmoc-group

was cleaved with piperidine (20%) in DMF for 3 min and 10 min. After each step the resin was washed 5 times with DMF.

The Rhodamine 110 marker was inserted into the peptide by a manual coupling procedure. A threefold excess of this marker was activated *in situ* with 3 equivalents HATU and 2 equivalents DIPEA and added to the resin bound peptide for one hour reaction time. Afterwards the resin was washed three times with DMF, dichloromethane (DCM) and isopropanol and dried. The following cleavage of the peptide from the resin and of the side chain protecting groups was performed with trifluoroacetic acid (TFA) (90%) and as scavenger triethylsilan/water (2.5%/2.5%) for 2.5 h at room temperature was used.

## Synthesis of the pAnt43-58 – cell penetrating peptide (CPP)



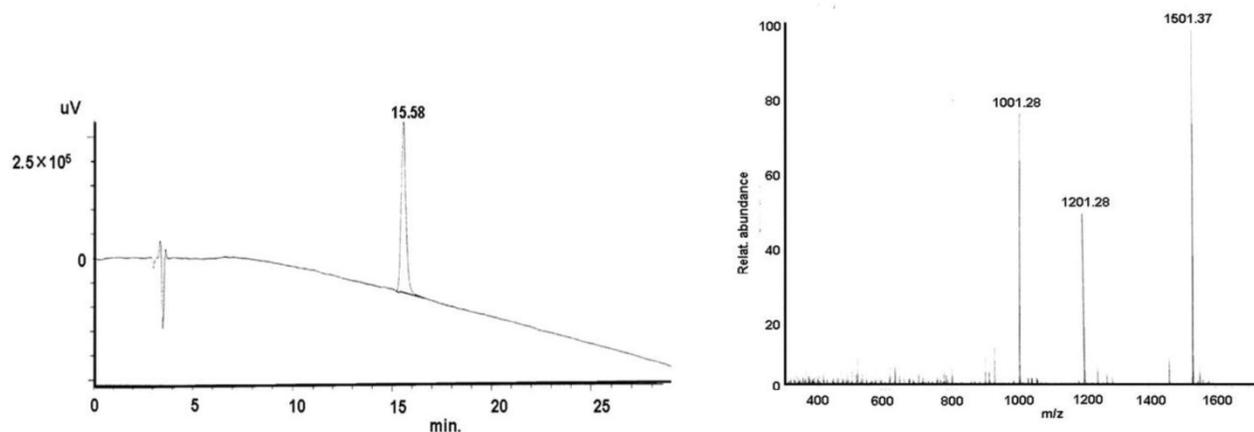
**Figure 4.** shows the structural formula of the pAnt43-58 (upper part). The lower part demonstrates the graphs of HPLC (left) and mass (right).

$C_{107}H_{174}N_{35}O_{21}S_2$

Found mass: 2349.305;

Exact Mass: 2349.3; Mol. Wt.: 2235.77

m/e: 2350.30 (100.0%), 2351.31 (77.8%), 2353.31 (43.9%), 2354.31 (19.8%), 2355.26 (2.4%), 2356.31 (0.7%), 2357.31 (0.18%), C, 55.33; H, 7.53; N, 20.67; O, 13.60; S, 2.87



**Figure 5.** shows exemplarily the HPLC and MS plots of the CtsB-BioShuttle construct **1** (shown in Table I) containing the antisense PNA I after synthesis.

Found mass:

Exact mass: MW Cal: 6006.21 1501.37 ( $M+4H$ )<sup>4+</sup>; 1201.39 ( $M+5H$ )<sup>5+</sup>; 1001.28 ( $M+6H$ )<sup>6+</sup>

### Purification of pAnt43-58 and Rhodamine-peptide-PNA conjugates

The crude material was purified by preparative HPLC on a Kromasil 100-10C 18  $\mu\text{m}$  reverse phase column (30' 250mm) using an eluent of 0.1% trifluoroacetic acid in water (A) and 80% acetonitrile in water (B). The peptide was eluted with a successive linear gradient of 10% B to 80% B in 30 min at a flow rate of 23 ml/min. The fractions containing the purified protein were lyophilized. The purified material was characterized with analytical HPLC and matrix assisted laser desorption mass spectrometry (MALDI-MS) (Figure 1 - 5Figure 1).

### Ligation via disulfide bridge formation

The disulfide bridge between the N-terminal cysteine of pAnt43-58 and the N-terminal cysteine residue of the NLS was achieved by the activation of one cysteine using 2,2'-dithiopyridine and the subsequent coupling to the cysteine of the other module. The coupling reaction was carried out in an ethanol / water buffer at 60°C.

The individual components and the complete modules were validated using LCMS (Shimadzu LC-10 and LCQ electrospray, Finnigan-Mat, (Thermo Fischer Scientific GmbH, Bremen, Germany). The purity levels were generally >90%.

After ligation the reaction products were tested in the following cell study; the ultimate conjugates are listed Table 1.

### Cell culture & CtsB-BioShuttle FFM-measurements

In order to characterize the CtsB-BioShuttle conjugates we used HeLa [57, 58] and MDA-MB-231 [59, 60], human cell lines which are characterized by their different *CtsB* expression and activity of the gene product. *CtsB* considered as a genetic marker is strongly expressed during proteolysis' and apoptosis' processes [61-63]. Therefore the *CtsB* mRNA was chosen as a target for a fluorescent imaging test of our CtsB-BioShuttle constructs.

In GenBank with the accession number M14221 the human cathepsin B mRNA sequence was searched. This sequence describes the total *CtsB* mRNA.

The bases sequences of the three investigated PNAs are represented as follows:

The **PNA I** is the corresponding antisense molecule and possesses the antiparallel complementary sequence of the *CtsB* mRNA Exon I at the position 70-80. As a control we synthesized the **PNA II** which is identical with the sequence of the *CtsB* mRNA at

the end of the Exon I position 70-80 [64]. The **PNA III**'s sequence is randomized.

The MDA-MB-231 cell line was cultivated in Dulbecco's modified Eagles Medium (DMEM) (Invitrogen Life Technologies, Carlsbad, USA) with 10% fetal calf serum and 1% Glutamine (Biochrom, Germany). The HeLa cells were cultivated in RPMI 1640 (GIBCO, Invitrogen) supplemented with 10% fetal Calf serum. All cell lines were cultivated without phenol red and maintained at 37°C in 5% CO<sub>2</sub> with high atmospheric humidity.

24 hours before fluorescence measurement studies the cells were washed in Hank's Balanced Salts (PAN-Biotech, Germany), after treatment with trypsin/EDTA solution (0.5/0.2 %) harvested and suspended in fresh medium. Cell suspension (400  $\mu\text{l}$ ) were transferred to chambers of the Lab-Tek™ Chamber Slides (Nunc, USA) and incubated under identical conditions as described above. For measurement studies the MDA-MB-231 cells were treated with the CtsB-BioShuttle conjugates **PNA I**, **PNA II**, and **PNA III** (final concentration 100 nM) for 1 h. Finally, the medium was removed and the cells were washed with Hank's and fresh medium was added. The HeLa cells were treated identically with the CtsB-BioShuttle **PNA I** variant (as a negative control). After 24 hours, the samples were measured as shown in Figure 6.

### Discussion and outlook

Progresses in both, the genome - and in the informatics research established proteomics and metabolomics and influenced the pharmaceutical research in diagnostic as well as in therapeutic areas [65-70]. "Old fashioned" diagnostic molecules and drugs are highly effective but not sensitive enough, hamper the therapeutic success and result in discontinuation of treatment. It is beyond controversy that these drugs, which represent the old paradigm "one size fits all", is not really up-to-date anymore [71, 72].

The Blockbusters' importance seems to approach perpetually its end, in parallel, the time for individual therapeutic interventions started [73, 74]. The situation changed and the new paradigm delineates the "patient specific medicine" [75]. This "personalized medicine" is based on the generation and application of the patient-specific drug [76, 77]. The development of such components is deemed to be a global challenge for "Big Pharma". In consideration of quality and specificity the biotech industry seems to have advances in providing breakthrough products more efficiently than pharmaceutical industrial manufacturing [78]. The recombinant chemistry together with the solid phase peptide synthesis chemistry (SPPS)

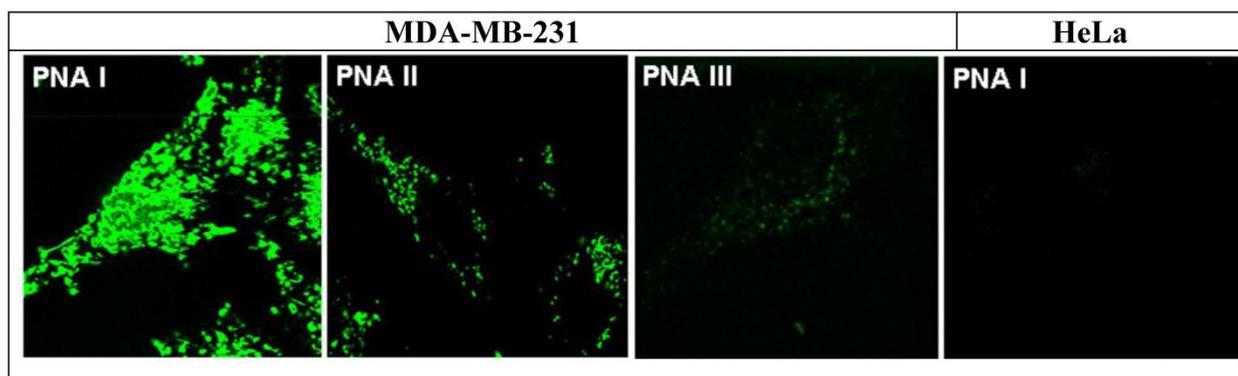
represent two methodologies to fulfil the demands of crucial criteria (high quality & high variability) for biotechnology products [79] and the synthesis of molecules designed for the patient's genetic disposition.

Here we discuss a SPPS variant featuring these properties which is based on the classical solid phase strategy but focuses on the temperature as physical parameter. However, folding processes after peptide syntheses can hamper the prolongation of the peptide bond formation during the solid phase synthesis.

Microwave heat assisted syntheses, however at elevated reaction temperature resulted in increased racemization [80, 81]. We circumvented this drawback using a concept based on a personal synthesizer, used for peptide synthesis and also for organic chemical syntheses that involves heating, cooling and filtration procedures [82]. A microwave methodology providing synthesis conditions avoiding racemization is still under investigation by the Nokihara group, HiPep Laboratories, Kyoto (unpublished data).

The temperature is assumed to be the critical parameter in limiting the peptide synthesis as documented in 1988 by Merrifield [83]. A heat-induced unfolding of peptide chains can be understood as an

initial point for the successful construction of large proteins, shown by the Dolphin group [56]. Here we studied this methodology in the solid phase synthesis of peptide nucleic acids as an initial step for the entry into "personalized medicine" in oncology using exemplarily the cathepsin B gene (*CtsB*) and the corresponding mRNA as molecular target. The SPPS, the attachment of a fluorescence dye and the following ligation of the modules by disulfide formation result in the final CtsB-BioShuttle conjugate designed for fluorescence imaging in the selected cell lines. HeLa cells were used as a control and the imaging of the CtsB mRNA was documented [55]. Here we confirmed the data of the non-invasive HeLa cervix carcinoma and the invasive MDA-MB-231 the breast cancer cell lines differentially expressing the CtsB (Figure 6). The CtsB-BioShuttle on MDA-MB-231 resulted in strong signal using the construct with the complementary PNA I. This would be the direct way to detect tumor cells and treat with patient specific PNA at the same time. Insistently, the complexity of the pharmaceutical research and in biotechnology, the dedicated chemical methodologies like the SPPS hold a key role in the development of patient-specific drugs and imaging agents.



**Figure 6.** The figure shows FFM-measurement pictures of the cell lines: MDA-MB-231 cells 24 hours after treatment (final concentration 100 nM) with the CtsB-BioShuttle conjugates PNA I, PNA II and PNA III, and HeLa control cells (right picture) treated with the PNA I (antisense). The fluorescence signals can be observed inside of both lines and indicate an uptake of all tested CtsB-BioShuttle conjugates. Whereas the fluorescence signal intensity inside of the cytoplasm of the MDA-MB-231 cells treated with PNA III (random) is hardly visible related to the signal intensities inside of the cytosol of MDA-MB-231 cells which are treated with PNA I and PNA II. Solely in the nuclei of the MDA-MB-231 cells treated with the PNA I clear morphologic structures with high fluorescence intensities are recognizable. The HeLa control cells treated with the PNA I (antisense) did not show fluorescence signals.

## Conflict of Interest

The authors have declared that no conflict of interest exists.

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